

# The $\text{Ca}_v\beta$ Subunit Protects the I-II Loop of the Voltage-gated Calcium Channel $\text{Ca}_v2.2$ from Proteasomal Degradation but Not Oligoubiquitination\*

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$\text{Ca}_v\beta$  subunits interact with the voltage-gated calcium channel  $\text{Ca}_v2.2$  on a site in the intracellular loop between domains I and II (the I-II loop). This interaction influences the biophysical properties of the channel and leads to an increase in its trafficking to the plasma membrane. We have shown previously that a mutant  $\text{Ca}_v2.2$  channel that is unable to bind  $\text{Ca}_v\beta$  subunits ( $\text{Ca}_v2.2$  W391A) was rapidly degraded (Waithe, D., Ferron, L., Page, K. M., Chaggar, K., and Dolphin, A. C. (2011) *J. Biol. Chem.* 286, 9598–9611). Here we show that, in the absence of  $\text{Ca}_v\beta$  subunits, a construct consisting of the I-II loop of  $\text{Ca}_v2.2$  was directly ubiquitinated and degraded by the proteasome system. Ubiquitination could be prevented by mutation of all 12 lysine residues in the I-II loop to arginines. Including a palmitoylation motif at the N terminus of  $\text{Ca}_v2.2$  I-II loop was insufficient to target it to the plasma membrane in the absence of  $\text{Ca}_v\beta$  subunits even when proteasomal degradation was inhibited with MG132 or ubiquitination was prevented by the lysine-to-arginine mutations. In the presence of  $\text{Ca}_v\beta$  subunit, the palmitoylated  $\text{Ca}_v2.2$  I-II loop was protected from degradation, although oligoubiquitination could still occur, and was efficiently trafficked to the plasma membrane. We propose that targeting to the plasma membrane requires a conformational change in the I-II loop that is induced by binding of the  $\text{Ca}_v\beta$  subunit.

Voltage-gated calcium channels have a crucial role in excitable cells, being involved in neurotransmitter release, gene transcription, muscle contraction, and calcium-induced calcium release from internal stores (2). High voltage-activated calcium channels are heteromultimeric complexes made of a pore-forming  $\text{Ca}_v\alpha1^3$  subunit (3) along with auxiliary  $\text{Ca}_v\beta$  and  $\text{Ca}_v\alpha2\delta$  subunits (4).

$\text{Ca}_v\beta$  subunits have two distinct effects on the expression on  $\text{Ca}_v\alpha1$  subunits: first, they influence the biophysical properties of the channels by hyperpolarizing the voltage dependence of activa-

tion and increasing the probability of opening (5), and second, they increase the trafficking of the channels to the plasma membrane (6). The binding site for  $\text{Ca}_v\beta$  subunits on  $\text{Ca}_v\alpha1$  was shown to be a highly conserved region of the intracellular loop between domains I and II (I-II loop) of high voltage-activated channels that became known as the  $\alpha$ -interaction domain (AID) (7). This conserved motif, consisting of 18 amino acids starting 24 residues after the end of the IS6 transmembrane segment of  $\text{Ca}_v2.1$ , was found to bind to all four  $\text{Ca}_v\beta$  subunits (8).

The mechanism by which the  $\text{Ca}_v\beta$  subunit promotes trafficking of the  $\text{Ca}_v\alpha1$  to the plasma membrane is under debate. Apart from  $\text{Ca}_v\beta2$ , which can associate with the plasma membrane either through palmitoylation (9) or by a polybasic region in the case of  $\text{Ca}_v\beta2e$  (10), the  $\text{Ca}_v\beta$  subunits are cytoplasmic proteins. It was first suggested that the  $\text{Ca}_v\alpha1$  I-II loop contained an ER retention motif that was masked in the presence of a  $\text{Ca}_v\beta$  (11), but more recently it has been postulated that the I-II loops contain an acidic ER export signal (12). Fusion proteins containing the  $\text{Ca}_v\alpha1$  I-II loops fused to a transmembrane CD4 sequence were not retained in the ER (13), and evidence suggests that binding of the  $\text{Ca}_v\beta$  subunits to the  $\text{Ca}_v1.2$  and  $\text{Ca}_v2.2$  I-II loop prevented ubiquitination and degradation of these channels (1, 13). Our previous work has shown that a mutant  $\text{Ca}_v2.2$  channel that is unable to bind  $\text{Ca}_v\beta$  subunits ( $\text{Ca}_v2.2$  W391A) was subjected to increased proteasomal degradation relative to the wild-type channel (1).

Efficient degradation by the proteasome requires the substrate to be ubiquitinated. Ubiquitin is a protein of 76 amino acids that is attached by a covalent bond to a lysine residue in the substrate; this then serves as a sorting signal (14). A single ubiquitin molecule can bind, resulting in monoubiquitination. More commonly, as ubiquitin contains seven lysine residues and can itself be ubiquitinated, polyubiquitination occurs. Different types of ubiquitination lead to different fates of the proteins; for example, Lys-48-linked ubiquitination is a signal to target the substrate for degradation via the proteasome (14).

In this study, we show that, in the absence of  $\text{Ca}_v\beta$  subunit, the isolated I-II loop of  $\text{Ca}_v2.2$  is directly ubiquitinated on a lysine residue within the loop and is rapidly degraded. Binding of a  $\text{Ca}_v\beta$  subunit prevents degradation of the ubiquitinated I-II loop but does not prevent oligoubiquitination. Surprisingly, we found that attaching a palmitoylation motif to the N terminus of the I-II loop is insufficient to target it to the plasma membrane in the absence of the  $\text{Ca}_v\beta$  even when protein degradation is inhibited. This suggests that membrane targeting

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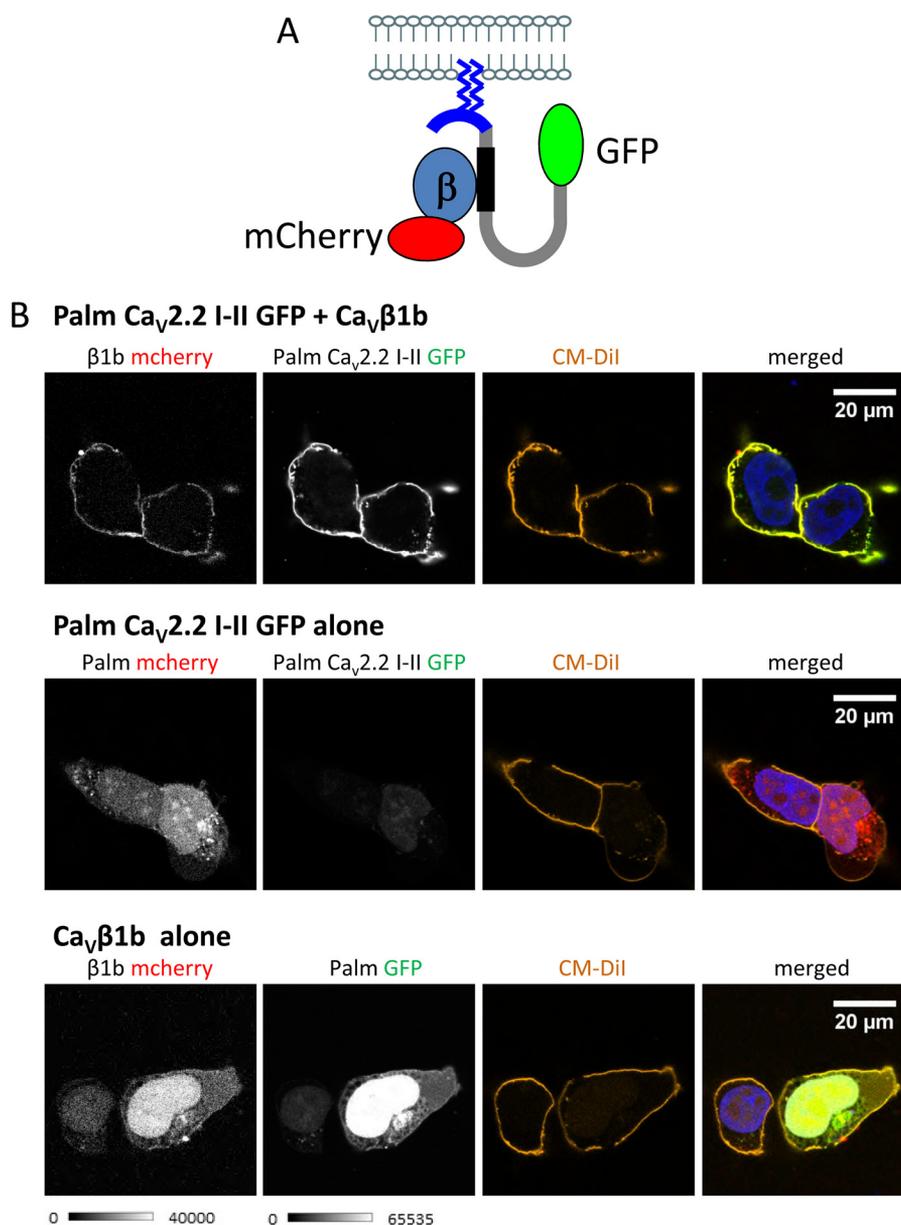
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<sup>3</sup> The abbreviations used are:  $\text{Ca}_v$ , voltage-gated calcium; AID,  $\alpha$ -interaction domain; Ub, ubiquitin; palm, palmitoylated; Bis-Tris, 2-[bis(2-hydroxyethyl)-amino]-2-(hydroxymethyl)propane-1,3-diol; pF, picofarad; nS, nanosiemens; ER, endoplasmic reticulum; WCL, whole cell lysate.

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**FIGURE 1. Palm  $Ca_v2.2$  I-II-GFP is trafficked to the plasma membrane in the presence of  $Ca_v\beta1b$ .** *A*, schematic showing the constructs used. The palmitoylation motif, MTLESIMACCL, containing two palmitoylated cysteines and shown in blue, was fused to the N terminus of the I-II loop of  $Ca_v2.2$  (amino acids 356–483). GFP was fused to the C terminus of the I-II loop.  $Ca_v\beta1b$ , which is fused to mCherry, interacts with the  $\beta$ -binding site on the I-II loop, shown in black. *B*, confocal images of tsA-201 cells co-expressing palm  $Ca_v2.2$  I-II-GFP (shown in grayscale; top row, second panel) with  $Ca_v\beta1b$ -mCherry (top left) or palm mCherry (in the absence of  $Ca_v\beta$ ; middle row). The bottom row shows  $Ca_v\beta1b$ -mCherry (left) co-expressed with palm GFP (second panel). For each condition, the third column shows the cells stained with the plasma membrane marker CM-Dil. Palm  $Ca_v2.2$  I-II-GFP and  $Ca_v\beta1b$ -mCherry are only trafficked to the plasma membrane when expressed together. The merged images on the right show mCherry in red, GFP in green, the nuclei stained with DAPI (blue), and the plasma membrane stained with CM-Dil (orange). The scale bar is 20  $\mu$ m, and the grayscale calibration bar is shown below.

requires the I-II loop to be correctly folded in the presence of the  $Ca_v\beta$  subunit.

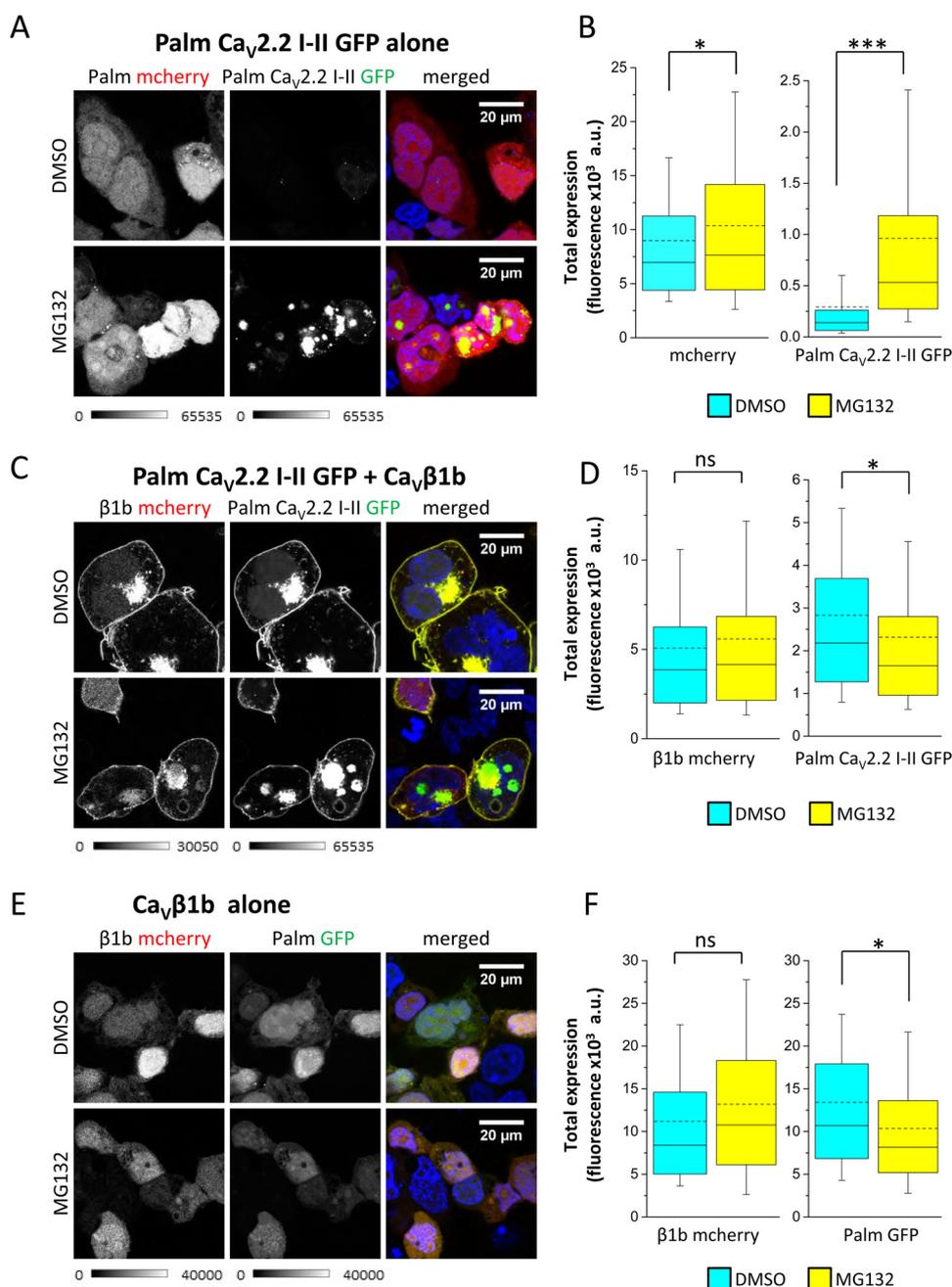
### Results

**Trafficking of the Palmitoylated I-II Loop of  $Ca_v2.2$  in tsA-201 Cells**—To investigate the mechanism by which the  $Ca_v\beta$  subunit protects  $Ca_v2.2$  from degradation and is involved in trafficking it to the plasma membrane, a construct was made containing the intracellular I-II loop of  $Ca_v2.2$  (amino acids 356–483), which is known to interact with the  $Ca_v\beta$  via the AID (7). A palmitoylation motif, MTLESIMACCL, the first 11 amino acids of the guanine nucleotide-binding protein  $G_q$  (15),

was added to its N terminus (palm  $Ca_v2.2$  I-II) to target it to the plasma membrane, and a tag, either GFP or HA, was added to the C terminus (Fig. 1A).

The expression and localization of the palmitoylated I-II loop of  $Ca_v2.2$  tagged with GFP (palm  $Ca_v2.2$  I-II-GFP) was investigated using confocal microscopy. In the presence of  $Ca_v\beta1b$  tagged with mCherry, palm  $Ca_v2.2$  I-II-GFP was efficiently trafficked to the plasma membrane as shown by its colocalization with the plasma membrane marker CM-Dil (Fig. 1B, top panel). In the absence of  $Ca_v\beta$  subunits, palm  $Ca_v2.2$  I-II-GFP showed very low expression levels in tsA-201 cells and was not found at the plasma membrane (Fig. 1B, middle panel)

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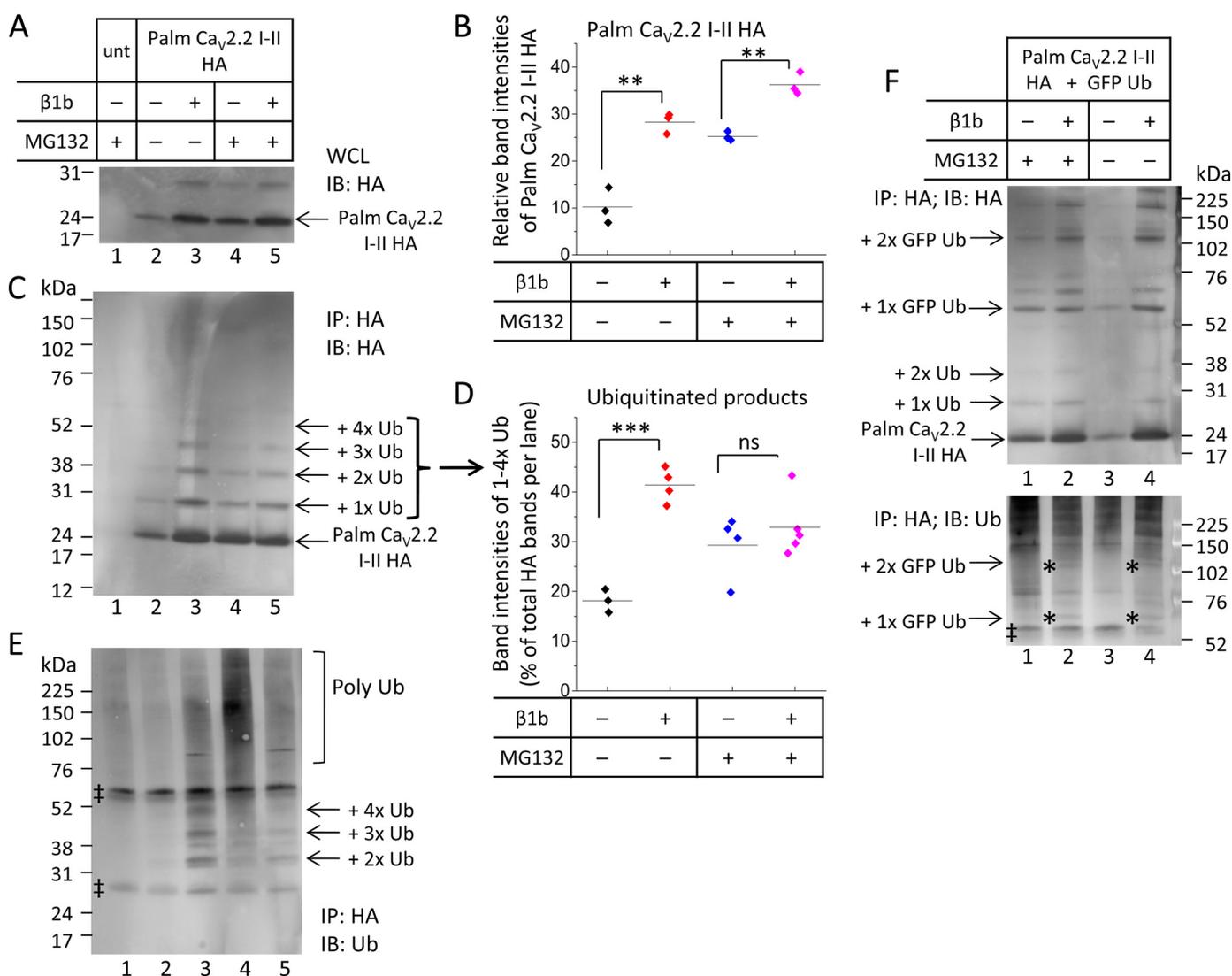


**FIGURE 2. In the absence of Ca<sub>v</sub>β subunit, palm Ca<sub>v</sub>2.2 I-II-GFP is rapidly degraded.** *A*, representative confocal images of tsA-201 cells co-expressing palm mCherry without Ca<sub>v</sub>β (shown in grayscale; left) and palm Ca<sub>v</sub>2.2 I-II-GFP (grayscale; middle) in the absence (top panels) and presence (bottom panels) of MG132 (4 μM). The panel on the right shows the merged image; DAPI was used to visualize the nucleus (blue). The scale bar is 20 μm, and the grayscale calibration bar is shown below each set of images. *B*, box and whisker plots showing fluorescence intensity in the absence of MG132 (DMSO control; cyan bar; *n* = 221 cells) or the presence of MG132 (yellow bar; *n* = 314) of the cells represented in *A*. The box shows the interquartile range, whiskers range from 10 to 90%, the median is shown as a solid line, and the mean is shown as a dotted line. GFP fluorescence was measured in all cells that were expressing mCherry. Palm mCherry fluorescence (left) was slightly increased in the presence of MG132 (\*, *p* = 0.04, Student's *t* test; mean fluorescence ± S.E. of 8977 ± 463 arbitrary units (a.u.) in DMSO and 10,379 ± 460 arbitrary units in MG132), whereas palm Ca<sub>v</sub>2.2 I-II-GFP (right) fluorescence was significantly increased by MG132 (\*\*\*, *p* < 0.0001, Student's *t* test; mean ± S.E. of 291 ± 40 (DMSO) and 962 ± 63 arbitrary units (MG132)). *C*, as for *A* but for cells co-expressing palm Ca<sub>v</sub>2.2 I-II-GFP with Ca<sub>v</sub>β1b-mCherry. *D*, box and whisker plots (as in *B*) showing fluorescence intensity of the cells in *C* in the absence (cyan; *n* = 243 cells) or the presence (yellow; *n* = 244) of MG132. Ca<sub>v</sub>β1b-mCherry (left) fluorescence was not significantly altered by MG132 (mean ± S.E. of 5067 ± 264 (DMSO) and 5595 ± 329 arbitrary units (MG132)); palm Ca<sub>v</sub>2.2 I-II-GFP (right) fluorescence was slightly (\*, *p* = 0.02, Student's *t* test) decreased in the presence of MG132 (mean ± S.E. of 2830 ± 145 (DMSO) and 2317 ± 156 arbitrary units (MG132)). *E*, control experiments showing the co-expression of Ca<sub>v</sub>β1b-mCherry and palm GFP as for *A*. *F*, box and whisker plots (as in *B*) showing fluorescence intensity of the cells in *E*; Ca<sub>v</sub>β1b-mCherry (left) or palm GFP (right) in the absence (cyan; *n* = 86 cells) or presence (yellow; *n* = 69) of MG132. Ca<sub>v</sub>β1b-mCherry fluorescence was not significantly different (mean ± S.E. of 11,200 ± 940 (DMSO) and 13,190 ± 1160 arbitrary units (MG132)); palm GFP fluorescence was slightly (\*, *p* = 0.03, Student's *t* test) decreased in the presence of MG132 (mean ± S.E. of 13,410 ± 1019 (DMSO) and 10,340 ± 871 arbitrary units (MG132)). ns, non-significant.

despite the presence of the palmitoylation sequence. Either the palm Ca<sub>v</sub>2.2 I-II-GFP was being expressed at much lower levels in the absence of Ca<sub>v</sub>β1b, or more likely it was being degraded.

When expressed alone, Ca<sub>v</sub>β1b-mCherry was found uniformly distributed throughout the cytoplasm (Fig. 1*B*, bottom left panel), but when co-expressed with palm Ca<sub>v</sub>2.2 I-II-GFP,

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**FIGURE 3. Palm Ca<sub>v</sub>2.2 I-II-HA is directly ubiquitinated.** *A*, Western blot showing aliquots (30 μg of protein) of WCLs from tsA-201 cells expressing palm Ca<sub>v</sub>2.2 I-II-HA and blotted with rat anti-HA antibody. *Lane 1* shows untransfected (*unt*) cells in the presence of the proteasomal inhibitor MG132 (4 μM). *Lanes 2–5* show cells transfected with palm Ca<sub>v</sub>2.2 I-II-HA in the absence (*lanes 2 and 3*) or presence (*lanes 4 and 5*) of MG132 and the absence (*lanes 2 and 4*) or presence (*lanes 3 and 5*) of Ca<sub>v</sub>β1b. *B*, quantification of band intensities expressed as a scatter plot for palm Ca<sub>v</sub>2.2 I-II-HA from Western blots of WCLs, including the example shown in *A*. Analysis was performed using ImageJ on gels from three separate transfections. The intensity of each band of palm Ca<sub>v</sub>2.2 I-II-HA was measured and expressed as a percentage of total palm Ca<sub>v</sub>2.2 I-II-HA band intensities on each gel. Mean band intensities (shown by *solid lines*) show that, in the absence of MG132 (*A, lanes 2 and 3*), Ca<sub>v</sub>β1b causes a significant increase in the amount of palm Ca<sub>v</sub>2.2 I-II-HA (10.22 ± 2.20, *black symbols* in the absence of Ca<sub>v</sub>β; 28.28 ± 1.27, *red symbols* in the presence of Ca<sub>v</sub>β1b; \*\*, *p* = 0.002, Student's *t* test). In the presence of MG132 (*A, lanes 4 and 5*), Ca<sub>v</sub>β1b also causes a significant increase in the amount of palm Ca<sub>v</sub>2.2 I-II-HA (25.23 ± 0.565, *blue symbols* in the absence of Ca<sub>v</sub>β; 36.27 ± 1.38, *pink symbols* in the presence of Ca<sub>v</sub>β1b; \*\*, *p* = 0.002, Student's *t* test). *C*, Western blot showing tsA-201 cells immunoprecipitated (*IP*) with rabbit anti-HA antibody and immunoblotted (*IB*) with rat anti-HA antibody. *Lanes* are the same as those in *A*. A ladder of ubiquitinated I-II loop proteins (indicated by *arrows*) is detected. *D*, quantification of HA-stained Western blots showing the I-II loop bound to one to four ubiquitins were combined and expressed as a percentage of total HA proteins per lane. Mean values are represented by *solid lines*. In the presence of Ca<sub>v</sub>β1b, ubiquitinated products make up a larger percentage of HA-detected proteins (mean ± S.E. of 41.38 ± 1.71%, *red symbols*, corresponding to *lane 3* of *C* compared with 18.12 ± 1.34% in the absence of Ca<sub>v</sub>β1b, *black symbols*, corresponding to *lane 2*; \*\*\*, *p* = 0.0002, Student's *t* test). In the presence of MG132 (*lanes 4 and 5*), Ca<sub>v</sub>β1b has no significant effect on the percentage of ubiquitinated products detected with the HA antibody (29.28 ± 3.23% in the absence of Ca<sub>v</sub>β1b, *blue symbols* compared with 32.88 ± 2.73% in the presence of Ca<sub>v</sub>β1b, *pink symbols*). *E*, Western blot showing tsA-201 cells immunoprecipitated with rabbit anti-HA antibody and immunoblotted with anti-Ub antibody for the same samples as those shown in *A* and *C*. The ladder of ubiquitinated products detected with the anti-HA antibody (shown in *C*) is also detected with anti-Ub (indicated with *arrows*) along with a smear of polyubiquitinated proteins of high molecular mass. Nonspecific IgG bands detected in all lanes are marked with ‡. *F*, Western blots showing tsA-201 cells transfected with palm Ca<sub>v</sub>2.2 I-II-HA and GFP-Ub, immunoprecipitated with rabbit anti-HA antibody, and immunoblotted with anti-HA (*top*) or anti-Ub (*bottom*) antibodies in the presence (*lanes 1 and 2*) and absence (*lane 3 and 4*) of MG132 and in the absence (*lanes 1 and 3*) and presence (*lanes 2 and 4*) of Ca<sub>v</sub>β1b. GFP-Ub competes with endogenous ubiquitin to bind to palm Ca<sub>v</sub>2.2 I-II-HA to give larger products (*arrows* for +1 × or +2 × GFP-Ub; also marked with *asterisks* in *bottom* blot). ‡ shows nonspecific IgG bands. *ns*, non-significant.

it too was found mainly at the plasma membrane (Fig. 1*B*, *top left panel*). Surprisingly, the palmitoylation motif alone was also insufficient to target mCherry (Fig. 1*B*, *middle row*) or GFP (Fig. 1*B*, *bottom row*) to the plasma membrane; trafficking to the

plasma membrane required both palm Ca<sub>v</sub>2.2 I-II and the Ca<sub>v</sub>β subunit (Fig. 1*B*).

Whether protein degradation was responsible for reduced palm Ca<sub>v</sub>2.2 I-II-GFP expression in the absence of Ca<sub>v</sub>β was

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investigated by measuring the levels of fluorescence in the absence and presence of the proteasomal inhibitor MG132 (Fig. 2, *A* and *B*). In the presence of MG132, the levels of palm Ca<sub>v</sub>2.2 I-II-GFP were significantly increased (Fig. 2, *A* and *B*), suggesting that it is usually rapidly degraded by the proteasome. However, it was still not associated with the plasma membrane.

In the presence of Ca<sub>v</sub>β1b tagged with mCherry, palm Ca<sub>v</sub>2.2 I-II-GFP was expressed at increased levels and was trafficked to the plasma membrane (Fig. 2*C*, *middle panel*). There was no increase in fluorescence in the presence of MG132 (Fig. 2*D*), suggesting that Ca<sub>v</sub>β protects palm Ca<sub>v</sub>2.2 I-II-GFP from being degraded. The same result was found using untagged Ca<sub>v</sub>β1b (data not shown). Control experiments showed that expression of either Ca<sub>v</sub>β1b-mCherry or palm GFP alone was not increased by MG132 (Fig. 2, *E* and *F*).

**Evidence for Multiple Stages of Ubiquitination of the Ca<sub>v</sub>2.2 I-II Loop**—To investigate the ubiquitination of the Ca<sub>v</sub>2.2 I-II loop in more detail, palmitoylated I-II loop constructs were tagged with hemagglutinin (HA) at the C terminus (palm Ca<sub>v</sub>2.2 I-II-HA) and expressed in tsA-201 cells in the presence or absence of Ca<sub>v</sub>β1b as well as in the presence or absence of the proteasomal inhibitor MG132. The anti-HA antibody identified palm Ca<sub>v</sub>2.2 I-II-HA from whole cell lysates (Fig. 3*A*). Quantification showed that there was significantly less palm Ca<sub>v</sub>2.2 I-II-HA in the absence of Ca<sub>v</sub>β than in the presence of Ca<sub>v</sub>β1b (Fig. 3*B*) in both the absence of MG132 (Fig. 3*B*, compare *columns 1* and *2*, which correspond to Fig. 3*A*, *lanes 2* and *3*) and the presence of MG132 (Fig. 3*B*, *columns 3* and *4*). Ca<sub>v</sub>β, therefore, is likely to protect palm Ca<sub>v</sub>2.2 I-II-HA from being degraded, although an effect on expression cannot be ruled out.

Palmitoylated Ca<sub>v</sub>2.2 I-II-HA proteins were immunoprecipitated from lysates with anti-HA antibody, and Western blotting analysis with both anti-HA and anti-ubiquitin antibodies was carried out. In this experiment, the protein A-Sepharose beads used for the pulldown were limiting; an excess of lysates was added in an attempt to immunoprecipitate equal amounts of palm Ca<sub>v</sub>2.2 I-II loop in the different conditions. Even so, there was less HA-tagged protein present in the absence of both Ca<sub>v</sub>β and MG132 (Fig. 3*C*, *lane 2* compared with *lanes 3–5*).

In the presence of Ca<sub>v</sub>β1b, palm Ca<sub>v</sub>2.2 I-II-HA was found at increased levels (Fig. 3*C*, *lane 3*). The anti-HA antibody identified the palm Ca<sub>v</sub>2.2 I-II-HA at the correct size (18 kDa) but also revealed a ladder of bands at higher molecular mass (Fig. 3*C*). Ubiquitin has a molecular mass of ~8 kDa. The ladder of bands corresponds to the size expected if one or more endogenous ubiquitins were bound to the Ca<sub>v</sub>2.2 I-II loop. Quantification of band intensity for ubiquitinated I-II loops expressed as a percentage of total protein per lane for HA-stained Western blots showed that the ubiquitinated palm Ca<sub>v</sub>2.2 I-II-HA represented a significantly lower proportion of total I-II loop protein in the absence of Ca<sub>v</sub>β than in the presence of Ca<sub>v</sub>β1b (Fig. 3*D*, *columns 1* and *2*). This difference was lost in the presence of MG132 (Fig. 3*D*, *columns 3* and *4*), suggesting that, in the absence of both Ca<sub>v</sub>β and MG132, ubiquitinated products are rapidly degraded. However, Ca<sub>v</sub>β1b does not appear to prevent the addition of one to four ubiquitins to the I-II loop (Fig. 3*C*, *lanes 3* and *5*).

The ladder of ubiquitinated I-II loop products was also observed using the anti-ubiquitin antibody (Fig. 3*E*), further evidence that these bands represent ubiquitinated I-II loop. The addition of one to four ubiquitins to palm Ca<sub>v</sub>2.2 I-II-HA was more evident in the presence of Ca<sub>v</sub>β1b (Fig. 3, *C* and *E*, *lanes 3* and *5*) than in its absence (*lanes 2* and *4*). It appears that Ca<sub>v</sub>β1b protects against degradation of these oligoubiquitinated products as well as or better than MG132.

There was no evidence of high molecular mass polyubiquitination in the absence of MG132 and Ca<sub>v</sub>β (Fig. 3*E*, *lane 2*), suggesting that, in the absence of the proteasomal inhibitor, the polyubiquitinated Ca<sub>v</sub>2.2 I-II loop was rapidly degraded by the proteasome. In contrast, in the presence of MG132 but the absence of Ca<sub>v</sub>β, palmitoylated Ca<sub>v</sub>2.2 I-II loop degradation was prevented, and there was an accumulation of high molecular mass polyubiquitinated proteins as seen by the smear above 76 kDa observed with the anti-ubiquitin antibody (Fig. 3*E*, *lane 4*). This smear was reduced in the presence of Ca<sub>v</sub>β1b (Fig. 3*E*, *lane 5*), suggesting that Ca<sub>v</sub>β1b protects the Ca<sub>v</sub>2.2 I-II loop from polyubiquitination as well as degradation. The Ca<sub>v</sub>β1b, therefore, appears to allow the addition of one to four ubiquitins to the Ca<sub>v</sub>2.2 I-II but prevents the polyubiquitination that usually leads to degradation.

Further evidence that the ladder of bands shown in Fig. 3, *C* and *E*, represents mono- or oligoubiquitinated products was obtained by including GFP-tagged ubiquitin in the transfection mixture. This competed with endogenous ubiquitin, reducing the density of bands at lower molecular mass (26 and 34 kDa) but producing bands at higher molecular mass (55 and 92 kDa) corresponding to the binding of one or two larger GFP-tagged ubiquitins (Fig. 3*F*). Addition of GFP-tagged ubiquitin was observed both in the presence and absence of Ca<sub>v</sub>β1b (Fig. 3*F*). The greater intensity of bands representing GFP-ubiquitin bound to I-II loop in the presence of Ca<sub>v</sub>β1b (Fig. 3*F*, *lane 4* compared with *lane 3*) suggests that there is less degradation when Ca<sub>v</sub>β1b is present. The smear of polyubiquitin-conjugated proteins visualized by the anti-ubiquitin antibody was much stronger in the presence of the highly expressed GFP-ubiquitin (Fig. 3*F*, *bottom panel*, compared with Fig. 3*E*). The anti-ubiquitin antibody binds to polyubiquitinated proteins much more efficiently than to monoubiquitinated conjugates (16).

**Ubiquitination Occurs on Lysine Residues within the Ca<sub>v</sub>2.2 I-II Loop**—Ubiquitination involves the formation of a covalent bond between the C terminus of ubiquitin and the ε-amino of a lysine residue on the substrate (17). The palmitoylated Ca<sub>v</sub>2.2 I-II-HA construct contains 12 lysines (Fig. 4*A*, \*). To determine whether ubiquitination occurs directly on the I-II loop, all 12 of the lysine residues were mutated to arginines (palm Ca<sub>v</sub>2.2 I-II K-R), and confocal and co-immunoprecipitation experiments were performed on the mutated constructs expressed in tsA-201 cells in the presence or absence of Ca<sub>v</sub>β1b subunits.

Confocal imaging experiments showed that, when expressed alone in tsA-201 cells, the palmitoylated Ca<sub>v</sub>2.2 I-II K-R mutant tagged with GFP (palm Ca<sub>v</sub>2.2 I-II K-R-GFP) was either expressed at higher levels or was less degraded than the wild-type I-II loop (Fig. 4*B* compared with wild type in Fig. 2*A*). However, it was not trafficked to the plasma membrane in the



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Ca<sub>v</sub>β1b-mCherry, the difference in GFP fluorescence intensities was much reduced (Fig. 4G).

Western blotting of the palmitoylated Ca<sub>v</sub>2.2 I-II K-R-HA-tagged construct immunoprecipitated with anti-HA antibody showed that the direct binding of ubiquitin to the I-II loop was abolished by mutation of all lysine residues (Fig. 5, A and B). The ladder of bands corresponding to the size expected for the addition of one or more ubiquitins to the palmitoylated I-II loop (Fig. 5A, lane 3) was absent for both the anti-HA (Fig. 5A, lanes 4–7) and anti-ubiquitin antibodies (Fig. 5B).

The Ca<sub>v</sub>β subunit is known to interact with the AID within the first half of the I-II loop (7). To identify which part of the I-II loop is involved in ubiquitination, two further K-R mutant HA-tagged constructs were made: one in which the first four lysine residues were mutated to arginines, referred to as 5' K-R, and the second in which the last eight lysines were mutated to arginines, 3' K-R. Western blots of the palm Ca<sub>v</sub>2.2 I-II K-R-HA-tagged constructs immunoprecipitated with the anti-HA and blotted with anti-HA and anti-ubiquitin antibodies show that both constructs were able to bind ubiquitin (Fig. 5, C and D). This indicates either that ubiquitin binds to more than one lysine residue or that it is able to switch to another lysine if the preferred lysine is no longer available.

To test whether the ladder of bands in Figs. 3 and 5 represents monoubiquitination of multiple lysine residues or oligoubiquitination of a single residue, we used a GFP-tagged mutant ubiquitin construct in which all seven lysine residues were mutated to arginines, GFP-Ub<sup>KO</sup> (18). This mutant ubiquitin can no longer form polyubiquitinated chains but can still compete with endogenous ubiquitin to monoubiquitinate the substrate. When included in the transfections, a single GFP-Ub<sup>KO</sup> was found to bind to the palmitoylated Ca<sub>v</sub>2.2 I-II loop (Fig. 6A, lanes 3 and 4; data quantified in Fig. 6B), whereas the ladder of multiple GFP-ubiquitin (Ub) moieties bound was absent (Fig. 6, A and B, compare lanes 4 and lanes 6). Taken together, these data suggest that only a single lysine residue on the I-II loop is oligoubiquitinated sequentially, but if the preferred lysine has been mutated to arginine, this oligoubiquitination can occur on another available lysine.

**Ca<sub>v</sub>β1b Does Not Protect Ca<sub>v</sub>2.2 I-II Loop from Oligoubiquitination**—Ca<sub>v</sub>β1b protected palm Ca<sub>v</sub>2.2 I-II-GFP from being degraded (Fig. 2, A–D). Western blotting, however, showed that palm Ca<sub>v</sub>2.2 I-II-HA was ubiquitinated even when co-transfected with Ca<sub>v</sub>β1b (Fig. 3, C and E). Although ubiquitination is usually the initial step on the proteasomal degradation pathway, mono- or oligoubiquitination can also lead to different outcomes for the protein (14). We therefore asked whether the I-II loop that was interacting with the Ca<sub>v</sub>β1b was protected from ubiquitination or was also ubiquitinated. To do this, we co-expressed palm Ca<sub>v</sub>2.2 I-II-HA with either GFP-tagged Ca<sub>v</sub>β1b (Ca<sub>v</sub>β1b-GFP) or with GFP (without Ca<sub>v</sub>β) as a control in tsA-201 cells, immunoprecipitated the Ca<sub>v</sub>β with anti-GFP antibody, and immunoblotted with anti-HA and anti-Ub antibodies.

Ca<sub>v</sub>β1b-GFP was able to co-immunoprecipitate non-ubiquitinated palm Ca<sub>v</sub>2.2 I-II-HA (Fig. 6C, lanes 4 and 5, shown with an arrow). In addition, the anti-HA antibody also detected a ladder of bands likely to represent ubiquitinated I-II loop

products (Fig. 6C, top, lanes 4 and 5) that were also detected using the anti-Ub antibody (Fig. 6C, bottom, marked with asterisks). Both non-ubiquitinated and ubiquitinated palm Ca<sub>v</sub>2.2 I-II-HA products immunoprecipitated directly with the anti-HA antibody are shown in lanes 6–9 for comparison. These results, quantified in Fig. 5D, could indicate that the I-II loop does not need to be ubiquitinated to interact with Ca<sub>v</sub>β subunits but that Ca<sub>v</sub>β is able to interact with both ubiquitinated and non-ubiquitinated I-II loop. Alternatively, ubiquitin may interact with the Ca<sub>v</sub>2.2 I-II after it has bound to Ca<sub>v</sub>β.

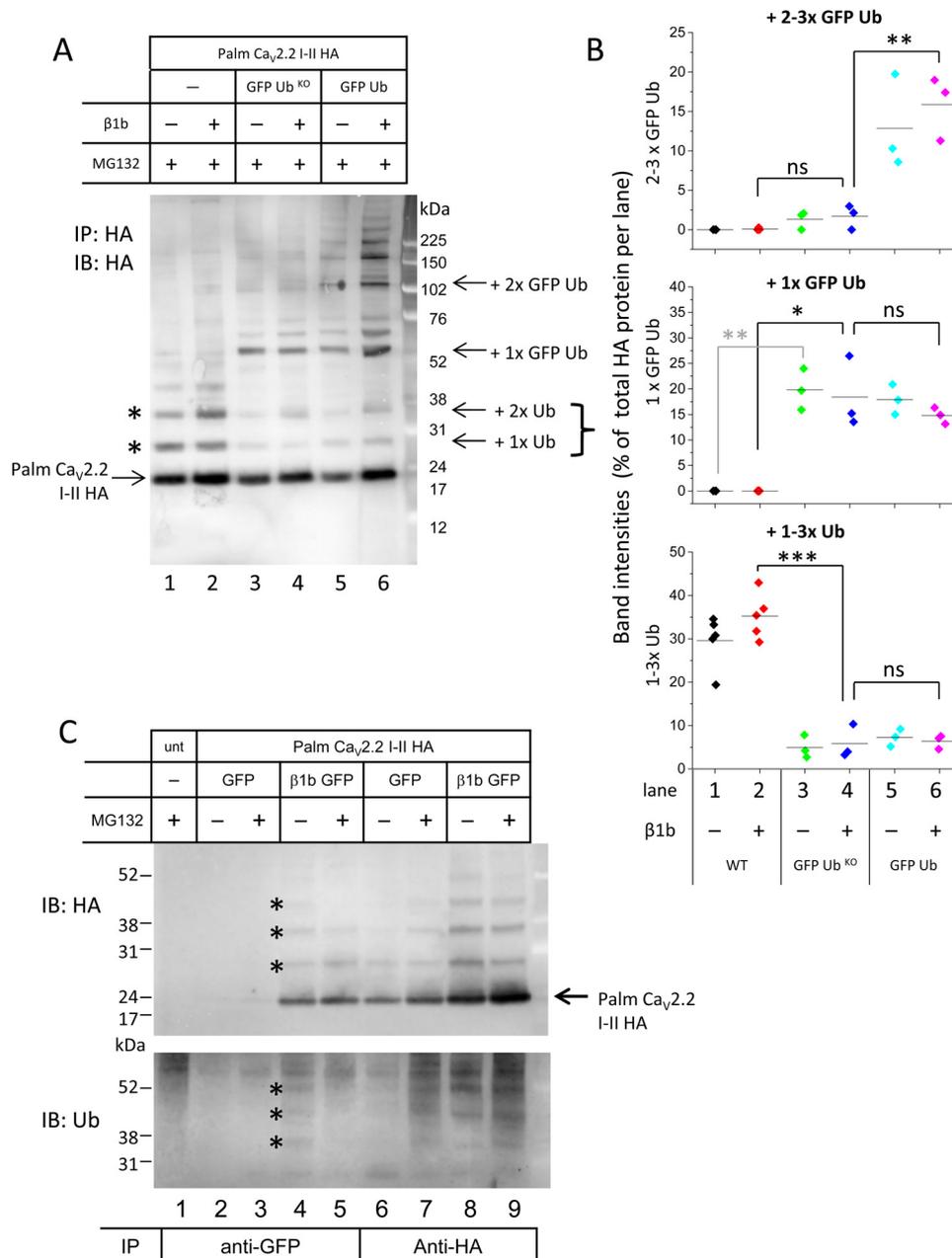
**Mutation of all Lysine Residues in the I-II Loop Reduces Degradation of Full-length Ca<sub>v</sub>2.2 Channels**—To determine whether ubiquitination of the I-II loop plays a role in degradation of the full-length channel, the 12 lysine residues in the I-II loop of Ca<sub>v</sub>2.2 (Fig. 4A) were mutated to arginines. The wild-type and mutated channels were expressed in tsA-201 cells and examined by confocal imaging. In this experiment, the Ca<sub>v</sub>2.2 α1 subunits contained an extracellular HA tag (6), and GFP was fused to the N terminus to give GFP-Ca<sub>v</sub>2.2-HA (wild type (WT)) or GFP-Ca<sub>v</sub>2.2 K-R-HA (containing 12 lysine-to-arginine mutations). Full-length channels were expressed with auxiliary Ca<sub>v</sub>α<sub>2</sub>δ-1 subunits in the presence or absence of MG132 and the presence or absence of Ca<sub>v</sub>β1b-mCherry (Fig. 7, A and C). As with the palm Ca<sub>v</sub>2.2 I-II loop, MG132 caused a significant increase in the total GFP-Ca<sub>v</sub>2.2-HA fluorescence measured in the absence of Ca<sub>v</sub>β1b (Fig. 7, A and B), whereas MG132 had no effect on the GFP fluorescence of GFP-Ca<sub>v</sub>2.2 K-R-HA (Fig. 7, C and D). In the absence of Ca<sub>v</sub>β1b, total GFP fluorescence intensity was significantly lower for GFP-Ca<sub>v</sub>2.2-HA WT than for GFP-Ca<sub>v</sub>2.2 K-R-HA (Fig. 7E). In contrast, in the presence of Ca<sub>v</sub>β1b, GFP fluorescence intensities were not significantly different between the two conditions (Fig. 7F). This suggests that the K-R mutations within the I-II loop protect the full-length channel from degradation in the absence of Ca<sub>v</sub>β.

We next wanted to determine whether mutation of the lysines in the I-II loop had any effect on trafficking of the full-length channel. Full-length GFP-Ca<sub>v</sub>2.2-HA (either WT or K-R mutant) was expressed in tsA-201 cells together with Ca<sub>v</sub>β1b and Ca<sub>v</sub>α<sub>2</sub>δ-1. In non-permeabilized conditions, an anti-HA antibody was used to label the channel in the proximity of the plasma membrane (Fig. 8A). HA labeling was measured on a line (width of 10 pixels) drawn around the cell, and this was compared with the intracellular GFP fluorescence of the same construct measured within the cell and excluding that at the plasma membrane. Although intracellular GFP fluorescence was slightly higher for GFP-Ca<sub>v</sub>2.2 K-R-HA than for the WT channel in this experiment, the HA fluorescence intensities at the plasma membrane were unchanged (Fig. 8B), indicating that the 12 mutations in the I-II loop have little effect on Ca<sub>v</sub>2.2 trafficking.

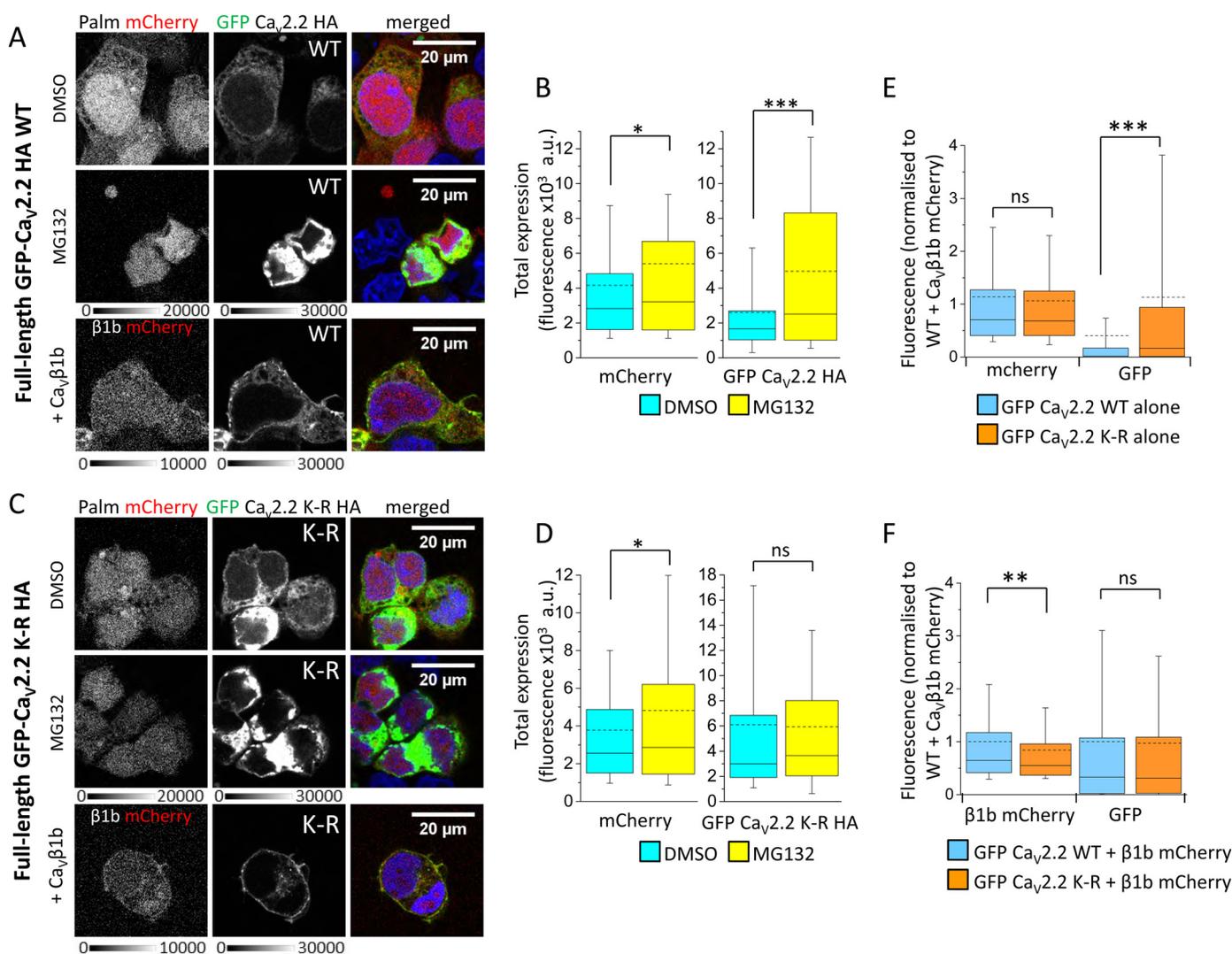
Electrophysiological examination of Ca<sub>v</sub>2.2-HA channels expressed in tsA-201 cells together with Ca<sub>v</sub>β1b-GFP and Ca<sub>v</sub>α<sub>2</sub>δ-1 showed that the K-R mutant produced a decrease in current density compared with the WT channel (Fig. 8, C and D). The reduction in current density attributed to the K-R mutation was paralleled by a reduction in whole cell conductance ( $G_{\max}$ ) through the mutated channel (Fig. 8D, left). The



## Ubiquitination and Degradation of the Ca<sub>v</sub> Channel I-II Loop



**FIGURE 6. Oligoubiquitination, occurring on a single lysine within the I-II loop, does not prevent an interaction with Ca<sub>v</sub>β1b-GFP.** *A*, Western blot showing tsA-201 cells expressing WT palm Ca<sub>v</sub>2.2 I-II-HA in the absence (*lanes 1, 3, and 5*) or presence (*lanes 2, 4, and 6*) of Ca<sub>v</sub>β1b and in the presence of GFP-Ub<sup>KO</sup> (*lanes 3 and 4*) or GFP-Ub (*lanes 5 and 6*) immunoprecipitated (IP) and immunoblotted (IB) with anti-HA antibody. *Lanes 1 and 2* show palm Ca<sub>v</sub>2.2 I-II-HA bound to endogenous ubiquitins (shown with asterisks). GFP-Ub competes with endogenous ubiquitin to give larger products (arrows for +1 × or +2 × GFP-Ub), but only one dominant band is seen when GFP-Ub<sup>KO</sup> is used. *B*, quantification of band intensities of ubiquitinated palm Ca<sub>v</sub>2.2 I-II-HA expressed as a percentage of total HA-detected products per lane. Samples are all in the presence of 4 μM MG132. Band intensities were measured using ImageJ and expressed as scatter plots and show the percentage of HA-products bound to two or three GFP-tagged ubiquitins (*top*), a single GFP-Ub (*middle*), or one to three endogenous ubiquitins (*bottom*). *Lanes* are the same as those shown in *A*. In the presence of GFP-Ub<sup>KO</sup>, there is a significant reduction in the amount of endogenous ubiquitins bound to palm Ca<sub>v</sub>2.2 I-II-HA; in the absence of Ca<sub>v</sub>β1b, mean band intensities (solid lines) ± S.E. were 29.6 ± 2.68 (WT alone; black symbols), 4.92 ± 1.51 (WT + GFP-Ub<sup>KO</sup>; green), and 7.24 ± 1.16% (WT + GFP-Ub; cyan), and in the presence of Ca<sub>v</sub>β1b, mean band intensities were 35.26 ± 2.35 (WT; red), 5.84 ± 2.26 (WT + GFP-Ub<sup>KO</sup>; blue; \*\*\*, *p* < 0.0001), and 6.39 ± 0.95% (WT + GFP-Ub; pink). Statistical analysis was carried out using one-way analysis of variance with Bonferroni post hoc test. For WT alone, there is no measurable amount of GFP-ubiquitin bound (*middle and top plots*). Transfecting GFP-Ub<sup>KO</sup> gives a statistically significant increase in the binding of a single GFP-Ub to palm Ca<sub>v</sub>2.2 I-II-HA in both the absence and presence of Ca<sub>v</sub>β1b (*middle plot*; 19.86 ± 2.34%; green; \*\*, *p* = 0.0011 and 18.40 ± 4.06%; blue; \*, *p* = 0.011), whereas there is no increase in the amount of palm Ca<sub>v</sub>2.2 I-II-HA bound to two or three GFP-Ub moieties (*top*, 1.31 ± 0.66 and 1.70 ± 0.88%). In contrast, in the presence of GFP-Ub, a single GFP-Ub bound (*middle plot*) increases to 17.91 ± 1.7 and 14.78 ± 0.92% in the absence and presence of Ca<sub>v</sub>β1b, respectively, and the addition of two to three GFP-Ub bound (*top*) also increases to 12.87 ± 3.47 and 15.89 ± 2.35% (\*\*, *p* = 0.0048). *C*, Western blots showing tsA-201 cells expressing WT palm Ca<sub>v</sub>2.2 I-II-HA in the presence of free GFP (*lanes 2, 3, 6, and 7*) or Ca<sub>v</sub>β1b tagged with GFP (*lanes 4, 5, 8, and 9*) immunoprecipitated with anti-GFP (*lanes 1–5*) or anti-HA (*lanes 6–9*) and immunoblotted with anti-HA (*top*) or anti-Ub (*bottom*) antibodies. Untransfected (*unt*) cells are shown in *lane 1*. MG132 (4 μM) is included where shown. The size of palm Ca<sub>v</sub>2.2 I-II-HA that is not ubiquitinated is shown with an arrow. Asterisks show ubiquitinated products detected with both anti-HA and anti-Ub antibodies. *ns*, non-significant.



**FIGURE 7. Full-length Ca<sub>v</sub>2.2 is protected from degradation by mutation of the lysine residues in its I-II loop.** *A*, representative confocal images of tsA-201 cells co-expressing palm mCherry (without Ca<sub>v</sub>β; *left panel*) and full-length GFP-Ca<sub>v</sub>2.2-HA (*middle panel*). The *top row* shows the WT Ca<sub>v</sub>2.2 in the absence of Ca<sub>v</sub>β and MG132, the *middle row* shows palm mCherry and GFP-Ca<sub>v</sub>2.2-HA in the presence of MG132 (4 μM), and the *third row* shows Ca<sub>v</sub>β1b-mCherry with GFP-Ca<sub>v</sub>2.2-HA. The *panel on the right* shows the merged image; DAPI was used to visualize the nucleus (*blue*). The *scale bar* is 20 μm, and the *grayscale calibration bar* is shown below each set of images. *B*, box and whisker plots showing fluorescence intensity in the absence of MG132 (DMSO control; *cyan bar*; *n* = 415 cells) or in the presence of MG132 (*yellow bar*; *n* = 75) of the cells represented in *A*. The *box* shows the range of 25–75%, the *whiskers* range from 10 to 90%, the *median* is shown as a *solid line*, and the *mean* is shown as a *dotted line*. GFP fluorescence was measured in all cells that were expressing mCherry. Palm mCherry fluorescence (*left*) was slightly increased in the presence of MG132 (\*, *p* = 0.046, Student's *t* test) with a mean fluorescence ± S.E. of 4155 ± 210 arbitrary units (a.u.) in DMSO versus 5403 ± 898 arbitrary units in MG132, whereas palm Ca<sub>v</sub>2.2 I-II-GFP (*right*) fluorescence was significantly increased by MG132 with mean ± S.E. of 2609 ± 153 (DMSO) and 4973 ± 598 arbitrary units (MG132) (\*\*\*, *p* < 0.0001, Student's *t* test). *C*, as for *A* but for cells expressing Ca<sub>v</sub>2.2 that had all 12 lysine residues in its I-II loop mutated to arginines (K-R). *D*, box and whisker plots (as in *B*) showing fluorescence intensity in the absence of MG132 (DMSO control; *cyan bar*; *n* = 269 cells) or in the presence of MG132 (*yellow bar*; *n* = 138) of the cells represented in *C*. Palm mCherry fluorescence (*left*) was slightly increased in the presence of MG132 (\*, *p* = 0.02, Student's *t* test) with mean fluorescence ± S.E. of 3787 ± 233 (DMSO) and 4822 ± 60 arbitrary units (MG132), whereas there was no difference in palm Ca<sub>v</sub>2.2 I-II K-R-GFP fluorescence (*right*; mean ± S.E. of 6098 ± 449 (DMSO) and 5936 ± 520 arbitrary units (MG132)). *E* and *F*, box and whisker plots (as in *B*) of normalized fluorescence intensity of mCherry and GFP for GFP-Ca<sub>v</sub>2.2-HA WT and GFP-Ca<sub>v</sub>2.2 K-R-HA. Data are taken from three independent experiments and normalized for mCherry and GFP fluorescence of WT in the presence of Ca<sub>v</sub>β1b-mCherry (shown in *F*). In the absence of Ca<sub>v</sub>β1b (*E*), mCherry fluorescence (*left*) was not significantly different between WT (*light blue*; *n* = 1678 cells) and the K-R mutant (*orange*; *n* = 1046) with mean normalized fluorescence values ± S.E. of 1.138 ± 0.033 and 1.064 ± 0.037, whereas normalized GFP fluorescence (*right*) was significantly lower for WT than the K-R mutant (\*\*\*, *p* < 0.0001, Student's *t* test) with mean values of 0.402 ± 0.032 and 1.132 ± 0.069. In the presence of Ca<sub>v</sub>β1b (*F*), although Ca<sub>v</sub>β1b-mCherry expression was slightly lower for K-R mutant (*orange*; *n* = 734; mean, 0.841 ± 0.033) than WT (*light blue*; *n* = 738; mean, 1.00 ± 0.039; \*\*, *p* = 0.0018, Student's *t* test), GFP fluorescence was not significantly different (mean values, 1.00 ± 0.065 for WT and 0.973 ± 0.065 for K-R mutant). *ns*, non-significant.

and palmitoylation of both residues leads to greater stability at the plasma membrane (22). G protein α subunits follow a two-signal model for targeting to the plasma membrane (23). For non-myristoylated G proteins, such as Gα<sub>q</sub>, an α-helical polybasic motif at the N terminus provides the initial signal (24, 25), and this signal, in conjunction with an association with the G protein βγ subunit (Gβγ) (26), allows targeting to the plasma

membrane and subsequent palmitoylation (25). Tethering of the Ca<sub>v</sub>β2e subunit to the plasma membrane also requires a polybasic motif at the N terminus (10). However, although the I-II loop of Ca<sub>v</sub>2.2 contains multiple basic amino acids at the N terminus (Fig. 4A), this region alone is insufficient to target it to the plasma membrane, even when it is palmitoylated, in the absence of Ca<sub>v</sub>β subunits.

## Ubiquitination and Degradation of the $\text{Ca}_v$ Channel I-II Loop

Binding of  $\text{Ca}_v\beta$  to the AID is thought to induce a dramatic change in secondary structure of the I-II loop (27). In the absence of  $\text{Ca}_v\beta$ , circular dichroism (CD) measurements showed that the AID exists as a random coil (28), whereas in the presence of  $\text{Ca}_v\beta$ , the AID was shown to have an  $\alpha$ -helical structure (28, 29). Secondary structure predictions suggested that the entire linker between the last transmembrane domain of domain I (IS6) and the AID could form a continuous  $\alpha$ -helix (28), and CD measurements showed that this was the case (30). The helical content was found to be significantly higher for this region of  $\text{Ca}_v2.2$  than that of  $\text{Ca}_v1.2$  (31). It appears that  $\text{Ca}_v\beta$  is able to interact with an unfolded I-II loop via its AID, inducing a conformational change and extending the  $\alpha$ -helical structure along the I-II linker toward the IS6 (28). The N terminus of the I-II loop contains a number of basic amino acids (see Fig. 4A). Fig. 9 shows two helical wheel diagrams of the region of the I-II linker upstream of the AID (*left*) and including the AID (*right*); formation of an  $\alpha$ -helix would allow two clusters of basic amino acids to align along one side and form a polybasic region. This may act as a trafficking signal to direct the construct to the plasma membrane and promote stable dipalmitoylation. Interestingly, the Trp-391 residue, which is critical for the interaction between the I-II loop and the  $\text{Ca}_v\beta$  subunit, is located along the same face of the helix although beyond the polybasic regions (Fig. 9).

Alternatively, formation of an  $\alpha$ -helix may allow another binding partner to interact with the I-II loop and direct it to the plasma membrane. The I-II linkers of  $\text{Ca}_v2.1$  (32) and  $\text{Ca}_v2.2$  (33) were found to interact directly with  $G\beta\gamma$ . We have previously shown that  $\text{Ca}_v\beta$  and  $G\beta\gamma$  subunits bind at the same time and that  $\text{Ca}_v\beta$  must be present for the  $G\beta\gamma$  subunit to induce voltage-dependent modulation of  $\text{Ca}_v2.2$  (5). Binding of  $G\beta\gamma$  subunits was found to be dependent on the formation of a rigid IS6-AID linker induced by the binding of  $\text{Ca}_v\beta$  (34). The G protein  $G\alpha_q$  has been shown to require an association with  $G\beta\gamma$  before it can be palmitoylated and stabilized at the plasma membrane (26). To examine whether  $G\beta\gamma$  subunits were involved in the trafficking of the I-II loop constructs, we included a minigene derived from the C terminus of  $\beta$ -adrenergic receptor kinase (5) to bind and remove free  $G\beta\gamma$  subunits. Confocal experiments showed that transfection of  $\beta$ -adrenergic receptor kinase had no effect on the trafficking of palm  $\text{Ca}_v2.2$  I-II in the presence of  $\text{Ca}_v\beta1b$ ,<sup>4</sup> providing no evidence that  $G\beta\gamma$  binding is involved.

In the absence of  $\text{Ca}_v\beta$ , the I-II loop remains in a misfolded state. The  $\text{Ca}_v\beta$  subunit is able to act as a molecular chaperone to induce correct folding of the I-II loop of the  $\text{Ca}_v\alpha1$  subunit. Misfolded proteins are more likely to aggregate as they have exposed hydrophobic domains, and a key role of the chaperone is to interact co-translationally as the nascent protein emerges from the ribosome and prevent misfolding and aggregation (35). In the absence of the chaperone, misfolded proteins are rapidly targeted to the ubiquitin-proteasome system for degradation.

Substrates for the ubiquitin-proteasome system are tagged with ubiquitin; addition of ubiquitin moieties is thought to

occur sequentially, one at a time, with the rate-limiting step being the addition of the first ubiquitin (36). It has previously been suggested that a chain of four ubiquitins is the minimum signal for targeting the substrate to the proteasome (37), but more recently it has been shown that only a single ubiquitin is needed for small proteins (20–150 amino acids) (38). Ubiquitination also has other functions, however, such as protein trafficking (14), endocytosis, and signaling (39), which are independent of the proteasome. Our data show that the palmitoylated I-II loop carries up to four ubiquitins (Fig. 3) even in the presence of the  $\text{Ca}_v\beta$  subunit and the proteasomal inhibitor MG132; conditions in which the protein is not degraded. Relevant to this, a mass spectrometry study has shown that in almost 50% of cases, ubiquitination of proteins has a non-proteasomal function (40).

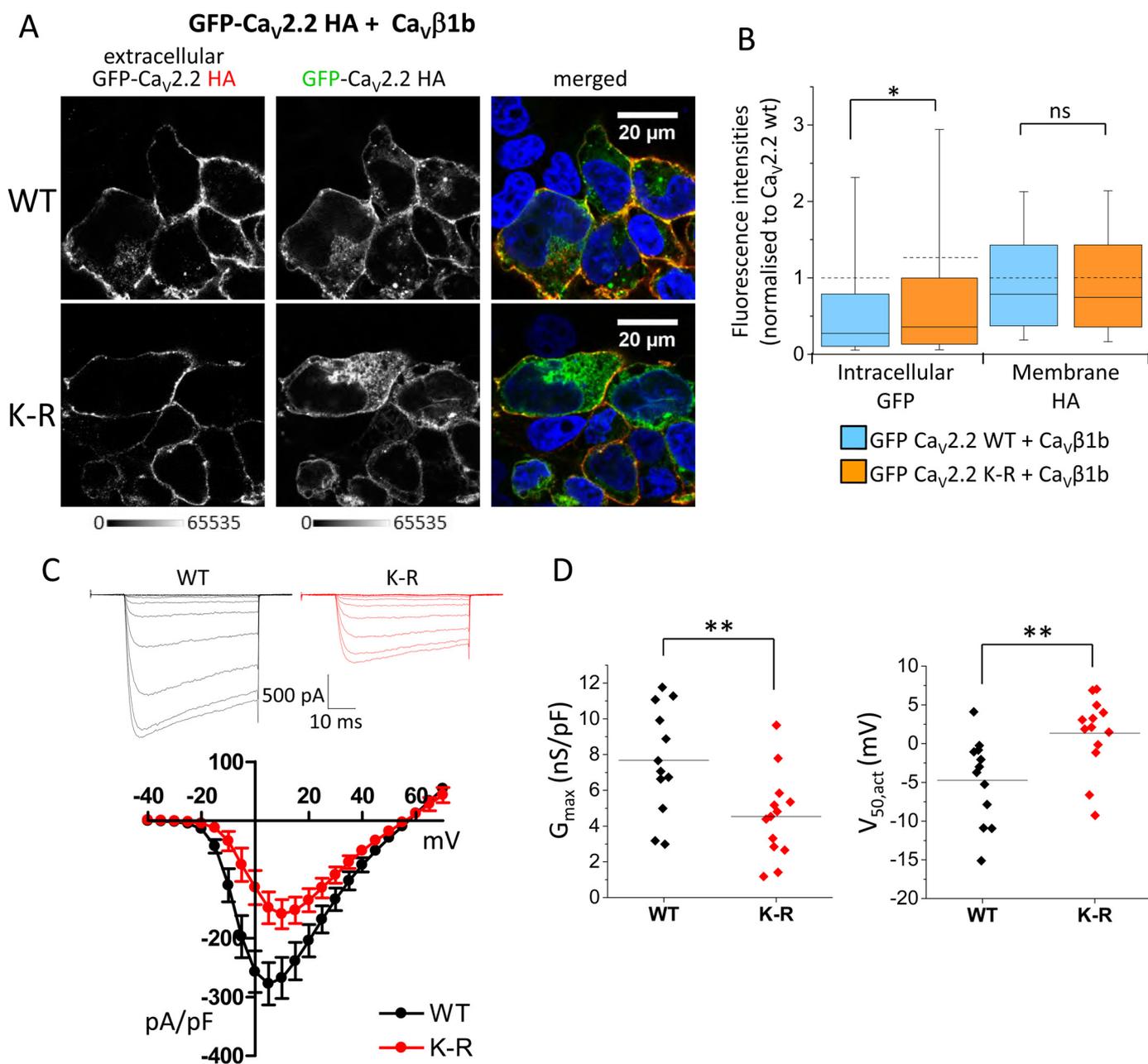
In the absence of  $\text{Ca}_v\beta$ , the I-II loop was rapidly degraded as shown in Fig. 2. It has been shown that nascent proteins can be ubiquitinated co-translationally and targeted for degradation before translation has finished (41). Ubiquitination does not necessarily lead to degradation of newly synthesized ubiquitinated proteins; if correct folding does occur with the help of a chaperone, the ubiquitins may be removed by the action of deubiquitinating enzymes (42). In this regard, binding of the  $\text{Ca}_v\beta$  subunit to the ubiquitinated I-II loop clearly rescues it from degradation, although our evidence does not indicate it promotes deubiquitination. By contrast, it is possible that oligoubiquitination is functionally important for the effect of  $\text{Ca}_v\beta$  because the full-length  $\text{Ca}_v2.2$  K-R mutant showed reduced functional expression, possibly reflecting reduced interaction with the  $\text{Ca}_v\beta$  subunit.

Mutation of the 12 lysine residues in the I-II loop of the full-length  $\text{Ca}_v2.2$  channel protected the channel from degradation. The full-length K-R mutant channel still contains multiple internal lysine residues that were not mutated and were therefore available as ubiquitination sites. However, the fact that the K-R channel was less degraded than WT shows that the 12 lysines within the I-II loop have a critical role in ubiquitination and degradation.

Trafficking of the palmitoylated  $\text{Ca}_v2.2$  I-II loop to the plasma membrane required the presence of the palmitoylation signal, the I-II loop, and the  $\text{Ca}_v\beta$  subunit. Although the palmitoylated K-R mutant I-II loop was no longer ubiquitinated and degraded, it also was not trafficked to the plasma membrane in the absence of the  $\text{Ca}_v\beta$  subunit; trafficking was restored when  $\text{Ca}_v\beta$  was also co-expressed. It is possible that correct folding, which is only achieved in the presence of the  $\text{Ca}_v\beta$  subunit, exposes a trafficking signal. Typical trafficking signals have been identified as short linear amino acid sequences, but it has been shown that export of the Kir2.1 channel from the Golgi to the plasma membrane requires the correctly folded N and C termini of the channel (43). The trafficking signal was only exposed when two separate domains were folded correctly in the tertiary structure of the channel.

In conclusion, our experiments indicate that, in the absence of  $\text{Ca}_v\beta$  subunit, the I-II loop of  $\text{Ca}_v2.2$  is directly ubiquitinated and rapidly degraded by the proteasome system. When  $\text{Ca}_v\beta$  is present, it is able to bind to the I-II loop even if the loop is already ubiquitinated. Binding appears to induce a conforma-

<sup>4</sup> K. M. Page, unpublished observations.



**FIGURE 8. Full-length GFP-Ca<sub>v</sub>2.2-HA containing 12 lysine-to-arginine mutations in the I-II loop is still trafficked and functionally active in the presence of Ca<sub>v</sub>β1b.** *A*, representative confocal images of tsA-201 cells expressing full-length GFP-Ca<sub>v</sub>2.2-HA with Ca<sub>v</sub>β1b and Ca<sub>v</sub>α<sub>2</sub>δ-1 and showing WT (top) and K-R mutant (bottom). Cells were non-permeabilized and incubated with rat anti-HA antibody overnight to show extracellular HA staining on the plasma membrane (left panels; grayscale) to be compared with intracellular GFP fluorescence (middle panels; grayscale). Merged images are shown on the right; DAPI was used to stain the nuclei (blue). Scale bars are 20 μm, and the grayscale calibration bar is shown below. *B*, box and whisker plots showing normalized fluorescence intensities of the images represented in *B* with intracellular GFP (left) for WT GFP-Ca<sub>v</sub>2.2-HA (light blue;  $n = 1048$  cells) and GFP-Ca<sub>v</sub>2.2 K-R-HA (orange;  $n = 998$  cells) and extracellular HA staining at the plasma membrane (right). GFP fluorescence at the plasma membrane (overlapping with membrane HA labeling) was excluded from the analysis to give intracellular GFP rather than total GFP. Data have been taken from three separate experiments involving three transfections. Although intracellular GFP fluorescence may be slightly increased for the K-R mutant compared with WT (mean fluorescent values  $\pm$  S.E. of  $1.000 \pm 0.069$  for WT and  $1.263 \pm 0.092$  for K-R; \*,  $p = 0.022$ , Student's *t* test), extracellular HA staining is unchanged (mean values,  $1.000 \pm 0.025$  for WT and  $1.003 \pm 0.029$  for K-R). *C*, current-voltage relationships of full-length WT and K-R mutant Ca<sub>v</sub>2.2 channels co-expressed with Ca<sub>v</sub>β1b-GFP and Ca<sub>v</sub>α<sub>2</sub>δ-1 in tsA-201 cells. Top, example traces of WT Ca<sub>v</sub>2.2/β1b-GFP/α<sub>2</sub>δ-1 (−40 to +10 mV in 5-mV increments; black traces) and K-R Ca<sub>v</sub>2.2/β1b-GFP/α<sub>2</sub>δ-1 currents (−40 to +15 mV in 5-mV increments; red traces) recorded from a holding potential of −80 mV. The charge carrier was 1 mM Ba<sup>2+</sup>. Cells expressing wild-type or the K-R mutant channel had the same holding currents of  $-3 \pm 4$  pA ( $n = 12$ ) and  $-3 \pm 4$  pA ( $n = 13$ ), respectively. The scale bars refer to both traces. Bottom, mean current density-voltage relationships (pA/pF) were calculated from current amplitude (pA) measurements taken 15 ms into a 50-s depolarizing voltage pulse. Error bars represent S.E.  $I_{Ba}$  at +5 mV was  $277 \pm 36$  pA/pF ( $n = 12$ ) and  $148 \pm 27$  pA/pF ( $n = 13$ ) for WT and K-R Ca<sub>v</sub>2.2, respectively ( $p = 0.0081$ , Student's *t* test). *D*, characteristic properties of the current density-voltage relationships of WT Ca<sub>v</sub>2.2 and the K-R mutant channel. Values were derived from modified Boltzmann fits of individual current density-voltage plots describing WT and K-R Ca<sub>v</sub>2.2 currents. Left,  $G_{max}$  was  $7.7 \pm 0.9$  nS/pF (mean (solid line)  $\pm$  S.E.;  $n = 12$ ) and  $4.5 \pm 0.7$  nS/pF ( $n = 13$ ) for WT and K-R Ca<sub>v</sub>2.2, respectively (\*\*,  $p = 0.0079$ , Student's *t* test). Right,  $V_{50}$  values were  $-4.7 \pm 1.6$  (n = 12) and  $1.3 \pm 1.3$  mV (n = 13) for WT and K-R Ca<sub>v</sub>2.2, respectively (\*\*,  $p = 0.0071$ , Student's *t* test). ns, non-significant.

## Ubiquitination and Degradation of the Ca<sub>v</sub> Channel I-II Loop

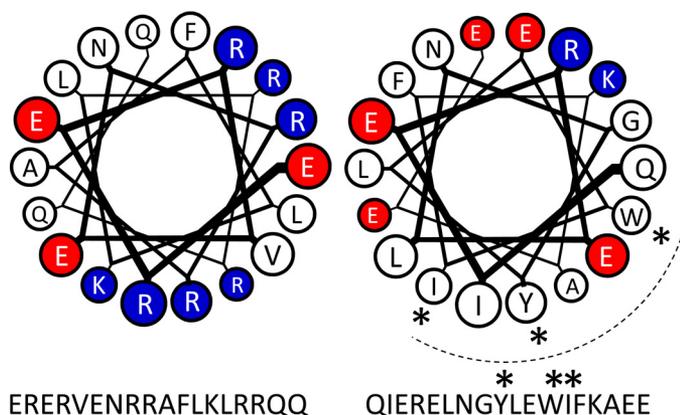


FIGURE 9.  $\alpha$ -Helical model of the 5' region of the Ca<sub>v</sub>2.2 I-II loop.  $\alpha$ -Helical wheel models of the 5' region of the Ca<sub>v</sub>2.2 I-II loop after the IS6 transmembrane domain and immediately preceding the AID (left) and the subsequent sequence including the AID (right; in the same orientation) show two clusters of basic amino acids along one face of the helix. Basic residues are shown in blue, acidic residues are shown in red, and the remainder are shown in white. Residues involved in the interaction with Ca<sub>v</sub> $\beta$  subunits lie along the dotted line; Trp-391, Ile-392, and Tyr-388, which are critical for this interaction (29), are labeled with asterisks.

tional change and the formation of an  $\alpha$ -helix in the I-II loop, thereby allowing the construct to associate with the plasma membrane.

### Experimental Procedures

**Molecular Biology**—The calcium channels used were rabbit Ca<sub>v</sub>2.2 (GenBank Accession number D14157), rat Ca<sub>v</sub> $\alpha$ 2 $\delta$ -1 (M86621), and rat Ca<sub>v</sub> $\beta$ 1b (X61394), all expressed in the pMT2 vector. The green fluorescent protein mut3bGFP (GFP) (44) or mCherry was fused to the C terminus of Ca<sub>v</sub> $\beta$ 1b. GFP-Ub and GFP-Ub<sup>KO</sup> (18) were obtained from Addgene. The first intracellular loop of Ca<sub>v</sub>2.2 (amino acids 356–483, beginning ESGEF... and ending ... KAQ) had a palmitoylation sequence, MTLESIMACCL, added to the N terminus and an HA tag (TSYPYDVPDYA) added to the C terminus to give a construct termed palm Ca<sub>v</sub>2.2 I-II-HA in this study. Further constructs were made where the HA tag was substituted with GFP or mCherry. The mutant palm Ca<sub>v</sub>2.2 I-II K-R-HA was made by mutating all 12 lysine residues in the I-II loop to arginines, and two further constructs were made by mutating either the first four (palm Ca<sub>v</sub>2.2 I-II 5' K-R-HA) or the last eight (palm Ca<sub>v</sub>2.2 I-II 3' K-R-HA) lysines to arginines. The I-II loop containing the 12 lysine-to-arginine substitutions was inserted into the full-length Ca<sub>v</sub>2.2, which also contained an extracellular HA tag (6) and GFP fused to the N terminus to give GFP-Ca<sub>v</sub>2.2 K-R-HA. The sequences of all constructs were confirmed by DNA sequencing.

**Cell Culture and Transfection**—tsA-201 cells (European Collection of Authenticated Cell Cultures (ECACC)) were cultured in Dulbecco's modified Eagle's medium in the presence of 10% fetal bovine serum, penicillin, streptomycin, and 2% GlutaMAX (Invitrogen). For immunocytochemistry and co-immunoprecipitation experiments, transfections were carried out using PolyJet (SigmaGen) according to the manufacturer's instructions using equal ratios of constructs unless otherwise stated. When included, the protease inhibitor MG132 (Calbiochem) was added at a concentration of 4  $\mu$ M 24 h after transfection,

and the cells were harvested or fixed a further 16–18 h afterward. At the concentration of MG132 used (4  $\mu$ M) and the time of incubation (16–18 h), MG132 was found to be effective at inhibiting proteasomal degradation without being detrimental to cell survival. Total protein yields from transfections in the absence and presence of MG132 were measured (for Western blotting and co-immunoprecipitation analysis) and found to be similar. For electrophysiological experiments, transfections were performed using FuGENE 6 (Promega) with GFP-Ca<sub>v</sub>2.2-HA, Ca<sub>v</sub> $\alpha$ 2 $\delta$ -1, and Ca<sub>v</sub> $\beta$ 1b subunits in a ratio of 3:2:2.

**Immunocytochemistry**—Cells were transfected on polylysine-coated coverslips and fixed with 4% paraformaldehyde in Tris-buffered saline (TBS; 20 mM Tris, 150 mM NaCl, pH 7.4) for 5 min. 4',6-Diamidino-2'-phenylindole dihydrochloride (DAPI) was used to stain the nuclei. When used, the plasma membrane stain CM-Dil (ThermoFisher Scientific) was used at a dilution of 1:200 for 20 min at room temperature. Coverslips were mounted in VectaShield (Vector Laboratories). When anti-HA staining was used, cells were incubated with blocking buffer (20% goat serum, 4% BSA in TBS) for 1 h at room temperature before being incubated with rat anti-HA (Roche Applied Science) diluted 1:200 in 0.5 $\times$  blocking buffer at 4  $^{\circ}$ C overnight. After washing, samples were incubated with secondary antibody anti-rat Alexa Fluor 594 at a dilution of 1:500 for 1 h at room temperature before being stained with DAPI and mounted. Samples were viewed on an LSM 780 confocal microscope (Zeiss) using a 63 $\times$ /1.4 numerical aperture oil immersion objective in 16-bit mode. The tile function (3  $\times$  3 tiles; each tile consisting of 1024  $\times$  1024 pixels) was used, and every transfected cell within the image was analyzed to remove collection bias. Confocal optical sections were 1  $\mu$ m, and acquisition settings were kept constant. Images were analyzed using NIH ImageJ. Images that were analyzed were not saturated.

**Western Blotting Analysis**—Transfected tsA-201 cells were harvested in phosphate-buffered saline (PBS) containing protease inhibitors (Complete tablet from Roche Applied Science). Cells were lysed by sonication for 10 s with 1% Igepal in PBS in the presence of protease inhibitors followed by incubation on ice for 30 min, and whole cell lysates (WCLs) were collected after centrifugation (14,000  $\times$  g for 30 min at 4  $^{\circ}$ C). Samples were incubated for 15 min at 55  $^{\circ}$ C with 100 mM dithiothreitol and 2 $\times$  Laemmli sample buffer, and proteins were separated by SDS-PAGE on 4–12% Bis-Tris gels and then transferred to polyvinylidene fluoride membranes. Membranes were incubated in blocking buffer (10 mM Tris, pH 7.4, 500 mM NaCl, 0.5% Igepal, 3% BSA) for 1 h followed by incubation with the primary antibody. The following primary antibodies were used: rat anti-HA (Roche Applied Science) at 1:1000 overnight, mouse anti-ubiquitin (P4D1, Santa Cruz Biotechnology) at 1:500 overnight, and mouse anti-GFP (Roche Applied Science) at 1:1000 for 1 h, all at 4  $^{\circ}$ C. The appropriate secondary antibodies coupled to horseradish peroxidase were incubated at a dilution of 1:2000 for 1 h at room temperature. Proteins were detected using the enhanced ECL Plus reagent (GE Healthcare) on a Typhoon 9410 scanner (GE Healthcare). Quantification of Western blotting was carried out using the gel analysis tool of ImageJ. A rectangular section was placed over the entire lane, and each selected lane was plotted as a line profile. The peak

band intensities above background were measured. This allows the changes in background along the length of the lane to be taken into account and accurately subtracted.

**Immunoprecipitation**—The total protein concentration was determined (Bradford assay, Bio-Rad) for each sample, and 1 mg of WCL was precleared on 50 μg of protein A-Sepharose (GE Healthcare) for 2 h at 4 °C. Sepharose beads were discarded, and supernatants were incubated with rabbit anti-HA (Sigma) or rabbit anti-GFP (Clontech) at a dilution of 1:200 overnight at 4 °C. Immunoprecipitated proteins were captured on 60 μg of protein A-Sepharose beads for 2 h at 4 °C. Beads were washed five times with PBS containing 0.1% Igepal and incubated for 15 min at 55 °C with 100 mM dithiothreitol and 2× Laemmli sample buffer. Eluted proteins were then resolved by SDS-PAGE.

**Electrophysiology**—Whole cell voltage clamp recordings were performed on tsA-201 cells at room temperature (20–24 °C). Single cells were clamped using an Axopatch 200B patch clamp amplifier (Axon instruments). Borosilicate glass patch pipettes were filled with a solution containing 140 mM cesium aspartate, 5 mM EGTA, 2 mM MgCl<sub>2</sub>, 0.1 mM CaCl<sub>2</sub>, 2 mM K<sub>2</sub>ATP, and 10 mM HEPES. CsOH was added to achieve pH 7.2. The external solution contained 150 mM tetraethylammonium bromide, 3 mM KCl, 1 mM NaHCO<sub>3</sub>, 1 mM MgCl<sub>2</sub>, 10 mM HEPES, 4 mM glucose, and 1 mM BaCl<sub>2</sub>. pH was adjusted to 7.4 with Tris base. Current density-voltage relationships were fitted with a modified Boltzmann equation as follows:  $I = G_{\max} \times (V - V_{\text{rev}}) / (1 + \exp(-(V - V_{50, \text{act}}) / k))$  where  $I$  is the current density (in pA/pF),  $G_{\max}$  is the maximum conductance (in nS/pF),  $V_{\text{rev}}$  is the reversal potential,  $V_{50, \text{act}}$  is the midpoint voltage for current activation, and  $k$  is the slope factor.

**Author Contributions**—A. C. D. and K. M. P. designed the experiments. K. M. P. made the cDNA constructs and performed all confocal microscopy, immunoprecipitation, and Western blotting experiments. K. M. P. analyzed data. S. W. R. designed, performed, and analyzed electrophysiology experiments. K. M. P. and A. C. D. wrote the manuscript.

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## References

1. Waithe, D., Ferron, L., Page, K. M., Chaggar, K., and Dolphin, A. C. (2011)  $\beta$ -Subunits promote the expression of Ca<sub>v</sub>2.2 channels by reducing their proteasomal degradation. *J. Biol. Chem.* **286**, 9598–9611
2. Simms, B. A., and Zamponi, G. W. (2012) Trafficking and stability of voltage-gated calcium channels. *Cell. Mol. Life Sci.* **69**, 843–856
3. Catterall, W. A. (2000) Structure and regulation of voltage-gated Ca<sup>2+</sup> channels. *Annu. Rev. Cell Dev. Biol.* **16**, 521–555
4. Dolphin, A. C. (2012) Calcium channel auxiliary  $\alpha_2\delta$  and  $\beta$  subunits: trafficking and one step beyond. *Nat. Rev. Neurosci.* **13**, 542–555
5. Meir, A., Bell, D. C., Stephens, G. J., Page, K. M., and Dolphin, A. C. (2000) Calcium channel  $\beta$  subunit promotes voltage-dependent modulation of  $\alpha_1B$  by  $G\beta\gamma$ . *Biophys. J.* **79**, 731–746
6. Cassidy, J. S., Ferron, L., Kadurin, I., Pratt, W. S., and Dolphin, A. C. (2014) Functional exofacially tagged N-type calcium channels elucidate the interaction with auxiliary  $\alpha_2\delta$ -1 subunits. *Proc. Natl. Acad. Sci. U.S.A.* **111**, 8979–8984
7. Pragnell, M., De Waard, M., Mori, Y., Tanabe, T., Snutch, T. P., and Campbell, K. P. (1994) Calcium channel  $\beta$ -subunit binds to a conserved motif in the I-II cytoplasmic linker of the  $\alpha_1$ -subunit. *Nature* **368**, 67–70
8. De Waard, M., Witcher, D. R., Pragnell, M., Liu, H., and Campbell, K. P. (1995) Properties of the  $\alpha_1$ - $\beta$  anchoring site in voltage-dependent Ca<sup>2+</sup> channels. *J. Biol. Chem.* **270**, 12056–12064
9. Chien, A. J., Carr, K. M., Shirokov, R. E., Rios, E., and Hosey, M. M. (1996) Identification of palmitoylation sites within the L-type calcium channel  $\beta_{2a}$  subunit and effects on channel function. *J. Biol. Chem.* **271**, 26465–26468
10. Miranda-Laferte, E., Ewers, D., Guzman, R. E., Jordan, N., Schmidt, S., and Hidalgo, P. (2014) The N-terminal domain tethers the voltage-gated calcium channel  $\beta_{2e}$ -subunit to the plasma membrane via electrostatic and hydrophobic interactions. *J. Biol. Chem.* **289**, 10387–10398
11. Bichet, D., Cornet, V., Geib, S., Carlier, E., Volsen, S., Hoshi, T., Mori, Y., and De Waard, M. (2000) The I-II loop of the Ca<sup>2+</sup> channel  $\alpha_1$  subunit contains an endoplasmic reticulum retention signal antagonized by the  $\beta$  subunit. *Neuron* **25**, 177–190
12. Fang, K., and Colecraft, H. M. (2011) Mechanism of auxiliary  $\beta$ -subunit-mediated membrane targeting of L-type (Ca<sub>v</sub>1.2) channels. *J. Physiol.* **589**, 4437–4455
13. Altier, C., Garcia-Caballero, A., Simms, B., You, H., Chen, L., Walcher, J., Tedford, H. W., Hermosilla, T., and Zamponi, G. W. (2011) The Cav $\beta$  subunit prevents RFP2-mediated ubiquitination and proteasomal degradation of L-type channels. *Nat. Neurosci.* **14**, 173–180
14. Hicke, L., and Dunn, R. (2003) Regulation of membrane protein transport by ubiquitin and ubiquitin-binding proteins. *Annu. Rev. Cell Dev. Biol.* **19**, 141–172
15. Thiyagarajan, M. M., Bigras, E., Van Tol, H. H., Hébert, T. E., Evanko, D. S., and Wedegaertner, P. B. (2002) Activation-induced subcellular redistribution of  $G\alpha_s$  is dependent upon its unique N-terminus. *Biochemistry* **41**, 9470–9484
16. Hicke, L. (2001) Protein regulation by monoubiquitin. *Nat. Rev. Mol. Cell Biol.* **2**, 195–201
17. Pickart, C. M. (2001) Mechanisms underlying ubiquitination. *Annu. Rev. Biochem.* **70**, 503–533
18. Bergink, S., Salomons, F. A., Hoogstraten, D., Groothuis, T. A., de Waard, H., Wu, J., Yuan, L., Citterio, E., Houtsmuller, A. B., Neeffjes, J., Hoeijmakers, J. H., Vermeulen, W., and Dantuma, N. P. (2006) DNA damage triggers nucleotide excision repair-dependent monoubiquitylation of histone H2A. *Genes Dev.* **20**, 1343–1352
19. Leroy, J., Richards, M. W., Butcher, A. J., Nieto-Rostro, M., Pratt, W. S., Davies, A., and Dolphin, A. C. (2005) Interaction via a key tryptophan in the I-II linker of N-type calcium channels is required for  $\beta_1$  but not for palmitoylated  $\beta_2$ , implicating an additional binding site in the regulation of channel voltage-dependent properties. *J. Neurosci.* **25**, 6984–6996
20. Gebhart, M., Juhasz-Vedres, G., Zuccotti, A., Brandt, N., Engel, J., Trockenbacher, A., Kaur, G., Obermair, G. J., Knipper, M., Koschak, A., and Striessnig, J. (2010) Modulation of Ca<sub>v</sub>1.3 Ca<sup>2+</sup> channel gating by Rab3 interacting molecule. *Mol. Cell. Neurosci.* **44**, 246–259
21. Kaur, G., Pinggera, A., Ortner, N. J., Lieb, A., Sinnegger-Brauns, M. J., Yarov-Yarovoy, V., Obermair, G. J., Flucher, B. E., and Striessnig, J. (2015) A polybasic plasma membrane binding motif in the I-II linker stabilizes voltage-gated Ca<sub>v</sub>1.2 calcium channel function. *J. Biol. Chem.* **290**, 21086–21100
22. Wedegaertner, P. B., Chu, D. H., Wilson, P. T., Levis, M. J., and Bourne, H. R. (1993) Palmitoylation is required for signaling functions and membrane attachment of  $G_q\alpha$  and  $G_{12}\alpha$ . *J. Biol. Chem.* **268**, 25001–25008
23. Marrari, Y., Crouthamel, M., Irannejad, R., and Wedegaertner, P. B. (2007) Assembly and trafficking of heterotrimeric G proteins. *Biochemistry* **46**, 7665–7677
24. Kosloff, M., Elia, N., and Selinger, Z. (2002) Structural homology discloses a bifunctional structural motif at the N-termini of  $G_\alpha$  proteins. *Biochemistry* **41**, 14518–14523
25. Crouthamel, M., Thiyagarajan, M. M., Evanko, D. S., and Wedegaertner, P. B. (2008) N-terminal polybasic motifs are required for plasma membrane localization of  $G\alpha_s$  and  $G\alpha_q$ . *Cell. Signal.* **20**, 1900–1910
26. Evanko, D. S., Thiyagarajan, M. M., and Wedegaertner, P. B. (2000) Inter-

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- action with Gβγ is required for membrane targeting and palmitoylation of Gα<sub>s</sub> and Gα<sub>q</sub>. *J. Biol. Chem.* **275**, 1327–1336
27. Buraei, Z., and Yang, J. (2010) The β subunit of voltage-gated Ca<sup>2+</sup> channels. *Physiol. Rev.* **90**, 1461–1506
  28. Opatowsky, Y., Chen, C. C., Campbell, K. P., and Hirsch, J. A. (2004) Structural analysis of the voltage-dependent calcium channel β subunit functional core and its complex with the α1 interaction domain. *Neuron* **42**, 387–399
  29. Van Petegem, F., Clark, K. A., Chatelain, F. C., and Minor, D. L. (2004) Structure of a complex between a voltage-gated calcium channel β-subunit and an α-subunit domain. *Nature* **429**, 671–675
  30. Arias, J. M., Murbartián, J., Vitko, I., Lee, J. H., and Perez-Reyes, E. (2005) Transfer of β subunit regulation from high to low voltage-gated Ca<sup>2+</sup> channels. *FEBS Lett.* **579**, 3907–3912
  31. Almagor, L., Chomsky-Hecht, O., Ben-Mocha, A., Hendin-Barak, D., Dascal, N., and Hirsch, J. A. (2012) The role of a voltage-dependent Ca<sup>2+</sup> channel intracellular linker: a structure-function analysis. *J. Neurosci.* **32**, 7602–7613
  32. De Waard, M., Liu, H., Walker, D., Scott, V. E., Gurnett, C. A., and Campbell, K. P. (1997) Direct binding of G-protein βγ complex to voltage-dependent calcium channels. *Nature* **385**, 446–450
  33. Zamponi, G. W., Bourinet, E., Nelson, D., Nargeot, J., and Snutch, T. P. (1997) Crosstalk between G proteins and protein kinase C mediated by the calcium channel α1 subunit. *Nature* **385**, 442–446
  34. Zhang, Y., Chen, Y. H., Bangaru, S. D., He, L., Abele, K., Tanabe, S., Kozasa, T., and Yang, J. (2008) Origin of the voltage dependence of G-protein regulation of P/Q-type Ca<sup>2+</sup> channels. *J. Neurosci.* **28**, 14176–14188
  35. Hartl, F. U., Bracher, A., and Hayer-Hartl, M. (2011) Molecular chaperones in protein folding and proteostasis. *Nature* **475**, 324–332
  36. Pierce, N. W., Kleiger, G., Shan, S. O., and Deshaies, R. J. (2009) Detection of sequential polyubiquitylation on a millisecond timescale. *Nature* **462**, 615–619
  37. Thrower, J. S., Hoffman, L., Rechsteiner, M., and Pickart, C. M. (2000) Recognition of the polyubiquitin proteolytic signal. *EMBO J.* **19**, 94–102
  38. Shabek, N., Herman-Bachinsky, Y., Buchsbaum, S., Lewinson, O., Haj-Yahya, M., Hejjaoui, M., Lashuel, H. A., Sommer, T., Brik, A., and Ciechanover, A. (2012) The size of the proteasomal substrate determines whether its degradation will be mediated by mono- or polyubiquitylation. *Mol. Cell* **48**, 87–97
  39. Mukhopadhyay, D., and Riezman, H. (2007) Proteasome-independent functions of ubiquitin in endocytosis and signaling. *Science* **315**, 201–205
  40. Wagner, S. A., Beli, P., Weinert, B. T., Nielsen, M. L., Cox, J., Mann, M., and Choudhary, C. (2011) A proteome-wide, quantitative survey of *in vivo* ubiquitylation sites reveals widespread regulatory roles. *Mol. Cell. Proteomics* **10**, M111.013284
  41. Turner, G. C., and Varshavsky, A. (2000) Detecting and measuring cotranslational protein degradation *in vivo*. *Science* **289**, 2117–2120
  42. Comyn, S. A., Chan, G. T., and Mayor, T. (2014) False start: cotranslational protein ubiquitination and cytosolic protein quality control. *J. Proteomics* **100**, 92–101
  43. Ma, D., Taneja, T. K., Hagen, B. M., Kim, B. Y., Ortega, B., Lederer, W. J., and Welling, P. A. (2011) Golgi export of the Kir2.1 channel is driven by a trafficking signal located within its tertiary structure. *Cell* **145**, 1102–1115
  44. Cormack, B. P., Valdivia, R. H., and Falkow, S. (1996) FACS-optimized mutants of the green fluorescent protein (GFP). *Gene* **173**, 33–38

# The Ca<sub>v</sub>β Subunit Protects the I-II Loop of the Voltage-gated Calcium Channel Ca<sub>v</sub>2.2 from Proteasomal Degradation but Not Oligoubiquitination

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