Running title:

MiRNAs in human oocytes and blastocysts

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Investigation of microRNA expression and DNA repair gene transcripts in human oocytes and blastocysts

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Capsule Summary

Detection of DNA damage and activation of correct DNA repair mechanisms play a crucial role during preimplantation embryo development. If the repair mechanisms are unable to repair the damage, apoptosis of an embryonic cell can be detrimental to the early developing embryo. Therefore correct activation of genes and proteins is critical for the survival of a preimplantation embryo.

The level of gene expression is affected by many factors and one of the regulatory factors is through microRNA (miRNA) regulation. Several studies have shown that miRNAs regulate their target genes in cancer cells and cell lines. In this study, we have shown that the miRNAs expressed in human oocytes and blastocysts are associated with their target genes that are involved in DNA repair.

Abstract

Purpose: The aim of the study is to investigate the regulation of DNA repair genes by microRNAs (miRNAs). MiRNAs are short non-coding RNAs that regulate transcriptional and post-transcriptional gene silencing. Several miRNAs that are expressed during preimplantation embryo development have been shown or are predicted to target genes that regulate cell cycle checkpoints and DNA repair in response to DNA damage.

Methods: This study compares the expression level of 20 miRNAs and nine target transcripts involved in DNA repair. The statistical significance of differential miRNA expression between oocytes and blastocysts was determined by T-test analysis using GraphPad prism. The possible regulatory roles of miRNAs on their target mRNAs were analysed by a Pearson correlation test.

Results: This study shows for the first time that several miRNAs are expressed in human oocytes and blastocysts that target key genes involved in DNA repair and cell cycle checkpoints. Blastocysts exhibited statistically significant lower expression levels for the majority of miRNAs compared to oocytes ($p < 0.05$). Correlation analyses showed that there were both inverse and direct association between miRNAs and their target mRNAs.

Conclusions: MiRNAs target many mRNAs including ones involved in DNA repair mechanisms. This study suggests that miRNAs and their target mRNAs involved in DNA repair are expressed in preimplantation embryos. Similar to the miRNAs expressed in adult tissues, these miRNAs seems to have regulatory roles on their target DNA repair mRNAs during preimplantion embryo development.

Keywords: microRNA, expression, DNA repair, human oocyte and blastocyst, preimplantation embryo development

INTRODUCTION

Coordination of the cell cycle and activation of DNA repair mechanisms upon DNA damage are crucial in the early developing embryo to preserve the genomic integrity. If the repair mechanisms are unable to repair the damage, apoptosis of an embryonic cell can be detrimental to the early developing embryo [1]. Therefore correct activation of genes and proteins is critical for the survival of a preimplantation embryo.

The regulation of transcripts is controlled by complex mechanisms. Recent studies proposed that more than half of the human transcriptome is regulated by microRNAs (miRNAs) [2, 3]. Several miRNAs have been shown to regulate and be regulated by genes functioning at cell cycle checkpoints [4-10] and in different repair mechanisms; nucleotide excision repair [11], mismatch repair [12-15] and double strand break repair [16-21] in cancer cells and cell lines.

MiRNAs are small non-coding RNAs of 17-25 nucleotide in length which either degrade or inhibit the expression and therefore function of their target genes [22-29]. More recently studies suggested that these non-coding RNAs may stabilise the expression of their target mRNAs [27, 30-36].

Expression of miRNAs has been observed in murine, bovine and human gametes and preimplantation embryos [22, 25, 37-40]. Similar expression profiles of miRNAs in mature mouse oocytes and in the early developing embryos were observed that may be due to the maternally inherited miRNAs in the early embryo [38]. Expression of miRNAs was also detected in the sperm and about 20% of these miRNAs are present within the nuclear or perinuclear part of the sperm that are transferred to the zygote [41]. However, these miRNAs are thought to not play a significant role since these miRNAs are already present in the oocytes [42]. Expression levels of these miRNAs vary during cleavage stage divisions, such that in murine embryos when the maternally inherited miRNAs are being degraded, a 60% decrease in the miRNA expression was observed [38, 39]. In this study, we investigate the expression of specific miRNAs that are known to target mRNA transcripts associated with DNA repair in human oocytes and blastocysts to correlate the expression profiles of these mRNA and

miRNAs and to elucidate the potential regulatory role and activity of the repair mechanisms in human oocytes and preimplantation embryos.

MATERIALS and METHODS

This work was licensed by the Human Fertilisation and Embryology Authority (HFEA project reference: RO113) and ethical approval was granted by the National Research Ethics Service (NRES), Research Ethics Committee (REC reference number: 10/H0709/26).

Sample collection and processing

Immature oocytes and surplus cryopreserved blastocysts were collected from patients who had given an informed consent following *in vitro* fertilisation (IVF) treatment. The zona pellucida of oocytes and blastocysts was removed following brief exposure to acidified Tyrode's solution (Medi-Cult, Surrey, UK). The samples were then washed in phosphate buffered saline with 0.1% polyvinyl alcohol solution (PBS/PVA, Sigma, USA) and 0.3U/µl RNasin plus RNAse inhibitor (Promega, UK) to prevent RNA degradation. Single oocytes and single blastocysts were transferred in a minimum volume of PBS/PVA with RNasin solution to an empty 0.2ml MicroAmp reaction tube.

Selection of mRNA and miRNA for analysis

An expression profile of DNA repair genes in human oocytes and blastocysts was previously established using microarrays [43, 44]. This study was used as a guideline to identify mRNAs that were differentially expressed in human oocytes relative to blastocysts. In order to deduce the activity of different DNA repair pathways one sensor gene or a gene functioning at the earlier stages of the particular repair pathway and one gene functioning at later stages of the repair pathway were selected. In addition to this group of repair genes, the expression of one miRNA processing gene and one house keeping gene was analysed as control genes. MiRNAs targeting the selected DNA repair genes were identified using miRNA databases (http://www.microrna.org/microrna/home.do, http://www.targetscan.org/ and http://mirdb.org/miRDB/) and previously published articles [22, 38,

40, 43, 45, 46]. A further literature search was conducted to select from these miRNAs that were shown to be expressed in human, murine and bovine embryos. Two miRNAs, namely hsa-miR-15a and hsa-miR-212, that were previously shown to be expressed in human oocytes and two miRNAs, hsa-let-7a, hsa-miR-21, that were reported to be expressed in human blastocysts were analysed as controls. An additional 16 miRNAs expressed in murine and bovine oocytes and blastocysts and embryonic stem cells were selected from the previous publications.

Oocytes and blastocysts were grouped according to morphological grade and maternal age. For each mRNA and miRNA, expression analyses were performed in six repeat samples (six individual oocytes and six individual blastocysts) to eliminate the sample variation with at least two replicates.

mRNA and miRNA expression

mRNA expression was analysed from nine oocytes and ten blastocysts using the TaqMan® gene expression cells-to-CT™ kit (Ambion, Life Technologies, UK). MiRNA expression was analysed using the TaqMan® microRNA cells-to-CT™ kit (Ambion, Life Technologies, UK) in a further 22 oocytes and 23 blastocysts. Both mRNA and miRNA quantification involved three main steps; lysis, reverse transcription and real time quantitative PCR (qPCR). Cell lysis was performed in 25µl lysis solution with 0.25µl DNase I at room temperature (RT) for 8 minutes. The lysis was stopped by addition of the 2.5µl stop solution and incubation at RT for 2 minutes.

Reverse transcription and qPCR for both mRNA and miRNA was performed following the respective manufacturer's protocol. For each reaction, negative controls in the absence of embryo lysates and cDNA products were performed. Overall 10 mRNAs and 20 miRNAs were analysed. Real time qPCR for each sample was carried out in duplicates and the expression level of each mRNA or miRNA was determined in a minimum of six different oocytes and six different blastocysts. All the samples that were used for mRNA expression analysis were also tested for *ACTB* expression. Similarly all samples that were used for miRNA expression analysis were tested for *RNU48*. *ACTB* and RNU48 were

selected for endogenous reference genes since both were shown to be expressed at a constant level in human oocytes and preimplantation embryos.

The comparative ΔΔCq method was used to examine the expression levels of miRNAs and mRNAs [40]. Determination of Cq values was performed using the LightCycler Nano software (Roche, UK) and ΔCq values were determined as follows after normalisation with the endogenous reference gene *ACTB* for mRNA and RNU48 for miRNA, respectively, for each oocyte and blastocyst sample:

ΔCq = Cq_ target gene - Cq_*ACTB*

ΔCq=Cq_ target gene - Cq_RNU48

The fold change was analysed by the relative quantification method, 2^-ΔΔCq:

ΔΔCq = (Cq_target-Cq_*ACTB*)blastocyst - (Cq_ target-Cq_*ACTB*)oocyte

ΔΔCq=(Cq_target-Cq_RNU48)blastocyst-(Cq_ target-Cq_RNU48)oocyte

Statistical analyses

All the statistical analyses were carried out using the GraphPad prism v6 software. The level of expression of each mRNA and miRNA in oocytes relative to blastocysts was examined by applying an unpaired two-tailed student's T-test with the Welch correction, respectively. The correlation between each miRNA and its target mRNA was investigated by the Pearson correlation test. An inverse correlation was defined as r=-1 and a direct correlation as r=+1 following Pearson correlation test. Similarly, a perfect correlation was defined as Pearson's r^2 coefficient equal to 1. For all the statistical analysis, $p<0.05$ indicated a statistical significance.

RESULTS

A total of 31 immature oocytes and 33 surplus cryopreserved blastocysts were collected from twenty women (maternal age; 35±5) and sixteen couples (maternal age; 38±7), respectively. mRNA

expression of nine transcripts involved in DNA repair and one involved in miRNA processing was investigated in a total of nine oocytes and ten blastocysts. Differentially expressed DNA repair genes in human oocytes and blastocysts were identified previously [43]. A literature search to identify miRNAs expressed in human oocytes and blastocysts was performed. This search was narrowed down on the miRNAs that were shown or predicted to regulate these differentially expressed DNA repair genes. From these mRNA and miRNA profiles, repair genes such as miRNAs targeting an important cell cycle checkpoint gene, sensor genes from each pathway and genes involved in different repair pathways, were selected to be analysed in this study (Table 1 and figure 1). MiRNAs were selected to target more than one gene that is involved in different repair pathways to be able to have a general idea of the possible regulatory roles of miRNAs in DNA repair pathways. The expression of 20 miRNAs targeting these repair genes was analysed in 22 oocytes and 23 blastocysts, respectively. The expression level of each mRNA or miRNA was determined in a minimum of six different oocytes and six different blastocysts. Each analysis was carried out in duplicate.

mRNA and miRNA expression

Quantification for each mRNA and miRNA was performed after normalisation with endogenous control genes, *ACTB* and RNU48, respectively. All ten mRNAs analysed (*PARP1*, *BRCA1*, *RAD50*, *MSH2*, *MSH3*, *GTF2H2*, *ERCC3*, *DCLRE1A*, *RB1* and *DICER*) were expressed both in oocytes and blastocysts. Of the twenty miRNAs analysed, eleven (hsa-let-7a, hsa-miR-16, hsa-miR-21, hsa-miR-31, hsa-miR-101, hsa-miR-145, hsa-miR-182, hsa-miR-192, hsa-miR-194, hsa-miR-210 and hsa-miR-212) were detected in oocytes and eighteen (hsa-let-7a, hsa-miR-7-2, hsa-miR-15a, hsa-miR-16, hsa-miR-21, hsa-miR-23b, hsa-miR-31, hsa-miR-34c, hsa-miR-101, hsa-miR-128, hsa-miR-130, hsa-miR-145, hsa-miR-155, hsa-miR-182, hsa-miR-192, hsa-miR-194, hsa-miR-210 and hsa-miR-212) in blastocysts, respectively. Two miRNAs analysed (hsa-miR-181c and hsa-miR-196) were neither detected in oocytes nor in blastocysts, whereas seven were only detected in blastocysts (hsa-miR-7-2, has-miR-15a, hsa-miR-23, hsa-miR-34, hsa-miR-128, hsa-miR-130 and hsa-miR-155). All miRNAs

detected in oocytes were also detected in blastocysts. The fold differences of these mRNAs and miRNAs between oocytes and blastocysts are listed in the supplementary table 2.

The expression levels of all mRNAs were significantly higher in oocytes compared to blastocysts ($p < 0.05$, figure 2). Six miRNAs (hsa-miR- 31, hsa-miR-16, let-miR-7a, hsa- miR-145, hsa-miR-210 and hsa-miR-212) were expressed at significantly higher levels in oocytes compared to blastocysts (Figure 3) whereas five miRNAs (hsa-miR-21, hsa-miR-101, hsa-miR-182, hsa-miR-192 and hsa-miR-194) were expressed at similar levels in oocytes and blastocysts.

Correlation between the expression of mRNA and miRNA

Two miRNAs, hsa-miR-181c and hsa-miR-196, that were neither expressed in oocytes nor in blastocysts were excluded from the correlation analysis. Pearson correlation test showed that there was a trend for both negative and positive correlations between miRNAs and their target mRNAs. Inverse associations were observed in blastocysts ($p < 0.05$) for hsa-miR-23b and its target gene *GTF2H2*, hsa-miR-128 and *DCLRE1A* and hsa-miR-128 and *RAD50* and direct association for hsa-miR-34c, and *RB1* that may indicate a possible stabilisation effect of miRNAs on their target mRNAs.

DISCUSSION

The level of gene expression is affected by many factors that may influence the DNA repair capacity. In the recent years, one of the regulatory mechanisms of transcripts was suggested to be through miRNA control. Although many studies have shown that several repair gene transcripts are regulated or regulate several miRNAs in cancer cells and cell lines, no studies were performed to analyse the association between miRNAs and their target mRNAs involved in DNA repair in human oocytes and embryos. The aim of this study was to analyse this association by correlating the expression levels of a selection of miRNAs and their target mRNAs.

In this study, the expression of 10 mRNAs and 20 miRNAs were analysed in human oocytes and blastocysts. The expression of all the mRNAs tested and 11 miRNAs were detected in both oocytes and blastocysts. Higher mRNA expression levels were detected in oocytes relative to the blastocysts (Figure 2). This is not surprising since the oocytes are required to be packed with mRNAs and passed on the early embryo to support itself until the embryonic genome activation and the maternal mRNAs are expected to be degraded post embryonic genome activation. Similar to the repair genes, majority of the miRNAs showed a higher expression level in oocytes relative to blastocysts. It has been well established that miRNAs silence many genes for translational inhibition, cleavage, degradation or destabilisation [23-28]. Therefore, the higher expression of miRNAs in oocytes may be required to degrade the maternally inherited mRNAs in the early developing embryos as also reported in zebrafish [47, 48] and in rainbow trout [49].

More recently studies have proposed that miRNAs may stabilise their target mRNAs as well [27, 30-36]. This study further investigated the possibility of the stabilisation effect of mRNAs on their target mRNAs by analysing whether individual samples with higher miRNA expression levels might tend to have higher or consistent expression levels of their target mRNAs. Alternatively a correlation in the other direction was questioned in order to understand the possible down-regulatory roles of miRNAs on their targets.

A trend for both direct and inverse relationships was observed between miRNAs and their target mRNAs. It is possible that these inverse and direct relationships are miRNA specific and each miRNA regulates each target differently. It may also depend on the embryonic development stage or it is possible that these observations do not hold a biological significance. A direct association was observed between the expression of miRNAs and *RB1*, active at G1/S transition, in oocytes and blastocysts. This direct relationship was also observed in oocytes among the miRNAs and mismatch repair genes, *MSH2* and *MSH3*, functioning at G2 cell cycle checkpoint. This association may be necessary to stabilise the target mRNAs in the early developing embryos to direct DNA repair until the embryonic genome activation. However both direct and indirect relationships were observed for

the nucleotide excision repair genes as also reported previously for miR-192 and *ERCC3* [11], double strand break repair and interstrand crosslink repair genes. The down-regulatory effect of some miRNAs on *BRCA1* and *PARP1* was also reported previously [50, 51]. The inverse relationships between miRNAs and their target mRNAs functioning at the G1/S phase of cell cycle may impair the detection and repair capacity of DNA damage. This may be the reason for oocytes to continue the meiotic divisions in the presence of DNA damage as reported previously by Marangos and Carroll (2012) [52].

One of the main limitations of this study was the practical inability of following the expression of miRNA and mRNA through different developmental stages. We have analysed the expression level of miRNAs and mRNAs in immature oocytes and at blastocyst stage of embryonic development. Previously published studies in humans have shown that 2% of the miRNAs analysed showed different expression levels between GV and MII oocytes [40]. Similarly, bovine studies have shown that miRNA expression varies during preimplantation embryo development and embryonic genome activation [38, 39]. However, in our study due to the scarcity of human samples, we could only analyse the expression of a limited number of miRNA and mRNA in limited sample types. Although the investigation of the miRNA and the target mRNA expression within the same group of oocytes and blastocysts would provide a more applicable analysis, due to technical reasons and scarcity of the samples, the analysis was performed on a separate group of oocytes and blastocysts obtained from different women and couples. While use of oocytes and embryos from different women and couples is a weakness of our study, it could also be considered as strength since it eliminates any possible hierarchal conclusions. Furthermore, even though there may be some differential miRNA expression among different oocytes and embryos, the difference is not expected to be significant since none of the patients were diagnosed with complications, such as polycystic ovarian syndrome or endometriosis that may have caused differences in miRNA expression [53, 54]. With the improvements in the amplification techniques and lowering the bias introduced, future experiments may be performed on amplified human samples enabling expression studies on the same set of oocytes and blastocysts. The amplification has the benefit of analysis many more miRNAs and

mRNAs from the same set of oocytes and blastocysts. Further studies focusing on expression analysis of more miRNAs and target mRNAs will provide better understanding the association between miRNAs and their target mRNAs. However, the expression analyses alone would still not be sufficient to prove a true regulation and functional studies are crucial for a definite conclusion.

CONCLUSIONS

Regulation of mRNAs involves complex mechanisms, amongst which are included the action of miRNAs. It is well established that miRNAs target many mRNAs including ones involved in DNA repair mechanisms. This study showed that miRNAs and their target mRNAs involved in DNA repair are expressed in preimplantation embryos. Similar to the miRNAs expressed in different tissue types, the miRNAs expressed in embryos seem to have regulatory roles on their target DNA repair mRNAs during preimplantion embryo development.

REFERENCES

1. Lukas, J., C. Lukas, and J. Bartek, *Mammalian cell cycle checkpoints: signalling pathways and their organization in space and time.* DNA Repair (Amst), 2004. **3**(8-9): p. 997-1007.
2. Bartel, D.P., *MicroRNAs: target recognition and regulatory functions.* Cell, 2009. **136**(2): p. 215-33.
3. Pasquinelli, A.E., *MicroRNAs and their targets: recognition, regulation and an emerging reciprocal relationship.* Nat Rev Genet, 2012. **13**(4): p. 271-82.
4. Liu, Y., et al., *miR-375 targets the p53 gene to regulate cellular response to ionizing radiation and etoposide in gastric cancer cells.* DNA Repair (Amst), 2013. **12**(9): p. 741-50.
5. Pok, S., et al., *Cyclin E facilitates dysplastic hepatocytes to bypass G1/S checkpoint in hepatocarcinogenesis.* J Gastroenterol Hepatol, 2013. **28**(9): p. 1545-54.
6. Noto, J.M. and R.M. Peek, *The Role of microRNAs in Helicobacter pylori Pathogenesis and Gastric Carcinogenesis.* Front Cell Infect Microbiol, 2012. **1**: p. 21.
7. Aqeilan, R.I., G.A. Calin, and C.M. Croce, *miR-15a and miR-16-1 in cancer: discovery, function and future perspectives.* Cell Death Differ, 2009. **17**(2): p. 215-220.
8. Krichevsky, A.M. and G. Gabriely, *miR-21: a small multi-faceted RNA.* Journal of Cellular and Molecular Medicine, 2009. **13**(1): p. 39-53.
9. Jurado, J.O., et al., *IL-17 and IFN-gamma expression in lymphocytes from patients with active tuberculosis correlates with the severity of the disease.* J Leukoc Biol, 2012. **91**(6): p. 991-1002.
10. Massirer, K.B., et al., *The miR-35-41 family of microRNAs regulates RNAi sensitivity in Caenorhabditis elegans.* PLoS Genet, 2012. **8**(3): p. e1002536.
11. Xie, Q.H., et al., *MiR-192 inhibits nucleotide excision repair by targeting ERCC3 and ERCC4 in HepG2.2.15 cells.* Biochem Biophys Res Commun, 2011. **410**(3): p. 440-5.
12. Svrcek, M., et al., *Overexpression of microRNAs-155 and 21 targeting mismatch repair proteins in inflammatory bowel diseases.* Carcinogenesis, 2013. **34**(4): p. 828-34.
13. Mao, G., et al., *Modulation of microRNA processing by mismatch repair protein MutLalpha.* Cell Res, 2012. **22**(6): p. 973-85.
14. He, Y.Q., et al., *Estradiol regulates miR-135b and mismatch repair gene expressions via estrogen receptor-beta in colorectal cells.* Exp Mol Med, 2012. **44**(12): p. 723-32.
15. Valeri, N., et al., *Modulation of mismatch repair and genomic stability by miR-155.* Proceedings of the National Academy of Sciences, 2010. **107**(15): p. 6982-6987.
16. Hoffman, A.E., et al., *microRNA miR-196a-2 and breast cancer: a genetic and epigenetic association study and functional analysis.* Cancer Res, 2009. **69**(14): p. 5970-7.
17. Neijenhuis, S., et al., *Identification of miRNA modulators to PARP inhibitor response.* DNA Repair (Amst), 2013. **12**(6): p. 394-402.
18. Krishnan, K., et al., *MicroRNA-182-5p targets a network of genes involved in DNA repair.* RNA, 2013. **19**(2): p. 230-42.
19. Jiang, Y., et al., *Genetic variation in a hsa-let-7 binding site in RAD52 is associated with breast cancer susceptibility.* Carcinogenesis, 2013. **34**(3): p. 689-93.
20. Mansour, W.Y., et al., *Aberrant overexpression of miR-421 downregulates ATM and leads to a pronounced DSB repair defect and clinical hypersensitivity in SKX squamous cell carcinoma.* Radiother Oncol, 2012. **106**(1): p. 147-54.
21. Wu, C.W., et al., *MicroRNA-18a attenuates DNA damage repair through suppressing the expression of ataxia telangiectasia mutated in colorectal cancer.* PLoS One, 2013. **8**(2): p. e57036.
22. McCallie, B., W.B. Schoolcraft, and M.G. Katz-Jaffe, *Aberration of blastocyst microRNA expression is associated with human infertility.* Fertil Steril, 2010.

23. Bagga, S., et al., *Regulation by let-7 and lin-4 miRNAs results in target mRNA degradation.* Cell, 2005. **122**(4): p. 553-63.
24. Hayashi, K., et al., *MicroRNA biogenesis is required for mouse primordial germ cell development and spermatogenesis.* PLoS One, 2008. **3**(3): p. e1738.
25. Tesfaye, D., et al., *Identification and expression profiling of microRNAs during bovine oocyte maturation using heterologous approach.* Mol Reprod Dev, 2009. **76**(7): p. 665-77.
26. Bartel, D.P., *MicroRNAs: genomics, biogenesis, mechanism, and function.* Cell, 2004. **116**(2): p. 281-97.
27. Bartel, D.P. and C.Z. Chen, *Micromanagers of gene expression: the potentially widespread influence of metazoan microRNAs.* Nat Rev Genet, 2004. **5**(5): p. 396-400.
28. Miranda, K.C., et al., *A pattern-based method for the identification of MicroRNA binding sites and their corresponding heteroduplexes.* Cell, 2006. **126**(6): p. 1203-17.
29. Place, R.F., et al., *MicroRNA-373 induces expression of genes with complementary promoter sequences.* Proc Natl Acad Sci U S A, 2008. **105**(5): p. 1608-13.
30. Salmena, L., et al., *A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language?* Cell, 2011. **146**(3): p. 353-8.
31. Seitz, H., *Redefining microRNA targets.* Curr Biol, 2009. **19**(10): p. 870-3.
32. Arvey, A., et al., *Target mRNA abundance dilutes microRNA and siRNA activity.* Mol Syst Biol, 2010. **6**: p. 363.
33. Lee, D.Y., et al., *A 3'-untranslated region (3'UTR) induces organ adhesion by regulating miR-199a* functions.* PLoS One, 2009. **4**(2): p. e4527.
34. Poliseno, L., et al., *A coding-independent function of gene and pseudogene mRNAs regulates tumour biology.* Nature, 2010. **465**(7301): p. 1033-8.
35. Franco-Zorrilla, J.M., et al., *Target mimicry provides a new mechanism for regulation of microRNA activity.* Nat Genet, 2007. **39**(8): p. 1033-7.
36. Cazalla, D., T. Yario, and J.A. Steitz, *Down-regulation of a host microRNA by a Herpesvirus saimiri noncoding RNA.* Science, 2010. **328**(5985): p. 1563-6.
37. Rosenbluth, E.M., et al., *MicroRNA expression in the human blastocyst.* Fertil Steril, 2012. **99**(3): p. 855-861.
38. Tang, F., et al., *Maternal microRNAs are essential for mouse zygotic development.* Genes Dev, 2007. **21**(6): p. 644-8.
39. Yang, Y., et al., *Determination of microRNAs in mouse preimplantation embryos by microarray.* Dev Dyn, 2008. **237**(9): p. 2315-27.
40. Xu, Y.W., et al., *Differentially expressed micoRNAs in human oocytes.* J Assist Reprod Genet, 2011. **28**(6): p. 559-66.
41. Liu, J., et al., *MicroRNA-dependent localization of targeted mRNAs to mammalian P-bodies.* Nat Cell Biol, 2005. **7**(7): p. 719-23.
42. Amanai, M., M. Brahmajosyula, and A.C. Perry, *A restricted role for sperm-borne microRNAs in mammalian fertilization.* Biol Reprod, 2006. **75**(6): p. 877-84.
43. Jaroudi, S., et al., *Expression profiling of DNA repair genes in human oocytes and blastocysts using microarrays.* Hum Reprod, 2009. **24**(10): p. 2649-55.
44. Kakourou, G., et al., *Investigation of gene expression profiles before and after embryonic genome activation and assessment of functional pathways at the human metaphase II oocyte and blastocyst stage.* Fertil Steril, 2012. **99**(3): p. 803-814 e23.
45. Yang, G.H., et al., *MicroRNAs are involved in erythroid differentiation control.* J Cell Biochem, 2009. **107**(3): p. 548-56.
46. Mondou, E., et al., *Analysis of microRNAs and their precursors in bovine early embryonic development.* Mol Hum Reprod, 2012.
47. Chen, P.Y., et al., *The developmental miRNA profiles of zebrafish as determined by small RNA cloning.* Genes Dev, 2005. **19**(11): p. 1288-93.

48. Giraldez, A.J., et al., *Zebrafish MiR-430 promotes deadenylation and clearance of maternal mRNAs.* Science, 2006. **312**(5770): p. 75-9.
49. Ramachandra, R.K., et al., *Cloning and characterization of microRNAs from rainbow trout (Oncorhynchus mykiss): their expression during early embryonic development.* BMC Dev Biol, 2008. **8**: p. 41.
50. Moskwa, P., et al., *miR-182-Mediated Downregulation of BRCA1 Impacts DNA Repair and Sensitivity to PARP Inhibitors.* Molecular cell, 2010. **41**(2): p. 210-220.
51. Volinia, S., et al., *Breast cancer signatures for invasiveness and prognosis defined by deep sequencing of microRNA.* Proc Natl Acad Sci U S A, 2012. **109**(8): p. 3024-9.
52. Marangos, P. and J. Carroll, *Oocytes progress beyond prophase in the presence of DNA damage.* Curr Biol, 2012. **22**(11): p. 989-94.
53. Sorensen, A.E., et al., *MicroRNAs Related to Polycystic Ovary Syndrome (PCOS).* Genes (Basel), 2014. **5**(3): p. 684-708.
54. Yang, L. and H.Y. Liu, *Small RNA molecules in endometriosis: pathogenesis and therapeutic aspects.* Eur J Obstet Gynecol Reprod Biol, 2014. **183**: p. 83-8.
55. McPhee, F., et al., *Resistance Analysis of the Hepatitis C Virus NS3 Protease Inhibitor Asunaprevir.* Antimicrob Agents Chemother, 2012.
56. Valeria, N., et al., *MicroRNA-21 induces resistance to 5-fluorouracil by down-regulating human DNA MutS homolog 2 (hMSH2).* PNAS, 2010. **107**: p. 21098-21103.
57. Lee, R.C., R.L. Feinbaum, and V. Ambros, *The C. elegans heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14.* Cell, 1993. **75**(5): p. 843-54.
58. Mueller, A.C., D. Sun, and A. Dutta, *The miR-99 family regulates the DNA damage response through its target SNF2H.* Oncogene, 2012.

FIGURE LEGENDS

Figure 1Schematic diagram of the complex association of miRNAs in DNA repair pathways.

MiRNAs that were shown or predicted to target genes involved in repair were shown in a target map. MiRNAs that were associated with microsatellite instability (indicative of defective mismatch repair) were also shown in this diagram. MiRNAs were colour coded according to their target mRNAs. Double strand break repair genes (*DCLRE1A*, *PARP1*, *BRCA1* and *RAD50*) are shown in pink, mismatch repair genes (*MSH2* and *MSH3*) and microsatellite instability in green, nucleotide excision repair genes (*ERCC3* and *GTF2H2*) in purple and checkpoint genes (*RB1* and) in blue shades, respectively.

Figure 2 mRNA expression levels quantified by real time PCR after normalisation with the endogenous control, *ACTB*.

Quantification of target mRNAs compared to *GTF2H2*, which was shown to be expressed at the highest level in a) oocytes and b) blastocysts. ANOVA followed by Dunnett's post test was applied to all the mRNA normalised to *ACTB*; where ****$p<0.0001$ indicates the most statistical significance, ***$0.0001<p<0.001$, **$0.001<p<0.01$, *$0.01<p<0.05$ less statistical significance and $p\geq 0.05$ indicates no statistical significance. The x-axis represents all the miRNAs analysed in this study and the y-axis represents the ΔCq values. *GTF2H2* is shown in purple, *PARP1* in red, *BRCA1* in pink, *RAD50* in light pink, *MSH2* in light green, *MSH3* in dark green, *ERCC3* in purple, *DCLRE1A* in mid pink, *RB1* in aqua, and *DICER1* in black shades, respectively.

Figure 3 miRNA expression levels quantified by real time PCR after normalisation with the endogenous control, RNU48.

Quantification of target miRNAs compared to hsa-miR-16 a) in oocytes and b) in blastocysts. ANOVA followed by Dunnett's post test was applied to all miRNAs compared to hsa-miR-16. The x-axis represents all the miRNAs analysed in this study and the y-axis represents the ΔCq values. In oocytes the expression of hsa-miR-16 was at a higher expression level compared to the endogenous control RNU48 represented by a negative profile for this miRNA. MiRNAs were colour-coded according to their target genes as in figure 2, i.e. miRNAs targeting *PARP1* is shown in red, *BRCA1* in pink, *RAD50* in mid pink, *DCLRE1A* in light pink, *MSH2* in light green, *MSH3* in dark green, *ERCC3* in purple, *GTF2H2* in lilac and *RB1* in aqua shades, respectively. **** represents that the difference in level of expression of the miRNA from that of hsa-miR-16 is statistically significant ($p<0.001$).

Tables

Table 1 List of miRNAs and their association with DNA repair.

Cell Cycle Checkpoint	Nucleotide Excision Repair		Base Excision Repair	Double Strand Break Repair				Mismatch Repair		
RB1	***GTF2H2***	***ERCC3***	***PARP1***	***DCLRE1A***	***PARP1***	***BRCA1***	***RAD50***	***MSH2***	***MSH3***	***MSI***
Let-7	miR-23b	miR-192	miR-7	miR-15a	miR-7	miR-7	miR-15a	miR-21	miR-7	Let-7a
miR-7	miR-101		miR-31	miR-16	miR-31	miR-145	miR-16	miR-145	miR-21	miR-31
miR-34c	miR-128		miR-130	miR-23b	miR-130	miR-155	miR-128	miR-155	miR-192	miR-101
miR-101	miR-181c		miR-182	miR-128	miR-182	miR-182	miR-155	miR-192		miR-145
miR-128	miR-192		miR-192	miR-145	miR-192	miR-196	miR-194			miR-155
miR-181c	miR-212		miR-196	miR-155	miR-196	miR-210	miR-212			miR-181c
miR-192			miR-210	miR-212	miR-210	miR-212				miR-192
miR-194										miR-196b
miR-212										miR-212

Genes involved in cell cycle checkpoint, nucleotide excision, base excision, double strand break and mismatch repair pathways and the miRNAs targeting these genes are listed. These associations were either published previously [11, 55-58] or bioinformatics studies showed that these miRNAs target the mRNAs (http://www.targetscan.org/, http://www.microrna.org/microrna/home.do, http://mirdb.org/miRDB/).

SUPPLEMENTAL TABLE LEGENDS

Supplemental Table 1 ΔCq values and fold changes for mRNA expression in blastocysts compared with oocytes.

List of a) mRNAs and b) miRNAs, fold difference and log2 of fold change in blastocyst compared with oocyte are shown. mRNAs and miRNAs and ΔCq values in oocytes and blastocysts are listed. Average normalised ΔCq values generated from six repeats with two replicates of each sample is shown. Standard deviation (SD) for each data point of ΔCq is shown in parenthesis. The greater Cq values reflect lower expression. b) MiRNAs, fold change difference and log2 of fold change in blastocyst compared with oocyte are listed. Fold change was calculated by relative quantification method (2^ΔΔCq). The log2 scale transformation of the fold change facilitates interpretation of the difference in expression between the two samples. The negative log2 values indicated increased expression of the gene in the blastocyst compared with the oocyte. N/A stands for not applicable.