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**Primate brains, the ‘island rule’ and the evolution of *Homo floresiensis***

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18 **Summary**

19 The taxonomic status of the small bodied hominin, *Homo floresiensis*, remains controversial.  
20 One contentious aspect of the debate concerns the small brain size estimated for specimen LB1  
21 (Liang Bua 1). Based on intraspecific mammalian allometric relationships between brain and  
22 body size it has been argued that the brain of LB1 is too small for its body mass and is therefore  
23 likely to be pathological. The relevance and general applicability of these scaling rules has,  
24 however, been challenged, and it is not known whether highly encephalised primates adapt to  
25 insular habitats in a consistent manner. Here, an analysis of brain and body evolution in seven  
26 extant insular primates reveals that although insular primates follow the ‘island rule’, having  
27 consistently reduced body masses compared to their mainland relatives, neither brain mass or  
28 relative brain size follow similar patterns, contrary to expectations that energetic constraints will  
29 favour decreased relative brain size. Brain:body scaling relationships previously used to assess  
30 the plausibility of dwarfism in *H. floresiensis* tend to underestimate body masses of insular  
31 primates. In contrast, under a number of phylogenetic scenarios, the evolution of brain and body  
32 mass in *H. floresiensis* is consistent with patterns observed in other insular primates.

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## 39 Introduction

40 The remains of a small brained, small-bodied hominin from the Indonesian island of Flores  
41 (Brown et al., 2004) are still cause for much debate. Ascribed to a new species within our own  
42 genus, *Homo floresiensis*, the findings raise questions about the timings of early human  
43 migrations, how early human species co-existed, and the cognitive significance of brain size  
44 (Brown et al., 2004; Falk et al., 2005; Aiello, 2009; Wood, 2011). However, their status as a new  
45 species remains controversial with several studies reporting to show similarities between *H.*  
46 *floresiensis* and various pathological disorders in modern humans (Jacob et al., 2006; Martin et  
47 al., 2006; Hershkovitz et al., 2008; Oxnard et al., 2010). These suggestions have been  
48 consistently refuted (e.g. Martinez and Hamsici, 2008; Falk et al., 2009a, b; Brown, 2012) and no  
49 proposed pathology encompasses the full range of phenotypes observed in *H. floresiensis*, or  
50 satisfactorily explains how a diseased population could persist for over 50,000 years (Brown et  
51 al., 2004; Morwood et al., 2005). In a recent review of these arguments Aiello (2009) concluded  
52 that, although the debate can only be settled with the discovery of new specimens, the current  
53 level of evidence supporting a pathological explanation is not convincing. If not pathological,  
54 the small brain and body size of *H. floresiensis* requires an evolutionary explanation.

55 Two evolutionary hypotheses have been proposed to explain the origins of *H.*  
56 *floresiensis*. Either the small body (16-41kg; (Brown et al., 2004; Aiello, 2009) and small brain  
57 (417cc, Falk et al., 2005; 426cc, Kubo et al., 2013) of *H. floresiensis* is a product of insular  
58 dwarfism from a larger bodied ancestor, perhaps *H. erectus* (Brown et al., 2004; Kubo et al.,  
59 2013), or *H. floresiensis* is a descendent of an earlier, small bodied hominin that left Africa  
60 before *H. erectus* (Brown et al., 2004; Brown and Maeda, 2009). The latter hypothesis is  
61 controversial as the long supported ‘Out of Africa 1’ model posits that *H. ergaster/erectus* were

62 the earliest hominins to leave Africa (Wood, 2011). This model has, however, been challenged  
63 by recent paleontological discoveries (Dennell and Roebroeks, 2005; Ferring et al., 2011) and a  
64 number of morphometric and cladistic analyses have suggested *H. floresiensis* bears most  
65 similarity to early African hominins, such as *H. habilis*, or the hominins discovered at Dmanisi,  
66 Georgia (Tocheri et al., 2007; Gordon et al., 2008; Argue et al., 2009; Baab and McNulty, 2009;  
67 Brown and Maeda, 2009), although not all studies agree with some providing evidence for  
68 morphological affinities with early Javanese *H. floresiensis* (Kaifu et al., 2011).

69         Although descent from an unknown, similarly sized hominin remains possible, the insular  
70 dwarfism hypothesis attracted the most attention immediately after the description of the remains  
71 (Martin et al., 2006; Falk et al., 2006; Bromham and Cardillo, 2007) and continues to be  
72 discussed in both the academic and popular press (e.g. Weston and Lister, 2009; Baab, 2012;  
73 Kubo et al., 2013). This is in spite of the generality of the ‘island rule’ being strongly disputed  
74 (Meiri et al., 2006, 2008, 2011). The ‘island rule’ suggests that large vertebrates generally  
75 experience a reduction in body size on islands due, perhaps, to energetic constraints or changes  
76 in predation rates (Foster, 1964; van Valen, 1973). Dwarfism on islands is not a general trend  
77 found across mammals but has occurred in a limited number of groups (Meiri et al., 2006, 2008),  
78 dependent upon the species’ ecology and evolutionary history (McClain et al., 2013). One order  
79 where the island rule may hold is primates (Bromham and Cardillo, 2007; Welch, 2009), support  
80 for the island rule is found using inter-specific datasets (Bromham and Cardillo, 2007; Welch,  
81 2009), but not with intra-specific datasets (Meiri et al., 2008; Schillaci et al., 2009) raising the  
82 possibility that dwarfism in primates develops over longer time frames (Meiri et al., 2008).

83         Which one of the two evolutionary hypotheses is correct has implications for interpreting  
84 the small brain size of *H. floresiensis*. Brain:body allometry between closely related species

85 closely follows intra-specific scaling relationships (Lande, 1979). Hence, if *H. floresiensis*  
86 descended from a small-bodied hominin it is reasonable to expect that its brain size would be  
87 predictable based on the degree of body size change and intraspecific allometry. However,  
88 several notable examples suggest that during episodes of insular dwarfism selection can  
89 dramatically reduce brain size in a non-allometric manner, both in absolute mass and relative to  
90 body size (Roth, 1992; Köhler and Moyà-Solà, 2004; Weston and Lister, 2009). One suggested  
91 explanation for this has been that in an environment with limited resources, energetically  
92 expensive tissues, such as the brain (Aiello and Wheeler, 1995), are decreased in size in order to  
93 balance energy expenditure (Köhler and Moyà-Solà, 2004). Hence, if *H. floresiensis* evolved  
94 from a larger bodied ancestor in a resource-limited environment, additional selective pressures  
95 may have driven the evolution of its small brain.

96         However, using both intraspecific scaling relationships between brain and body mass in  
97 humans and other mammals, and an example of insular dwarfism, Martin et al. (2006) argued  
98 that the estimated brain size for LB1 (*H. floresiensis*) is too small for its body mass. They  
99 suggest this departure from expected patterns of brain:body allometry points towards a  
100 pathological origin for the specimens (Jacob et al., 2006; Martin et al., 2006). The relevance of  
101 these scaling relationships has, however, been questioned (Falk et al., 2006; Niven, 2007; Kaifu  
102 et al., 2011) and some alternative scaling relationships are more accommodating of *H.*  
103 *floresiensis*' small brain (Weston and Lister, 2009; Montgomery et al., 2010). Weston and Lister  
104 (2009), for example, showed that the predicted decrease in *H. floresiensis*' brain size during  
105 descent from African *H. erectus* is within the range observed in dwarfed Hippos, whilst  
106 Montgomery et al. (2010) found it was within the range observed in primate genera in which  
107 body mass decreased, if *H. floresiensis* was a descendent of either *H. habilis* or Dmanisi

108 hominins. These studies show that major phenotypic changes in brain size can occur during  
109 episodes of dwarfism. However, comparisons with *H. floresiensis* are not entirely straight  
110 forward. Comparing *H. floresiensis* with particular non-primate dwarfs assumes conservation in  
111 the genetic, developmental, physiological and behavioural constraints acting on brain size across  
112 large phylogenetic distances. The use of these examples to assert that extensive dwarfism and  
113 brain reduction is ‘mechanistically possible’ (Weston and Lister, 2009; Kubo et al., 2013) in  
114 hominins therefore has limitations. Likewise, comparing intergeneric patterns in non-insular  
115 primates assumes conservation in allometry between taxonomic scales and ecological niches.

116         Unfortunately our expectations of patterns of brain evolution on islands are based on only  
117 a handful of examples, and it remains unclear if there is a consistent pattern among insular  
118 dwarfs. Assessing whether or not *H. floresiensis* departs from expected evolutionary patterns of  
119 brain:body allometry relies on choosing taxa with which to compare the evolution of *H.*  
120 *floresiensis* to in a biologically meaningful way. In this respect a comparison with other insular  
121 primates is of direct relevance (Bromham and Cardillo, 2007). If primates follow the island rule  
122 for body mass (Bromham and Cardillo, 2007; Welch, 2009) examining how brain size evolved in  
123 these insular species arguably provides the best reference for contextualizing the small brain of  
124 *H. floresiensis* and assessing the arguments put forward against its taxonomic status, and  
125 different evolutionary hypotheses.

126         Based on a thorough analysis of 7 mainland/island pairs of extant primates, the present  
127 study examines patterns of brain evolution in insular primates and re-analyses predicted patterns  
128 of brain evolution during the origin of *H. floresiensis*. I first test whether or not there is an ‘island  
129 rule’ for brain size and then examine whether the patterns of brain:body allometry in extant  
130 insular primates are in line with scaling models previously used to assess the plausibility of the

131 dwarfism hypothesis for *H. floresiensis*. Finally I examine predicted patterns of brain evolution  
132 during the descent of *H. floresiensis* using both the observed allometric scaling among extant  
133 insular primates and typical intra-specific mammalian scaling. By doing so I aim to identify the  
134 phylogenetic scenarios under which the brain and body size of *H. floresiensis* is acceptable under  
135 either, or both, the hypotheses that it descended through insular dwarfism, or from a similarly  
136 sized hominin. These results are discussed in the context of morphological similarities between  
137 *H. floresiensis* and other hominins.

138

## 139 **Materials and methods**

### 140 *Mainland/Island pairs*

141 This study makes use of the dataset of mainland/island taxon pairs compiled by Bromham and  
142 Cardillo (2007) that was used to test the island rule for primate body size. Endocranial volume  
143 (ECV) and body mass data from wild individuals are available for 7 mainland/island primate  
144 pairs; all are catarrhines (Table 1, Isler et al., 2008). Although ECV is an indirect measure of  
145 brain size it is a more readily measurable trait and scales isometrically with brain mass (Isler et  
146 al., 2008). Data on cranial capacities were converted to brain mass by multiplication by the  
147 density of fresh brain tissue (1.036 g/cc) (Isler et al., 2008). Of the 14 species included in the  
148 analysis 3 were classed as ‘data deficient’ by the original authors (Isler et al., 2008) and are based  
149 on less than 3 individuals. Whilst poor sampling of intraspecific variation may introduce error in  
150 these cases, the data are included in the analysis in order to maximize an already small sample  
151 size. Comparisons involving these species are not outliers and the conclusions are robust to their  
152 exclusion.

153 Isler et al.'s data on species means are used throughout, but analyses using data from  
154 each sex separately are also presented in the Supplementary Material and lead to the same  
155 conclusions. In addition to low intraspecific sampling, variation in the sex ratios of the data and  
156 the geographic distribution of the mainland species could introduce error into the analyses.  
157 However, I assume this error affects both brain and body mass equally and would not bias the  
158 results towards finding differences in the patterns of evolution between the two traits, or bias  
159 estimates of brain:body scaling in any particular direction. Isler et al. (2008) suggest sample size  
160 has a more important effect on parameter estimation than data quality so it is likely the size of  
161 the dataset is the most limiting factor of this study.

162 Identifying the true ancestor of insular populations/species is challenging, and a potential  
163 source of error in this study. For their study on primate body size Bromham and Cardillo selected  
164 the closest mainland relative of each island population by taking into account published  
165 phylogenies, taxonomies, species distribution data and consultation with experts. The divergence  
166 times between most island/mainland pairs are unknown, however Bromham and Cardillo suggest  
167 most of the islands in their dataset became separated from the mainland after the last glacial  
168 maximum (<12,000 years ago) although divergence date estimates between species pairs, where  
169 available, are typically older (Chatterjee et al., 2009; Perelman et al., 2011).

170 Recently several new species-level phylogenies have been published for primates based  
171 on either a supermatrix approach (Bininda-Emonds et al., 2007; Chatterjee et al., 2009; Arnold et  
172 al., 2010) or new, larger datasets that are likely to give more accurate phylogenetic  
173 reconstructions but contain fewer species (Perelman et al., 2011). Unfortunately the relationships  
174 among species, and occasionally genera, in these phylogenies do not always agree and none of  
175 these phylogenies include all 14 taxa considered here. Discerning the species level relationships

176 within some genera of catarrhines is particularly troublesome given an apparently complex  
177 pattern of hybridization and gene flow between species (Chatterjee et al., 2009). Hence, whilst  
178 phylogenetics may ultimately confirm or revise which extant species is most closely related to  
179 each insular population, which may in turn prompt a revision of the analyses presented below,  
180 for the present study Bromham and Cardillo's mainland/island pairs are adopted as no new and  
181 robust evidence is available to support alternative pairs.

182

### 183 *Tests of the island rule*

184         Although it is generally assumed that brain size decreases on islands, this hypothesis has  
185 not been well tested (but see Mace and Eisenberg, 1982 for a test within *Peromyscus* mice).  
186 Although the dataset is limited, performing explicit tests of this hypothesis is preferable to  
187 assuming it is true. For the current analyses I take a broad view of an 'island rule' for brain size  
188 and simply search for consistent patterns among insular primates. If insularity has a consistent  
189 effect on brain size several patterns could be observed. First, brain mass could decrease  
190 coincidentally with body mass such that relative brain size remains constant. In this case, brain  
191 size would account for the same proportion of the species' energetic budget (Aiello and Wheeler,  
192 1995). Alternatively, if energetically constrained, brain mass may decrease more rapidly than  
193 predicted by allometric scaling leading to a decrease in relative brain size. Finally, if brain mass  
194 remains relatively constant, perhaps due to cognitive or behavioural constraints (Deaner et al.,  
195 2007), but body mass decreases, relative brain size will increase.

196         There is considerable debate over how best to test the island rule (Lomolino, 1985;  
197 Bromham and Cardillo, 2007; Price and Phillimore, 2007; Meiri et al., 2008; Welch, 2009).

198 Welch (2009) reviewed the three main alternative tests of the island rule, their assumptions and  
199 performance. Here I provide a brief overview of his conclusions.

200 1) 'Null hypothesis A' states that evolutionary size changes on islands do not depend on  
201 the ancestral state. This hypothesis is tested using an Ordinary Least Squares (OLS) regression  
202 between the island-mainland size ratio,  $R$  ( $R = S_i/S_m$ ), and the size of the mainland species ( $S_m$ ).  
203 The null hypothesis is rejected when the slope is significantly negative (Lomolino, 1985).

204 2) 'Null hypothesis B' states that there are no differences between the patterns of  
205 evolution of mainland and island species, and mainland relatives remain close to the ancestral  
206 state. This is tested using a Standardized Major Axis (SMA) regression between the size of the  
207 island species ( $S_i$ ) and the size of the mainland species ( $S_m$ ) (Price and Phillimore, 2007) where a  
208 slope that is significantly less than one is consistent with the island rule. Welch (2007) proposed  
209 an alternative permutation test for Null hypothesis B that can be implemented by generating all  
210 possible combinations of the data where the island and mainland species are assigned randomly  
211 within each pair and repeating the regression on these data to generate a null distribution. The p-  
212 value is the proportion of tests with lower significance than the real data.

213 3) 'Null hypothesis C', states that insular evolution is characterized by a directional  
214 change in size determined by a heritable trait. This is tested using phylogenetically controlled  
215 regression between  $R$  and  $\ln(S_m)$  (Meiri et al., 2008).

216 Using simulated datasets Welch found that the test of Null hypothesis A has an  
217 unacceptable false positive rate when one of the other null models held and therefore  
218 recommended against using this test. The tests of Null B and C were found to have acceptable  
219 false positive rates and are recommended as tests of the island rule (Welch, 2009). I test the

220 island rule for body mass, brain mass and the encephalization quotient (EQ) (Jerison, 1973), a  
221 commonly used measure of relative brain size. The island rule has previously been shown to  
222 apply to body mass using the full Bromham and Cardillo dataset and different measures of body  
223 size (Bromham and Cardillo, 2007; Welch, 2009). The test is repeated here to confirm the pattern  
224 is detectable using only the 7 pairs for which brain mass data is available. Tests of Null A and B  
225 were performed using the SMATR package (Warton and Ormerod, 2007). The test for Null C  
226 was performed using Phylogenetic Generalized Least Squares (PGLS) regression between  $R$  and  
227  $S_m$  as implemented in BayesTraits (Pagel, 1999). For this test it is necessary to include a  
228 phylogeny of taxon pairs (i.e. the relationships between mainland species). Following previous  
229 test of the island rule in primates I used the mammal supertree (Bininda-Emonds et al., 2007),  
230 but also accounted for phylogenetic uncertainty by repeating the regressions across a distribution  
231 of the 1000 most supported phylogenies from Arnold et al. (2010). In the latter not all mainland  
232 species were present in the phylogeny, so in some cases con-generics were used to generate the  
233 phylogeny. Note divergence date estimates between mainland/island pairs are not required for  
234 this analysis because the regression is between the mass ratio of the species pair and the  
235 mainland mass.

236

### 237 *Scaling exponents and insular dwarfism in primates*

238 In previous studies of *H. floresiensis* the scaling relationships between brain and body  
239 mass between mainland ancestors and insular dwarfs have been used to model expected degrees  
240 of body mass reduction in *H. floresiensis* given the estimated difference in brain size between *H.*  
241 *floresiensis* and a range of putative ancestors (Martin et al., 2006; Montgomery et al., 2010;

242 Weston and Lister, 2009). These models were applied to *H. floresiensis* to assess whether it  
243 departs from expected patterns of brain:body allometry during insular dwarfism but no  
244 consideration has been given to whether or not these models accurately predict patterns of  
245 evolution in other insular dwarfs. Here, I assess how well these models are able to predict  
246 observed patterns of body mass reduction in extant insular primates given the observed  
247 difference in brain mass between the mainland and island taxa.

248         The following scaling exponents (b) were used to estimate the expected body mass of  
249 each insular species given the observed decrease in body mass: Martin et al.'s (2006) Model A  
250 (*Elphas antiquus* to *E. falconeri*;  $b = 0.32-0.35$ ) and Weston and Lister's (2010) 'late ontogenetic  
251 scaling' ( $b = 0.35$ ) and 'ontogenetic scaling' ( $b = 0.47$ ) models calculated from dwarfed, insular  
252 hippopotami. Dwarfed hippos have smaller brains than predicted based on these models with  
253 observed brain sizes up to 30% smaller than predicted by the 'late ontogenetic scaling' model  
254 and up to 17% smaller than the 'ontogenetic scaling' model (2010). For these models, the  
255 expected body mass was therefore also calculated within these bounds and are referred to as the  
256 'late ontogenetic-plus scaling' and 'ontogenetic-plus scaling'. In many cases this results in the  
257 island brain mass exceeding the mainland brain mass suggesting this model is not applicable to  
258 some insular primates which show more modest differences in brain size, I only show the results  
259 where this is not the case.

260

### 261 *Intraspecific scaling components*

262 In addition to scaling components from known cases of insular dwarfism, the brain and body  
263 mass of *H. floresiensis* has also been assessed using intraspecific scaling components. The logic

264 of applying these models to cases of insular dwarfism derives from Lande's (1979) observation  
265 that the allometric scaling relationship between brain and body mass is the same within  
266 populations as between closely related species. If insular dwarfism is the product of phyletic  
267 evolution within populations they may therefore follow intraspecific scaling (see Weston and  
268 Lister, 2009 for further discussion). Across mammals brain mass typically scales with body mass  
269 allometrically with a slope of between 0.2-0.4 (Gould, 1975; Lande, 1979). Among non-human  
270 primates a similar range is observed, from 0.124-0.349 (Holloway, 1980), and Pilbeam and  
271 Gould (1974) derived a similar value (0.329) among australopithecines.

272         Martin et al. (2006) applied a 'typical mammalian' intraspecific model (Martin et al.'s  
273 Model B) with a scaling component of 0.25 to *H. floresiensis*, and three values of human  
274 intraspecific scaling components (Model C) for males (0.1), females (0.03) and both sexes  
275 combined (0.17). As noted by Kubo et al. (2013) these values are derived from a modern Danish  
276 sample (Holloway, 1980) and may not represent global patterns, although other regionally  
277 restricted datasets produce a similar range of scaling components (Peters et al., 1998).

278         I apply similar models here, but interpret them in a slightly different way. Intraspecific  
279 scaling may poorly predict patterns of brain:body allometry in insular dwarfs (Weston and Lister,  
280 2009). Variation in life history and development is a potential explanatory factor if this causes a  
281 grade-shift between ancestor and descendent populations (Weston and Lister, 2009). However,  
282 another possible source of discrepancy is identified in Lande's original analysis of brain:body  
283 allometry (1979). Lande demonstrated that a scaling component of 0.2-0.4 is expected if the  
284 population evolved according to random genetic drift or directional selection acting only on body  
285 size. As discussed in the introduction, during episodes of insular dwarfism selection may not act  
286 solely on body size but may also have consequences for brain size and other traits. Directional

287 selection on both brain and body size could potentially lead to steeper allometric slopes (Lande,  
288 1979). Hence, for the current purposes, intraspecific scaling components can effectively form a  
289 null model which excludes the action of ‘special’ selective forces acting on both brain and body  
290 mass.

291 Kubo et al. (2013) recently used an indirect measure of body mass, femoral head  
292 diameter (FHD), to quantify brain:body scaling in humans and assess whether *H. floresiensis*  
293 scales in a similar manner. Their data consists of geographically disperse populations, includes  
294 both modern and ancient populations, and a number of modern human pygmy populations. For  
295 completeness, in the Supplementary Material I derive body mass estimates from the FHD data to  
296 discuss how their scaling relationships compare to other intraspecific datasets and the results  
297 presented below. However, these results should be viewed with caution as converting the FHD  
298 dataset to body mass may introduce a bias.

299

### 300 *Assessing H. floresiensis, insular dwarfism and intra-specific scaling*

301 The approach taken in the present study is to look for general patterns of brain evolution  
302 in insular primates, to calculate a range of brain:body scaling components observed among  
303 insular primates, and to use these to assess the evolution of *H. floresiensis* rather than to focus on  
304 specific case studies (e.g. Köhler and Moyà-Solà, 2004; Martin et al., 2006; Weston and Lister,  
305 2009). I argue this is preferable as there is a potential for taxon specific responses to selection  
306 pressures experienced on islands, and variation in what those selection pressure may be could  
307 cloud the interpretation of studies comparing *H. floresiensis* to one species/population, insular or  
308 otherwise. By focusing on insular primates the response to island ecology is incorporated with

309 allometric effects in the comparison between the mainland and island taxa, it is therefore not  
310 necessary to invoke additional amounts of change observed in non-primate insular dwarfs in  
311 addition to observed changes (Weston & Lister, 2009; Kubo et al., 2013).

312 In the first set of analyses patterns of brain evolution in insular primates are compared to  
313 that of *H. floresiensis*, assuming recent common ancestry with one of a number of hominins  
314 previously proposed as possible ancestors or closely related species, and a range of body mass  
315 estimates for *H. floresiensis*. Kubo et al. (2013) recently published a revised estimate of LB1's  
316 ECV of 426cc, and this value is used throughout. Previous estimates have ranged from 380-  
317 430cc (Brown et al., 2004; Falk et al., 2005; Holloway et al., 2011). Estimating hominin body  
318 masses is a challenging exercise (Uhl et al., 2013) and estimates of *H. floresiensis*' body mass  
319 vary greatly, from 16-41.3kg (Brown et al., 2004; Aiello, 2009) but, unless otherwise stated, in  
320 the following analyses body mass is not assumed, being the trait estimated in the analyses.  
321 Where it is assumed a range of masses are used.

322 The phylogenetic affinities of *H. floresiensis* continue to be debated. Initially it was  
323 thought possible that *H. floresiensis* was a descendent of *H. erectus* (Brown et al., 2004) and  
324 some still favour this hypothesis (e.g. Lieberman, 2009; Kaifu et al., 2011; Kubo et al., 2013).  
325 However, several subsequent analyses of morphological similarity increasingly point towards a  
326 relationship with early *Homo* species (Morwood et al., 2005; Tocheri et al., 2007; Gordon et al.,  
327 2008; Argue et al., 2009). Here I assume a range of proposed ancestors in order to identify  
328 phylogenetic scenarios under which the brain:body scaling of LB1 is plausible:

- 329 • *H. erectus*: following Martin et al. (2006) I include *H. erectus* 'broadly defined' as well as  
330 the geographically localized Ngandong hominins from Java, and Dmanisi hominins from

331 Georgia which are (tentatively) assigned to *H. erectus*, but a primitive form (Vekua et al.,  
332 2002; Wood, 2011). I also include *H. ergaster*, a candidate African ancestor of *H. erectus*  
333 (Wood and Collard, 1999). Unfortunately limited post-cranial material are available from  
334 which to estimate body mass estimates for Javan *H. erectus*, including the Ngandong  
335 specimens. The body mass estimates are therefore uncertain and should be viewed with some  
336 caution. They are included here to allow direct comparison with previous studies (Martin et  
337 al., 2006; Montgomery et al., 2010; Kubo et al., 2013).

338 • *H. habilis* and *H. rudolfensis*: a cladistic analysis by Argue et al. (2009) suggested *H.*  
339 *floresiensis* was an early member of the genus *Homo* which emerged after *H. rudolfensis* and  
340 either before or after *H. habilis*. LB1 also shows some morphological similarities to *H.*  
341 *habilis* (Gordon et al., 2008; Martinez and Hamsici, 2008).

342 • *Australopithecus africanus* and *A. sediba*: finally I include two representatives of the  
343 australopithecines. *A. sediba* is proposed to be a descendent or close relative of *A. africanus*  
344 (Berger et al., 2010) and these species may be closely allied with early *Homo* and are two of  
345 the younger members of the genus (Irish et al., 2013 but see also Wood and Harrison, 2011;  
346 Kimbel, 2013). Some authors have argued descent from an australopithecine may be a  
347 possible explanation for the origin of *H. floresiensis* (Brown and Maeda, 2009)

348 Body and brain mass estimates for these species were taken from published sources (Table 2).  
349 ECVs were converted to mass as described above. Of course, both traits are estimates and are not  
350 known without error and there is debate surrounding the taxonomic classification of some named  
351 species, and their relationships to one another. The analyses should therefore not be viewed as

352 giving a precise description of the ancestor but a range of phylogenetic scenarios under which  
353 the brain size of LB1 does not require special reasoning.

354 Each ancestor/descendant hominin pairing was used to calculate the expected *H.*  
355 *floresiensis* body masses, given the observed decrease in brain mass, using the ratio of change in  
356  $\text{Log}_{10}(\text{brain mass})$  and  $\text{Log}_{10}(\text{body mass})$  for each extant, insular primate mainland/island pair  
357 following Martin et al. (2006), and as described above.

358 Finally, as an alternative method of comparing *H. floresiensis* to insular primates, the  
359 relationship between  $R_{\text{brain}}$  and  $R_{\text{body}}$  for *H. floresiensis* was compared to the extant  
360 mainland/island pairs by assessing how the ratio for *H. floresiensis* fits the general primate  
361 relationship across a range of assumed body masses. Specifically, for each ancestor I calculated  
362 the  $R_{\text{brain}}$  and  $R_{\text{body}}$  for *H. floresiensis* across a range of body mass values for *H. floresiensis*  
363 taking 1kg incremental increases between 0kg to 45kg. I then assessed the leverage of these sets  
364 of *H. floresiensis* data points on the  $R_{\text{brain}}:R_{\text{body}}$  regression across the extant species pairs.  
365 Leverage was measured using Cook's Distance ( $D_i$ ) (Cook, 1977), which quantifies the effect on  
366 a regression of deleting an observation.  $D_i$  values greater than  $4/n$  indicate a data point has high  
367 leverage (Bollen and Jackman, 1990). High leverage scores could be due to deviation from the  
368 regression line or a large difference along either or both the x and y axis, i.e. either unexpected  
369 deviation in brain:body allometry (x or y) or unexpectedly large phenotypic shifts (x and y). I  
370 assume the regression for extant pairs represents the true relationship and interpret high leverage  
371 scores for *H. floresiensis* as indicating that the data point departs from the expected relationship.  
372 It is therefore possible to plot the leverage scores for a range of body masses for *H. floresiensis*  
373 to identify the range of body masses that fit the pattern observed in extant insular primates for

374 each assumed ancestor. These can then be compared to the range of estimated body masses for  
375 *H. floresiensis*.

376 The analyses described above assume *H. floresiensis* was an insular dwarf and should  
377 therefore have brain:body scaling within the range of other insular primates. As a test the  
378 hypothesis that *H. floresiensis* was not a dwarf, but a descendent of an early species of *Homo*  
379 that left Africa as a small bodied, small-brained hominin, I apply the typical mammalian (b =  
380 0.2-0.4), primate (b = 0.124-0.349) and Danish (b = 0.03-0.17) intraspecific scaling components  
381 described above.

382

## 383 **Results**

### 384 *Primate brains do not follow an 'island rule'*

385 For body mass, the SMA test of Null B is significant ( $p = 0.006$ , slope = 0.419) as is the  
386 permutation test of Null B ( $p = 0.016$ ) and the PGLS test of Null C using both the mammal  
387 supertree ( $p = 0.014$ ,  $R^2 = 0.734$ ) and when accounting for phylogenetic uncertainty using the  
388 10k Trees Project phylogenies ( $p = 0.008$ ,  $R^2 = 0.782$ ). This subset of data therefore reproduces  
389 support for the hypothesis that the island rule holds for primate body mass (Bromham and  
390 Cardillo, 2007; Welch, 2009).

391 In contrast, support for the applicability of an island rule to brain mass or EQ is not  
392 found. For brain mass the SMA test of Null B is not significant ( $p = 0.978$ , slope = 0.989) nor is  
393 the permutation test of Null B ( $p = 0.938$ ). The test of Null C is also narrowly non-significant  
394 (mammal supertree:  $p = 0.055$ ; 10K Trees:  $p = 0.056$ ). Similarly for EQ the SMA test ( $p = 0.813$ ,

395 slope = 0.966) and permutation test ( $p = 0.813$ ) of Null B and the PGLS test of Null C (mammal  
396 supertree:  $p = 0.580$ ; 10k Trees:  $p = 0.762$ ) are not significant. Indeed, even the OLS test of Null  
397 hypothesis A, which has high false positive rates (Welch, 2009), fails to find support for the  
398 applicability of the island rule to brain mass ( $p = 0.226$ ) or EQ ( $p = 0.571$ ) despite the test being  
399 significant for body mass ( $p = 0.001$ ). There is therefore no evidence for a consistent effect of  
400 insularity on primate brain size.

401

402 *Brain:body allometry in insular primates does not fit either non-primate mammalian models of*  
403 *insular dwarfism or typical intraspecific scaling components*

404 The ratio of brain and body masses between mainland/island pairs produce steeper allometric  
405 scaling relationships ( $b = 0.242-1.244$ , mean = 0.645) than would typically be predicted by  
406 intraspecific mammalian brain:body scaling ( $b = 0.2-0.4$ ). Only two pairs fall within this range,  
407 one of which is the *Macaca fascicularis fascicularis/M. f. fusca* pair in which the insular species  
408 is larger than its mainland relative. On average, all the scaling components previously applied to  
409 model *H. floresiensis* underestimate the body mass of the insular species in the extant  
410 mainland/island pairs (Table 2) with average percentage errors from the observed body mass  
411 between 12.2% and 57.9%. Weston and Lister's (2010) 'late ontogenetic scaling' model has the  
412 lowest average percentage error, however there is substantial variation in the performance of all  
413 models.

414 In some cases the average error across models is substantial, with values over 15% for  
415 4/7 pairs under the insular dwarfism models and even greater disparity under intraspecific  
416 scaling models. This suggests a lack of conservation in patterns of brain:body allometry

417 governing the evolution of primates and non-primate mammals on islands, and further refutes the  
418 expectation that insular dwarfs should show patterns of brain:body evolution within the range  
419 predicted by intraspecific allometry. The observation that these scaling exponents underestimate  
420 the body mass of *H. floresiensis* (Martin et al., 2006) is therefore an unconvincing argument  
421 against the status of *H. floresiensis* as a dwarfed hominin.

422

#### 423 *Testing the insular dwarfism hypothesis in H. floresiensis*

424 The relationships between mainland and island primate species pairs were used to  
425 provide a reinterpretation of the *H. floresiensis* data. When the brain:body scaling relationship  
426 from each species pair was used to predict the body mass of *H. floresiensis* (Figure 1, Table S1)  
427 the average body mass predicted is 12.9 kg based on descent from a Ngandong *H. erectus* like  
428 population, and 16.2kg for generalized *H. erectus*. These values lie below the probable range of  
429 body mass for *H. floresiensis* (Brown et al., 2004; Morwood et al., 2005; Aiello, 2009). In  
430 contrast descent from earlier *Homo* produces estimates within the acceptable range. Descent  
431 from Dmanisi hominins results in an average predicted body mass of 25.1kg, from *H. ergaster*  
432 19.7kg, from *H. habilis* 21.8kg and from *H. rudolfensis* 20.7kg. If *H. floresiensis* is instead a  
433 direct ancestor of an australopithecine, such as *A. africanus* or *A. sediba*, body mass estimates of  
434 32.3kg and 34.3kg are predicted respectively, close to the values predicted from LB1's femoral  
435 head diameter (Brown et al., 2004; Aiello, 2009). All of the scaling components from the 7 pairs  
436 of primates predict body masses within the acceptable range for descent from either  
437 australopithecines. Among *Homo*, the most consistent results are assuming ancestry with *H.*  
438 *habilis*, for which 6/7 pairs produce acceptable body masses. In this case the pair that doesn't is

439 *M. f. fascicularis/M. f. fusca*. Descent from either Dmanisi hominoids or *H. rudolfensis* is  
440 consistent with 5/7 pairs.

441 Across the extant species pairs the ratio of island/mainland brain mass ( $R_{\text{brain}}$ ) and body  
442 mass ( $R_{\text{body}}$ ) are significantly associated ( $p = 0.003$ ,  $R^2 = 0.858$ , Figure 2a). Assuming recent  
443 common ancestry with a range of hominins, the fit of *H. floresiensis* to this regression was  
444 assessed by calculating Cook's Distances across a range of body mass values for *H. floresiensis*,  
445 from 1kg up to 45kg (Figure 2b), just above the upper end of the range of body mass estimates  
446 derived from post-cranial morphology (Brown et al., 2004; Aiello, 2009). Cook's Distances  
447 indicate that dwarfism of Ngandong *H. erectus*, or *H. erectus* more generally, is not consistent  
448 with the relationship across primates unless *H. floresiensis* had a body mass of less than 3kg or  
449 11kg respectively (Table 4; Figure 2b), outside the range of body mass estimates for *H.*  
450 *floresiensis* (Brown et al., 2004; Aiello, 2009). Descent from *H. ergaster* requires a narrowly  
451 permissible final body mass of <19kg. But the evolution of *H. floresiensis* is most consistent  
452 with patterns of primate insular dwarfism ( $D_i < 0.5$ ) if it had a body mass above 14kg and was a  
453 dwarfed sister-lineage to Dmanisi hominins, above 8kg for *H. rudolfensis* or between 12kg and  
454 38kg for *H. habilis*. These are well within the range of body mass estimates for *H. floresiensis*  
455 (Brown et al., 2004). Similarly, descent from either *A. africanus* or *A. sediba* is acceptable if *H.*  
456 *floresiensis* had a body mass between 22-42kg or 22-43kg respectively. For the  
457 australopithecines, *H. habilis* and Dmanisi hominins the body mass at the lowest  $D_i$  values is  
458 within the acceptable range. Figure 2 also illustrates that, depending on the final body mass for  
459 *H. floresiensis*, some phylogenetic scenarios result in a proportional decrease in both brain and  
460 body size within the range observed between the extant mainland/island pairs.

461

462 *Testing the descent without dwarfism hypothesis in H. floresiensis*

463 The analyses above show that insular primates deviate from typical intraspecific patterns  
464 brain:body scaling, supporting previous evidence that insular dwarfs do not necessarily scale in  
465 this manner (Weston and Lister, 2009). This deviation, which is presumably caused by shifts in  
466 selection pressure or life history traits associated with insularity, permits the use of intraspecific  
467 brain:body allometry as a model of descent without insular dwarfism, as these effects should be  
468 absent. When the range of typical mammalian intraspecific brain:body scaling components are  
469 applied to *H. floresiensis*, as described above, the range of body mass estimates for all scenarios  
470 involving descent from a member of the genus *Homo* lie at the lower limit or below the  
471 acceptable range (Table 5). Only if *H. floresiensis* descended from an australopithecine are the  
472 body mass estimates reasonable. This pattern is even stronger if the range of intraspecific scaling  
473 within extant primate species ( $b = 0.12-0.33$ ), or within the Danish modern human population ( $b$   
474  $= 0.03-0.17$ ), is used.

475

## 476 **Discussion**

477 Despite generally being smaller than their mainland counterparts little evidence is found to  
478 support the applicability of an island rule to primate brain size using formal statistical tests.  
479 Although the dataset is limited in size it is sufficient to find strong statistical support for the  
480 island rule holding for body mass in primates, as previously reported (Bromham and Cardillo,  
481 2007; Welch, 2009). At the very least these results therefore suggests that the effects of insularity  
482 on brain and body size differ. For EQ in particular there is little hint of any consistent affect of  
483 insularity with increases being as common as decreases (Table 1). The scaling components

484 between brain and body mass vary greatly across pairs, from 0.24-1.24, this variability in  
485 response is a likely cause of the lack of an 'island rule' for relative brain size.

486         Although this conclusion may need to be revised when additional data is available, given  
487 the support for the island rule for body mass obtained using this dataset, the lack of support for  
488 the island rule for brain mass and EQ is perhaps surprising. Brain tissue is energetically  
489 expensive (Aiello, 2009) and it is expected that on islands resource limitation (Foster, 1964) will  
490 lead to an advantageous reduction in brain mass and relative brain size to reduce the total energy  
491 budget of the brain (Köhler and Moyà-Solà, 2004; Niven, 2007). This is observed in several  
492 insular dwarfs (Roth, 1992; Köhler and Moyà-Solà, 2004; Weston and Lister, 2009) and is  
493 further supported by evidence that food scarcity or seasonality in food abundance is associated  
494 with smaller relative brain sizes in strepsirhines (Taylor and Schaik, 2007; van Woerden et al.,  
495 2010). However, in primates, larger brains are associated with higher rates of innovative  
496 behaviour (Reader and Laland, 2002) and, in catarrhines, an ability to reduce the effects of food  
497 scarcity (van Woerden et al., 2012). Hence, one could hypothesize that for some insular primates  
498 the cognitive advantage of maintaining brain size on islands outweighs the energetic costs of  
499 doing so (Sol, 2009). The balance between these two selective forces is likely to depend on the  
500 environment of the island and the species' ecology and physiology (Navarrete et al., 2011;  
501 McClain et al., 2013) and it is clear from these results that the outcome is not necessarily  
502 consistent between species.

503         The range of responses observed in insular primates provides a meaningful framework  
504 within which to interpret the brain size of *H. floresiensis*. Assumptions surrounding the effects of  
505 insularity on brain size have had a major influence on the debate over the remains ascribed to *H.*  
506 *floresiensis* (Martin et al., 2006; Niven, 2007; Köhler and Moyà-Solà, 2008) despite being

507 largely untested. The poor performance of intraspecific brain:body scaling rules when applied to  
508 extant insular primates implies that the application of these scaling relationships as a test of  
509 whether or not the brain of *H. floresiensis* is too small to be non-pathological may be  
510 uninformative. Based on the observed difference in brain size and expected brain:body scaling  
511 relationship these models tend to underestimate the body mass of the insular species, just as they  
512 do for *H. floresiensis* (Martin et al., 2006). The size of these errors are similar to most, if not all,  
513 models applied to *H. floresiensis* (Martin et al., 2006). This discrepancy is caused by brain and  
514 body mass scaling with a higher scaling exponent than is generally expected (Lande, 1979). It is  
515 also notable that in many cases dwarfism models based on Hippopotamidae or *Elephas* also  
516 produce large errors, again suggesting variability in the effects on insularity on brain size in  
517 different mammalian taxa.

518         In contrast, when the scaling relationships between mainland and island primate species  
519 pairs are used to predict the body mass of *H. floresiensis* assuming recent shared ancestry with a  
520 range of hominin populations, the estimated mass is within the range predicted for *H. floresiensis*  
521 (Brown et al., 2004) under a number of scenarios (Table S1; Figure 1, 2). Estimates for the body  
522 mass of *H. floresiensis* vary and I here take the full range (16-41kg) as acceptable with the  
523 following caveat. In the original description of LB1, Brown et al. (2004) derived a body mass  
524 estimate of 16-28.7kg based on a stature of 106cm, and a mass of 36kg derived from femur  
525 cross-sectional area. Based on femoral head diameters body mass is estimated to be between  
526 31.4-41.31kg (Aiello, 2009). Brown et al. initially favored the lower end of this range, but  
527 subsequent finds have seen the consensus shift towards higher values (Morwood et al., 2005). A  
528 body mass of around 30kg therefore seems more reasonable, but the intraspecific range requires  
529 confirmation.

530 A close phylogenetic relationship with *H. erectus s.l.*, Ngandong *H. erectus* or *H.*  
531 *ergaster* would result in estimated body masses below, or at the very bottom of, the range of  
532 body masses for *H. floresiensis* (16.2kg, 12.9kg and 19.7kg respectively). In contrast, if *H.*  
533 *floresiensis* descended from an early *Homo*, represented here by *H. habilis*, *H. rudolfensis* or  
534 Dmanisi hominins, or an australopithecine-like hominin the predicted body mass is within the  
535 acceptable range. Within *Homo*, ancestry with *H. habilis* or Dmanisi hominoids produce the  
536 most consistent results with average estimated body masses for *H. floresiensis* of 21.8kg and  
537 25.1kg respectively. Similar results are obtained from the Cook's Distance analysis, which  
538 provide an approximate range of body masses for *H. floresiensis* that would fit the relationship  
539 between changes in brain and body mass observed in extant insular primates. For *H. habilis*, *H.*  
540 *rudolfensis*, Dmanisi hominins and the australopithecines this range encompasses the higher  
541 estimates of body mass derived from postcranial morphology of *H. floresiensis*. These results  
542 provide a range of scenarios under which descent with insular dwarfism can explain the patterns  
543 of brain:body allometry in *H. floresiensis*. Across a range of body masses, descent from *H.*  
544 *habilis*, Dmanisi hominins or Australopithecines would also result in proportional decreases in  
545 brain and body size within the range observed in insular primates (Figure 2).

546 Importantly, for the purposes of the present analysis, the fact that insular primates deviate  
547 from typical intraspecific scaling relationships permits the use of these models as a null  
548 hypothesis for descent without additional selection pressures such as those leading to insular  
549 dwarfing. When these intraspecific scaling components are used to model the descent of *H.*  
550 *floresiensis* the range of acceptable scenarios is much narrower. Essentially only descent from  
551 australopithecines produces body mass estimates consistent with those derived from post-cranial  
552 morphology of *H. floresiensis*, although descent from Dmanisi hominins or *H. habilis* produces a

553 range of estimates narrowly within the lowermost estimates. The convergence in results for  
554 australopithecines using intraspecific scaling relationships and the relationship among extant  
555 insular primates is due to the small differences in size between these species and *H. floresiensis*.

556 Unless *H. floresiensis* is a descendent of an australopithecine, the results suggest that that  
557 both evolutionary hypotheses regarding the origin of *H. floresiensis* may be true; that is, *H.*  
558 *floresiensis* descended from an early, small-bodied member of the genus *Homo* but its descent  
559 was characterized, at least in part, by insular dwarfism. Whilst resolution of the characteristics of  
560 the founding population is necessary to confirm that *H. floresiensis* was indeed an insular dwarf,  
561 the earliest human artifacts on Flores suggest there was sufficient time for dwarfism to occur  
562 (Bromham and Cardillo, 2007; Brumm et al., 2010) and the island fauna suggest environmental  
563 conditions may have rendered dwarfism likely and advantageous for some species (Meijer et al.,  
564 2010). Whilst equivocal, some features of *H. floresiensis* do point towards paedomorphic  
565 dwarfism (van Heteren, 2008, 2012) and patterns of brain and body size evolution in *H.*  
566 *floresiensis* are not inconsistent with other insular (this study) or otherwise dwarfed primates  
567 (Montgomery et al., 2010; Montgomery and Mundy, 2013).

568 The phylogenetic scenarios suggested by these analyses are consistent with previous  
569 reports of morphological similarities between *H. floresiensis* and early hominins. Several studies  
570 of cranial shape and morphology have suggested a close relationship between *H. floresiensis* and  
571 either Dmanisi hominins or *H. habilis* suggesting *H. floresiensis* was a descendent of an early  
572 member of the genus *Homo* (Gordon et al., 2008; Martinez and Hamsici, 2008; Baab and  
573 McNulty, 2009). Analysis of post-cranial morphology support this contention, with several  
574 studies demonstrating the retention of traits typical of early *Homo*, including wrist morphology  
575 (Tocheri et al., 2007), foot morphology (Jungers et al., 2009a), lower limb (Jungers et al., 2009b)

576 and shoulder morphology (Larson *et al.*, 2007). A cladistic analysis of cranial and postcranial  
577 characters also suggests *H. floresiensis* was most closely related to early members of the genus  
578 *Homo* (Argue *et al.*, 2009). Whether or not the immediate ancestry of *H. floresiensis* can be  
579 traced back to the australopithecines is more contentious, with some studies directly refuting this  
580 hypothesis (Gordon *et al.*, 2008; Baab and McNulty, 2009; Kaifu *et al.*, 2011) but others arguing  
581 that it remains a possibility (Brown and Maeda, 2009).

582         The earliest evidence for *Homo* outside Africa comes from specimens found at Dmanisi  
583 Gorge in Georgia, which extend back to c. 1.85 Ma and suggests primitive members of the genus  
584 radiated out of Africa earlier than previously suspected (Ferring *et al.*, 2011; Wood, 2011). The  
585 Dmanisi remains have been preliminarily attributed to early *H. erectus* but they retain many  
586 primitive characteristics and are smaller-bodied and have smaller brains than African *H. erectus*  
587 leading some to associate them with *H. habilis* or a previously unnamed species (Vekua *et al.*,  
588 2002; Rightmire *et al.*, 2006; Lordkipanidze *et al.*, 2007). The earliest evidence of *Homo* on  
589 Flores date to c. 1 Ma (Brumm *et al.*, 2010), which, assuming favorable climatic conditions,  
590 leaves a reasonable time period for dispersal assuming *H. floresiensis* is closely related to the  
591 Dmanisi hominins, and that these were the earliest non-African *Homo*. Characterizing this early  
592 out-of-Africa migration is, however, at an early stage and in need of productive new sites  
593 (Ferring *et al.*, 2011; Wood, 2011). It remains possible that the Dmanisi hominins were not the  
594 first Eurasians, and that an earlier lineage emerged from Africa and ultimately terminated with  
595 *H. floresiensis* (Brown and Maeda, 2009).

596         However, not all authors agree that *H. floresiensis* has clear taxonomic affinities with  
597 early *Homo*. Criticisms have been raised against some of the analyses (Trueman, 2009 and Kaifu  
598 *et al.*, 2011) and a reanalysis of cranial morphology using larger datasets of Indonesian *H.*

599 *erectus* which capture temporal variation in the population's morphology suggested *H.*  
600 *floresiensis* may be derived from early Javanese *H. erectus* (Kaifu et al., 2011), a possibility not  
601 rejected by previous studies (Baab and McNulty, 2009). Unfortunately the paucity of post-cranial  
602 material associated with these crania has so far limited the potential for similar comparisons with  
603 other anatomical traits, such as those indicating an affinity with early *Homo* (but see Kaifu et al.,  
604 2011 for discussion.).

605         The goal of the present study was to identify the phylogenetic scenarios, and range of  
606 body masses, under which patterns of brain and body size evolution in *H. floresiensis* does not  
607 deviate from expected patterns. A recent analysis by Kubo et al. (2013) produced a wider range  
608 of acceptable ancestors based on intraspecific scaling between brain size and femoral head  
609 diameter (FHD) in modern humans. They suggest descent from *H. habilis* can be explained  
610 solely by the allometric relationship between brain and body mass, whilst descent from early  
611 Javanese *H. erectus*, which the authors favour (Kaifu et al., 2011; Kubo et al. 2013), requires a  
612 similar decrease in brain mass beyond that predicted by allometry as observed in dwarfed Hippos  
613 (Weston and Lister, 2009). This approach takes advantage of large datasets and avoids error  
614 introduced when identifying ancestor/descendent relationships, but does not directly compare *H.*  
615 *floresiensis* with other insular populations and, as discussed in the introduction, assumes  
616 constraints on brain evolution are conserved between distantly related insular mammals.  
617 However, the difference between the analysis of Kubo et al. and the results based on intraspecific  
618 scaling in the present study must ultimately stem from differences in the estimated allometric  
619 slope between brain and body mass. Indeed, when converted from FHD to body mass Kubo et  
620 al.'s data give an intra-specific allometric slope of between 0.44-0.79, greater than typical  
621 estimates for mammals (see Supplementary Material).

622           This raises two questions; first, how robust is this high allometric slope? And second, is it  
623 human-specific? Kubo et al.'s dataset contains means of different human populations that may  
624 not be phylogenetically independent; indeed some are likely descendents of others. Failing to  
625 correct for this non-independence could bias estimates of scaling components (Harvey and Pagel,  
626 1991). The steep allometric slope therefore needs confirmation using phylogenetically controlled  
627 regressions. If confirmed, the question is then whether this high allometric slope is unique to  
628 humans and whether it can be applied to extinct hominins. The available data suggests extant  
629 primate populations scale within the range typical for other mammals (Holloway, 1980), as do  
630 australopithecines (Pilbeam and Gould, 1974). An upwards shift in intraspecific allometry within  
631 *Homo* is possible (Pilbeam and Gould, 1974), perhaps due to increased selection on brain mass,  
632 but further data and analysis are necessary to test this hypothesis.

633           Kubo et al.'s general conclusion however, that allometry can explain more of the  
634 reduction in brain size in *H. floresiensis* than previously thought, is in agreement with the current  
635 analysis. Where they have added variation in the estimated allometric slope by including global  
636 geographic human variation, here I have done so by examining allometry between insular  
637 primates and their mainland relatives. Although I reject descent from *H. erectus* as unlikely,  
638 descent from early Javanese *H. erectus* was not modeled here, as I know of no body mass  
639 estimates for this population. Given the lack of material for direct measurement Kubo et al.  
640 (2013) estimated FHD as 45-50mm based on size expectations and comparison with other  
641 hominins. Using this range, together with an ECV of 815cc (Kubo et al. 2013) as a model for the  
642 ancestral population of *H. floresiensis*, body mass estimates towards the lower end of the  
643 acceptable range for *H. floresiensis* are obtained (see Supplementary Material). I therefore

644 cannot rule out this scenario but, given the uncertainty in the body mass estimates, further data is  
645 required to make firm conclusions.

646 In summary, I find that two evolutionary scenarios are plausible explanations for the  
647 origin of *H. floresiensis* without the need to invoke an exceptional degree of phenotypic change.  
648 If a contribution from insular dwarfism is rejected, patterns of brain size evolution are only  
649 consistent with descent from an australopithecine-like ancestor. This would require a pre-*Homo*  
650 radiation out of Africa. If instead, it is accepted that *H. floresiensis* is an insular dwarf, a wider  
651 range of phylogenetic scenarios are consistent with descent from an early *Homo* species, similar  
652 in morphology to *H. habilis* or Dmanisi hominins, being most consistent with both patterns of  
653 brain size evolution, cranial (Gordon et al., 2008; Martinez and Hamsici, 2008; Baab and  
654 McNulty, 2009) and post-cranial morphological analysis (Tocheri et al., 2007; Jungers et al.,  
655 2009a, b; Larson et al., 2007). Of course, the taxonomic affinities of *H. floresiensis* are still  
656 debated (Kaifu et al., 2011) and many interesting questions about *H. floresiensis* remain  
657 unanswered (Aiello, 2009). What were the selection pressures favoring body mass reduction and  
658 how rapid was this change? What were the behavioural consequences of decreasing brain size  
659 and what neurological adaptations evolved to off-set size reduction (Falk et al., 2005, 2009)?  
660 What path out of Africa did the lineage leading to *H. floresiensis* take, and what drove this early  
661 migration (Brown and Maeda, 2009; Wood, 2011)? How much contact did this small bodied  
662 hominin have with modern humans (Aiello, 2009)? And what level of hominin diversity remains  
663 to be discovered? One can only hope further paleontological finds in Flores and elsewhere  
664 provides further resolution to the evolutionary history, phylogenetic affinities and phenotypic  
665 adaptations of *H. floresiensis*.

666

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673

674 **References**

- 675 1. Aiello, L. C. 2010. Five years of *Homo floresiensis*. *Am. J. Phys. Anthropol.* 142, 167–179.
- 676 2. Aiello, L.C., Wheeler, P. 1995. The expensive-tissue hypothesis: the brain and the digestive  
677 system in human and primate evolution. *Curr. Anthropol.* 36, 199-221.
- 678 3. Argue, D., Morwood, M., Sutikna, T., Jatmiko, Saptomo, W.E. 2009. *Homo floresiensis*: A  
679 cladistic analysis. *J. Hum. Evol.* 57 (5), 623-639.
- 680 4. Arnold, C., Matthews, L.J., Nunn, C.L. 2010. The 10k trees website: a new online resource  
681 for primate phylogeny. *Evol. Anthropol.* 19,114-8.
- 682 5. Baab, K. L. 2012. *Homo floresiensis*: Making sense of the small-bodied hominin fossils from  
683 Flores. *Nature Education Knowledge* 3(9), 4.
- 684 6. Baab, K.L., McNulty, K.P. 2009. Size, shape, and asymmetry in fossil hominins: the status of  
685 the LB1 cranium based on 3D morphometric analyses. *J. Hum. Evol.* 57 (5), 608-622.

- 686 7. Berger, L.R., de Rieter, D.J., Churchill, S.E., Schmid, P., Carlson, K.J., Dirks, P.H.G.M.,  
687 Kibii, J.M. 2010. *Australopithecus sediba*: A new species of Homo-like australopith from  
688 South Africa. *Science* 328 (5975), 195-204.
- 689 8. Bininda-Emonds, O.R.P., Cardillo, M., Jones, K.E., MacPhee, R.D.E., Beck, R.M.D.,  
690 Grenyer, R., Price, S.A., Vos, R.A., Gittleman, J.L., Purvis, A. 2007. The delayed rise of  
691 present-day mammals. *Nature* 446, 507-512.
- 692 9. Bollen, K.A., Jackman, R.W. 1990. Regression diagnostics: An expository treatment of  
693 outliers and influential cases. In *Modern Methods of Data Analysis* (eds J. Fox and J.S.  
694 Long), pp. 257-91. Newbury Park, CA, Sage.
- 695 10. Bromham, L., Cardillo, M. 2007. Primates follow the 'island rule': implications for  
696 interpreting *Homo floresiensis*. *Biol. Lett.* 3 (4), 398-400.
- 697 11. Brown, P. 2012. LB1 and LB6 *Homo floresiensis* are not modern human (*Homo sapiens*)  
698 cretins. *J. Hum. Evol.* 62, 201-224.
- 699 12. Brown, P., Maeda, T. 2009. Liang Bua *Homo floresiensis* mandibles and mandibular teeth: a  
700 contribution to the comparative morphology of a new hominin species. *J. Hum. Evol.* 57 (5),  
701 571-596.
- 702 13. Brown, P., Sutikna, T., Morwood, M.J., Soejono, R.P., Jatmiko, Saptomo, W.E., Rokus Awe  
703 Due. 2004. A new small-bodied hominin from the Late Pleistocene of Flores, Indonesia.  
704 *Nature* 431, 1055-1061.

- 705 14. Brumm, A., Jensen, G.M., van den Bergh, G.D., Morwood, M.J., Kurniawan, I., Aziz, F.,  
706 Storey, M. 2010. Hominins on Flores, Indonesia, by one million years ago. *Nature* 464, 748-  
707 752.
- 708 15. Carlson, K.J., Stout, D., Jashashvili, T., de Ruiter, D.J., Tafforeau, P., Carlson, K., Berger,  
709 L.R. 2011. The endocast of MH1, *Australopithecus sediba*. *Science* 333 (6048), 1402-1407.
- 710 16. Chatterjee, H.J., Ho, S.Y.W., Barnes, I., Groves, C. Estimating the phylogeny and divergence  
711 times of primates using a supermatrix approach. *BMC Evol. Biol.* 9, 259.
- 712 17. Cook, D.R. 1977. Detection of influential observation in linear regression. *Technometrics* 19  
713 (1), 15-18.
- 714 18. Deaner, R.O., Isler, K., Burkart, J., van Schaik, C. 2007. Overall brain size, and not  
715 encephalization quotient, best predicts cognitive ability across non-human primates. *Brain*  
716 *Behav. Evol.* 70, 115-124.
- 717 19. Dennell, R., Roebroeks, W. 2005. An Asian perspective on early human dispersal from  
718 Africa. *Nature* 438, 1099-1104.
- 719 20. Falk, D., Hildebolt, C., Smith, K., Jungers, W., Larson, S., Morwood, M., Sutikna, T.,  
720 Jatmiko, Saptomo, E. W., Prior, F. 2009. The type specimen (LB1) of *Homo floresiensis* did  
721 not have Laron Syndrome. *Am. J. Phys. Anthropol.* 140, 52–63.
- 722 21. Falk, D., Hildebolt, C., Smith, K., Morwood, M.J., Sutikna, T., Brown, P., Jatmiko, Saptomo,  
723 W.E., Brunnsden, B., Prior, F. 2005. The brain of LB1, *Homo floresiensis*. *Science* 308, 242-  
724 245.

- 725 22. Falk, D., Hildebolt, C., Smith, K., Morwood, M.J., Sutikna, T., Jatmiko, Saptomo, E.W.,  
726 Brunnsden, B., Prior, F. 2006. Response to comment on “The brain of LB1 *Homo*  
727 *floresiensis*”. Science 312: 999.
- 728 23. Falk, D., Hildebolt, C., Smith, K., Morwood, M.J., Sutikna, T., Jatmiko, Wayhu Saptomo, E.,  
729 Prior, F. 2009. LB1's virtual endocast, microcephaly, and hominin brain evolution. J. Hum.  
730 Evol., 57(5), 597-607.
- 731 24. Ferring, R., Oms, O., Agusti, J., Berna, F., Nioradze, M., Shelia, T., Tappen, M., Vekua, A.,  
732 Zhvania, D., Lordkipanidze. 2011. Earliest human occupations at Dmanisi (Georgian  
733 Caucasus) dated to 1.85–1.78 Ma. Proc. Natl. Acad. Sci. USA 108 (26), 10432019436.
- 734 25. Foster, J.B. 1964. Evolution of mammals on islands. Nature 202, 234-235.
- 735 26. Gordon, A.D., Nevell, L., Wood, B. 2008. The *Homo floresiensis* cranium (LB1): Size,  
736 scaling, and early Homo affinities. Proc. Natl. Acad. Sci. USA. 105, 4650-4655.
- 737 27. Gould, S.J. 1975. Allometry in primates, with emphasis on scaling and the evolution of the  
738 brain. Contrib. Primatol. 5, 244-292.
- 739 28. Harvey, P.H., Pagel, M.D.. 1991. The comparative method in evolutionary biology. Oxford  
740 University Press, Oxford.
- 741 29. Hershkovitz, I., Kornreich, L., Laron, Z. 2008. Comparative skeletal features between *Homo*  
742 *floresiensis* and patients with primary growth hormone insensitivity (Laron syndrome). Am.  
743 J. Phys. Anthropol. 134 (2), 198-208.

- 744 30. Holloway, R., Schoenemann, T., Monge, J. 2011. The LB1 endocast: un-adorned,  
745 unsmoothed, a replication study based on the original CT scan data. *Am. J. Phys. Anthropol.*  
746 S52, 165-166.
- 747 31. Holloway, R.L. 1980. Within-species brain-body weight variability: A reexamination of the  
748 Danish data and other primate species. *Am. J. Phys. Anthropol.* 53 (1), 109-121.
- 749 32. Irish, J.D., Gautelli-Steinberg, D., Legge, S.S., de Ruyter, D.J., Berger, L.R. 2013. Dental  
750 morphology and the phylogenetic "place" of *Australopithecus sediba*. *Science* 340 (6129).
- 751 33. Isler, K., Kirk, E.C., Miller, J.M.A., Albrecht, G.A., Gelvin, B.R., Martin, R.D. 2008.  
752 Endocranial volumes of primate species: scaling analyses using a comprehensive and reliable  
753 data set. *J. Hum. Evol.* 55 (6), 967–978.
- 754 34. Jacob, T., Indriati, E., Soejono, R.P., Hsu, K., Frayer, D.W., Eckhardt, R.B., Kuperavage,  
755 A.J., Thorne, A., Henneberg, M. 2006. Pygmoid Australomelanesian *Homo sapiens* skeletal  
756 remains from Liang Bua, Flores: Population affinities and pathological abnormalities. *Proc.*  
757 *Natl. Acad. Sci. USA* 103 (36), 13421-13426.
- 758 35. Jerison, H.J. 1973. *Evolution of the Brain and Intelligence*. New York: Academic Press.
- 759 36. Jungers, W.L., Harcourt-Smith, W.E.H., Wunderlich, R.E., Tocheri, M.W., Larson, S.G.,  
760 Sutikna, T., Due Awe, R., Morwood, M.J. 2009a. The foot of *Homo floresiensis*. *Nature* 459,  
761 81-84.
- 762 37. Jungers, W.L., Larson, S.G., Harcourt-Smith, W., Morwood, M.J., Sutikna, T., Due Awe, R.,  
763 Djubiantono, T. 2009b. Descriptions of the lower limb skeleton of *Homo floresiensis*. *J.*  
764 *Hum. Evol.* 57 (5), 538-554.

- 765 38. Kappelman, J. 1996. The evolution of body mass and relative brain size in fossil hominids. J.  
766 Hum. Evol. 30, 243.
- 767 39. Kimbel, W.H. 2013. Hesitation on hominin history. Nature 497, 573-574.
- 768 40. Köhler, M., Moyà-Solà, S. 2008. Island rules cannot be broken. Trends Ecol. Evol. 23 (1), 6-  
769 7.
- 770 41. Köhler, M., Moyà-Solà, S. 2004. Reduction of brain and sense organs in the fossil insular  
771 bovid *Myotragus*. Brain Behav. Evol. 63, 125-140.
- 772 42. Kaifu, Y., Baba, H., Sutikna, T., Morwood, M.J., Kubo, D., Saptomo, E.W., Jatmiko, Awe,  
773 D.R., Djubiantono, T. 2011. Craniofacial morphology of *Homo floresiensis*: Description,  
774 taxonomic affinities, and evolutionary implication. J. Hum. Evol. 61 (6), 644-682.
- 775 43. Kubo, D., Kono, R.T., Kaifu, Y. 2013. Brain size of *Homo floresiensis* and its evolutionary  
776 implications. Proc. Roy. Soc. B 280 (1760), 20130338.
- 777 44. Lande, R. 1979. Quantitative genetic analysis of multivariate evolution, applied to brain:  
778 body size allometry. Evolution 33(1), 402-416.
- 779 45. Larson, S.G., Jungers, W.L., Morwood, M.J., Sutikna, T., Jatmiko, Saptomo, E.W., Due,  
780 R.A., Djubiantono, T. 2007. *Homo floresiensis* and the evolution of the hominin shoulder. J.  
781 Hum. Evol. 53 (6), 718-731.
- 782 46. Lieberman, D.E. 2009. *Homo floresiensis* from head to toe. Nature 459, 41-42.
- 783 47. Lomolino, M.V. 1985. Body size of mammals on islands: the island rule reexamined. Am.  
784 Nat. 125, 310-316.

- 785 48. Lordkipanidze, D., Jashashvili, T., Vekua, A., Ponce de Leon, M.S., Zollikofer, C.P.E.,  
786 Rightmire, G.P., Pontzer, H., Ferring, R., Oms, O., Tappen, M., Bukhsianidze, M., Agusti, J.,  
787 Kahlke, R., Kiladze, G., Martinez-Navarro, B., Mouskhelishvili, A., Nioradze, M., Rook, L.,  
788 2007. Postcranial evidence from early Homo from Dmanisi, Georgia. *Nature* 449, 305–310.
- 789 49. Mace, G.M., Eisenberg, J.F. 1982. Competition, niche specialization and the evolution of  
790 brain size in the genus *Peromyscus*. *Biol. J. Linn. Soc.* 17: 243–257.
- 791 50. Martin, R.D., MacLarnon, A.M., Phillips, J.L., Dussubieux, L., Williams, P.R., Dobyns,  
792 W.B. 2006. Comment on “the brain of LB1 *Homo floresiensis*”. *Science* 312, 999b.
- 793 51. Martinez, A.M., Hamsici, O.C. 2008. Who is LB1? Discriminate analysis for the  
794 classification of specimens. *Pattern Recogn.* 41 (11), 3436-3441.
- 795 52. McClain, C.R., Durst, P.A.P., Boyer, A.G., Francis, C.D. 2013. Unraveling the determinants  
796 of insular body size shifts. *Biol. Lett.* 9(1).
- 797 53. Meijer, H.J.M., van den Hoek Ostende, L.W., van den Bergh, G.D., de Vos, J. 2010. The  
798 fellowship of the hobbit: the fauna surrounding *Homo floresiensis*. *J. Biogeog.* 37 (6), 995–  
799 1006.
- 800 54. Meiri, S., Cooper, N., Purvis, A. 2008. The island rule: made to be broken? *Proc. R. Soc. B*  
801 275 (1631), 141-148.
- 802 55. Meiri, S., Dayan, T., Simberloff, D. 2006. The generality of the island rule reexamined. *J.*  
803 *Biogeog.* 33: 1571–1577.
- 804 56. Meiri, S., Raia, P., Phillimore, A.B. 2011. Slaying dragons: limited evidence for unusual  
805 body size evolution on islands. *J. Biogeog.* 38: 89–100.

- 806 57. Montgomery, S.H., Capellini, I., Barton, R.A., Mundy, N.I. 2010. Reconstructing the ups and  
807 downs of primate brain evolution: implications for adaptive hypotheses and *Homo*  
808 *floresiensis*. BMC Biol. 8 (9).
- 809 58. Montgomery, S.H., Mundy, N.I. 2013. Parallel episodes of phyletic dwarfism in callitrichid  
810 and cheirogaleid primates. J. Evol. Biol. in press.
- 811 59. Morwood, M.J., Brown, P., Jatmiko, Sutikna, T., Saptomo, E.W., Westaway, K.E., Due,  
812 R.A., Roberts, R.G., Maeda, T., Wasisto, S., Djubiantono, T. 2005. Further evidence for  
813 small-bodied hominins from the Late Pleistocene of Flores, Indonesia. Nature 437, 1012-  
814 1017.
- 815 60. Navarrete, A., van Schaik, C.P., Isler, K. 2011. Energetics and the evolution of human brain  
816 size. Nature 480, 91-93.
- 817 61. Niven, J.E. 2007. Brains, islands and evolution: breaking all the rules. Trends Ecol. Evol. 22,  
818 57–59.
- 819 62. Oxnard, C., Obendorf, P.J., Kefford, B.J. 2010. Post-cranial skeletons of hypothyroid cretins  
820 show a similar anatomical mosaic as *Homo floresiensis*. PLoS ONE 5(9), e13018.
- 821 63. Pagel, M. 1999. Inferring the historical patterns of biological evolution. Nature 401, 877-  
822 884.
- 823 64. Perelman, P., Johnson, W.E., Roos, C., Seuánez, H.N., Horvath, J.E., Moreira, M.A.M.,  
824 Kessing, B., Pontius, J., Roelke, M., Rumpler, Y., Schneider, M.P.C., Silva, A., O'Brien, S.J.,  
825 Pecon-Slattery, J. 2011. A molecular phylogeny of living primates. PLoS Genet 7(3):  
826 e1001342.

- 827 65. Peters, M., Jäncke, K., Staigner, J.F., Schlaug, G., Huang, Y., Steinmetz, H. 1998. Unsolved  
828 problems in comparing brain sizes in *Homo sapiens*. *Brain Cogn.* 37, 254-285.
- 829 66. Pilbeam, D., Gould, S.J. 1974. Size and scaling in human evolution. *Science* 186 (4167),  
830 892-901.
- 831 67. Price, T.D., Phillimore, A.B. 2007. Reduced major axis regression and the island rule. *J.*  
832 *Biogeog.* 34 (11), 1998-1999.
- 833 68. Reader, S.M., Laland, K.N. 2002. Social intelligence, innovation, and enhanced brain size in  
834 primates. *Proc. Natl. Acad. Sci. USA* 99 (7), 4436-4441.
- 835 69. Rightmire, G.P., Lordkipanidze, D., Vekua, A. 2006. Anatomical descriptions, comparative  
836 studies and evolutionary significance of the hominin skulls from Dmanisi, Republic of  
837 Georgia. *J. Hum. Evol.* 50 (2), 115-141.
- 838 70. Robson, S.L., Wood, B. 2008. Hominin life history: reconstruction and evolution. *J. Anat.*  
839 212 (4), 394-425.
- 840 71. Roth, V.L. 1992. Inferences from allometry and fossils: dwarfing of elephants on islands.  
841 *Oxf. Surv. Evol. Biol.* 8, 259-288.
- 842 72. Schillaci, M.A., Meijaard, E., Clark, T. 2009. The effect of island area on body size in a  
843 primate species from the Sunda Shelf Islands. *J. Biogeog.* 36, 362-371.
- 844 73. Sol, D. 2009. Revisiting the cognitive buffer hypothesis for the evolution of large brains.  
845 *Biol. Lett.* 5 (1), 130-133.

- 846 74. Stanyon, R., Consigliere, S., Morescalchi, M.A. 1993. Cranial capacity in hominid evolution,  
847 Hum. Evol. 8, 205.
- 848 75. Taylor, A.B., van Schaik, C.P. 2007. Variation in brain size and ecology in Pongo. J. Hum.  
849 Evol. 52, 59-71.
- 850 76. Tocheri, M.W., Orr, C.M., Larson, S.G., Sutikna, T., Jatmiko, Saptomo, E.W., Due, W.A.,  
851 Djubiantono, T., Morwood, M.J., Jungers, W.L. 2007. The primitive wrist of *Homo*  
852 *floresiensis* and its implications for hominin evolution. Science 317 (5845), 1743-1745.
- 853 77. Trueman, J.W. 2010. A new cladistic analysis of *Homo floresiensis*. J. Hum. Evol. 59(2),  
854 223-226.
- 855 78. Uhl, N.M., Rainwater, C.W., Konigsberg, L.W. 2013. Testing for size and allometric  
856 differences in fossil hominin body mass estimation. Am. J. Phys. Anthropol. 151 (2), 215-  
857 229.
- 858 79. van Heteren, A.H. 2008. *Homo floresiensis* as an island form. Palarch's J. Vert. Palaeontol.  
859 (5)2, 1-19.
- 860 80. van Heteren, A.H. 2012. The hominins of Flores: Insular adaptations of the lower body.  
861 Comptes. Rendus. Palevol. 11(2-3), 169-179.
- 862 81. Van Valen, L. 1973. Pattern and the balance of nature. Evol. Theory. 1, 31-49.
- 863 82. van Woerden, J.T., van Schaik, C.P., Isler, K. 2010. Effects of seasonality on brain size  
864 evolution: evidence from strepsirrhine primates. Am. Nat. 176 (6), 758-767.

- 865 83. van Woerden, J.T., Willems, E.P., van Schaik, C.P., Isler, K. 2012. Large brains buffer  
866 energetic effects of seasonal habitats in catarrhine primates. *Evolution* 66(1), 191-199.
- 867 84. Vekua, A., Lordkipanidze, D., Rightmire, G.P., Agusti, J., Ferring, R., Maisuradze, G.,  
868 Mouskhelishvili, A., Nioradze, M., Ponce de Leon, M., Tappen, M., Tvalchredlidze, M.,  
869 Zollikofer, C. 2002. A new skull of early *Homo* from Dmanisi, Georgia. *Science* 297 (5578),  
870 85-89.
- 871 85. Warton, D. I., Ormerod, J. 2007. Smatr package, v. 2.1. See  
872 <http://web.maths.unsw.edu.au/~dwarton>.
- 873 86. Welch, J.J. 2009. Testing the island rule: primates as a case study. *Proc. Roy. Soc. B* 276,  
874 675-682.
- 875 87. Weston, E.M., Lister, A.M. 2009. Insular dwarfism in hippos and a model for brain size  
876 reduction in *Homo floresiensis*. *Nature* 459 (7), 85-88.
- 877 88. Wood, B. 2011. Did early *Homo* migrate "out of" or "in to" Africa? *Proc. Natl. Acad. Sci.*  
878 108 (26), 10375-10376.
- 879 89. Wood, B., Collard, M. 1999. The human genus. *Science* 284, 65-71.
- 880 90. Wood, B., Harrison, T. 2011. The evolutionary context of the first hominins. *Nature* 470,  
881 347-352.

## Figure legends

**Figure 1: Estimated body masses for *H. floresiensis* using the brain:body scaling relationship from each mainland/island primate pairs.** The range of estimates is shown for each assumed ancestor, given along the x-axis. The lower grey box gives the range of body mass estimates for LB1 derived from femoral cross-sectional area, the upper grey box gives the range derived from the femoral head diameter (Brown et al., 2004; Aiello, 2009).

## Figure 2:

**A) Relationship between  $R_{\text{brain}}$  and  $R_{\text{body}}$  for extant mainland-island pairs.** The regression line is shown with the range of minimum and maximum residuals from the extant data (grey area). The  $R_{\text{brain}}$  and  $R_{\text{body}}$  for each ancestral hominin is shown across a range of body mass estimates for *H. floresiensis*, from 16-41kg (colored horizontal lines). Color coding: black = *A. africanus*; red = *A. sediba*; orange = *H. rudolfensis*; yellow = *H. habilis*; fuchsia = *H. ergaster*; green = Dmanisi hominins; dark blue = Ngandong *H. erectus*; light blue = *H. erectus s. l.*

**B) Leverage of *H. floresiensis* on the Relationship between  $R_{\text{brain}}$  and  $R_{\text{body}}$ .** Cook's Distances were calculated for the *H. floresiensis* data point across a range of *H. floresiensis* body masses (0-45kg). The dashed grey line is the cut-off threshold above which the *H. floresiensis* data point has high leverage. Color coding is the same as panel A. Note that for *H. erectus s. l.*, Ngandong *H. erectus* and *H. ergaster*, after the curve's peak the  $D_i$  begins to fall again (dashed lines), however this is because these data so strongly influence the regression that the slope of the relationship flattens and significance is lost.

**Table 1: Phenotypic data for 7 pairs of insular primates and their closest mainland relative<sup>1</sup>**

	<b>Species</b>	<b>Brain mass [g]</b>	<b>Body mass [g]</b>	<b>EQ</b>	<b>N Brain mass<sup>2</sup></b>	<b>N body mass<sup>2</sup></b>
<i>Mainland</i>	<i>Macaca mulatta</i>	92.188	6792.766	2.080	103 (44/59)	83 (37/46)
<i>Island</i>	<i>Macaca cyclopis</i>	84.952	5470.000	2.216	2 (1/1)	11 (7/4)
<i>Mainland</i>	<i>Macaca radiata</i>	77.569	5084.083	2.125	9 (6/3)	9 (6/3)
<i>Island</i>	<i>Macaca sinica</i>	72.207	4440.000	2.166	13 (12/1)	78 (23/55)
<i>Mainland</i>	<i>Macaca fascicularis fascicularis</i>	64.297	3950.314	2.086	72 (45/27)	67 (44/23)
<i>Island</i>	<i>Macaca fascicularis fusca</i>	68.140	5017.875	1.883	11 (7/4)	11 (7/4)
<i>Mainland</i>	<i>Presbytis femoralis</i>	76.390	6847.878	1.714	14 (9/5)	14 (9/5)
<i>Island</i>	<i>Presbytis natunae</i>	59.254	5065.333	1.627	3 (0/3)	4 (0/4)
<i>Mainland</i>	<i>Nasalis larvatus</i>	95.627	14560.724	1.294	45 (24/21)	37 (10/17)
<i>Island</i>	<i>Simias concolor</i>	60.528	7975.000	1.226	6 (3/3)	6 (3/3)
<i>Mainland</i>	<i>Trachypithecus johnii</i>	87.646	11600.000	1.382	1 (1/0)	10 (7/3)
<i>Island</i>	<i>Trachypithecus vetulus</i>	63.499	6237.000	1.517	6 (3/3)	6 (3/3)
<i>Mainland</i>	<i>Hylobates moloch</i>	106.708	6577.000	2.460	1 (1/0)	1 (1/0)
<i>Island</i>	<i>Hylobates klossii</i>	91.159	5795.000	2.288	6 (2/4)	6 (2/4)

<sup>1</sup> species pairs from Bromham and Cardillo (2007), phenotypic data from Isler et al. (2008);<sup>2</sup> total number and number of males/females in brackets

**Table 2: Phenotypic data for the hominin taxa used to model the descent of *H. floresiensis***

<b>Grouping</b>	<b>Species/taxa</b>	<b>Body mass (kg)</b>	<b>Brain mass (g)</b>	<b>Notes and references</b>
<i>Homo erectus</i> <i>and allies</i>	<i>H. erectus s.l.</i> (broadly defined)	60.0	1026.7	Following Martin et al. (2006); Stanyon et al. (1993) and Kappleman et al. 1996
	<i>H. erectus</i> (Ngandong)	60.0	1190.4	Following Martin et al. (2006); Stanyon et al. (1993) and Kappleman et al. 1996
	Dmanisi homoids	50.0	687.9	Following Martin et al. (2006); Vekua et al. (2002) and Rightmire et al. (2006)
	<i>H. ergaster</i>	58.0	884.7	Wood and Collard (1999)
Early <i>Homo</i>	<i>H. habilis</i>	32.6	571.9	Kappelman et al. (1996)
	<i>H. rudolfensis</i>	50.0	779.1	Kappelman et al. (1996); Robson and Wood (2008)
Australopithecines	<i>A. sediba</i>	33.6	435.1	Wood and Collard (1999)
	<i>A. africanus</i>	36.0	473.5	Carlson et al. (2011)

**Table 3. Application of scaling models to extant mainland/island species pairs<sup>1</sup>**

**a) Non-primate models of insular dwarfism**

Species pair	Island	Martin et al.'s Model A		Weston and Lister, late ontogenetic		Weston and Lister, late ontogenetic + 30% <sup>2</sup>	
	body mass [g]	estimate	%age error	estimate	%age error	estimate	%age error
<i>Macaca mulatta/M. cyclopis</i>	5470	5260.9-5377.4	3.8-1.7	5377.4	1.7	-	-
<i>Macaca radiata/M. sinica</i>	4440	4064.7-4143.4	8.5-6.7	4143.4	6.7	-	-
<i>Macaca f. fascicularis/M. f. fusca</i>	5018	4735.0-4662.0	5.6-7.1	4662.0	7.1	-	-
<i>Presbytis femoralis/P. natunae</i>	5065	3095.5-3313.5	38.9-34.6	3313.5	34.6	-	-
<i>Nasalis larvatus/Simias concolor</i>	7975	3487.3-3941.8	56.3-50.6	3941.8	50.6	10921.3	-36.9
<i>Trachypithecus johnii/T. vetulus</i>	6237	4236.7-4618.7	32.1-25.9	4618.7	25.9	-	-
<i>Hylobates moloch/H. klossii</i>	5795	4020.5-4193.7	30.6-27.6	4193.7	27.6	-	-

**a) cont'd**

Species pair	Island	Weston and Lister, early ontogenetic		Weston and Lister, early ontogenetic +17% <sup>2</sup>	
	body mass [g]	estimate	%age error	estimate	%age error
<i>Macaca mulatta/M. cyclopis</i>	5470	5708.0	-4.4	-	-
<i>Macaca radiata/M. sinica</i>	4440	4365.6	1.7	-	-
<i>Macaca f. fascicularis/M. f. fusca</i>	5018	4468.8	10.9	-	-
<i>Presbytis femoralis/P. natunae</i>	5065	3988.2	21.3	5928.6	-17.1
<i>Nasalis larvatus/Simias concolor</i>	7975	5502.9	31.0	8180.2	-2.6
<i>Trachypithecus johnii/T. vetulus</i>	6237	5842.9	6.3	8685.7	-39.3
<i>Hylobates moloch/H. klossii</i>	5795	4704.3	18.8	-	-

**b) Intraspecific scaling models**

<b>Species pair</b>	Island	Typical mammalian (Martin et al.'s Model B)		Non-human primates		Human (Martin et al.'s Model C)	
	body mass [g]	b = 0.2-0.4		b = 0.124-0.349		b = 0.03-0.17	
		estimate	%age error	estimate	%age error	estimate	%age error
<i>Macaca mulatta/M. cyclopis</i>	5470	4512.9-5536.8	17.5--1.2	3512.4-5373.8	35.8-1.8	444.7-4198.7	91.9-23.2
<i>Macaca radiata/M. sinica</i>	4440	3554.0-4250.7	20.0-4.3	2853.8-4141.0	35.7-6.7	467.4-3336.5	89.5-24.9
<i>Macaca f. fascicularis/M. f. fusca</i>	5018	5279.0-4566.4	-5.2-9.0	6306.0-4664.2	-25.7-7.1	27308.8-5556.2	-444.2—10.7
<i>Presbytis femoralis/P. natunae</i>	5065	1922.3-3628.2	62.0-28.4	882.3-3306.6	82.6-34.7	1.4-1536.2	100.0-69.7
<i>Nasalis larvatus/Simias concolor</i>	7975	1479.4-4641.2	81.5-41.8	364.2-3927.1	95.4-50.8	0.0-988.1	100.0-87.6
<i>Trachypithecus johnii/T. vetulus</i>	6237	2315.0-5182.2	62.9-16.9	862.2-4606.5	86.2-26.1	0.3-1742.0	100.0-72.1
<i>Hylobates moloch/H. klossii</i>	5795	2992.4-4436.4	48.4-23.4	1846.8-4188.3	68.1-27.7	34.5-2604.2	99.4-55.1

<sup>1</sup> percentage errors:  $100 - [(estimate/observed)*100]$

<sup>2</sup> results are only shown for pairs where the additional range does not result in the island species having a larger brain than the mainland species

**Table 4. Range of *H. floresiensis* body masses that produce low Cook's Distance values for each ancestor using the mean scaling component between mainland/island pairs of extant primates**

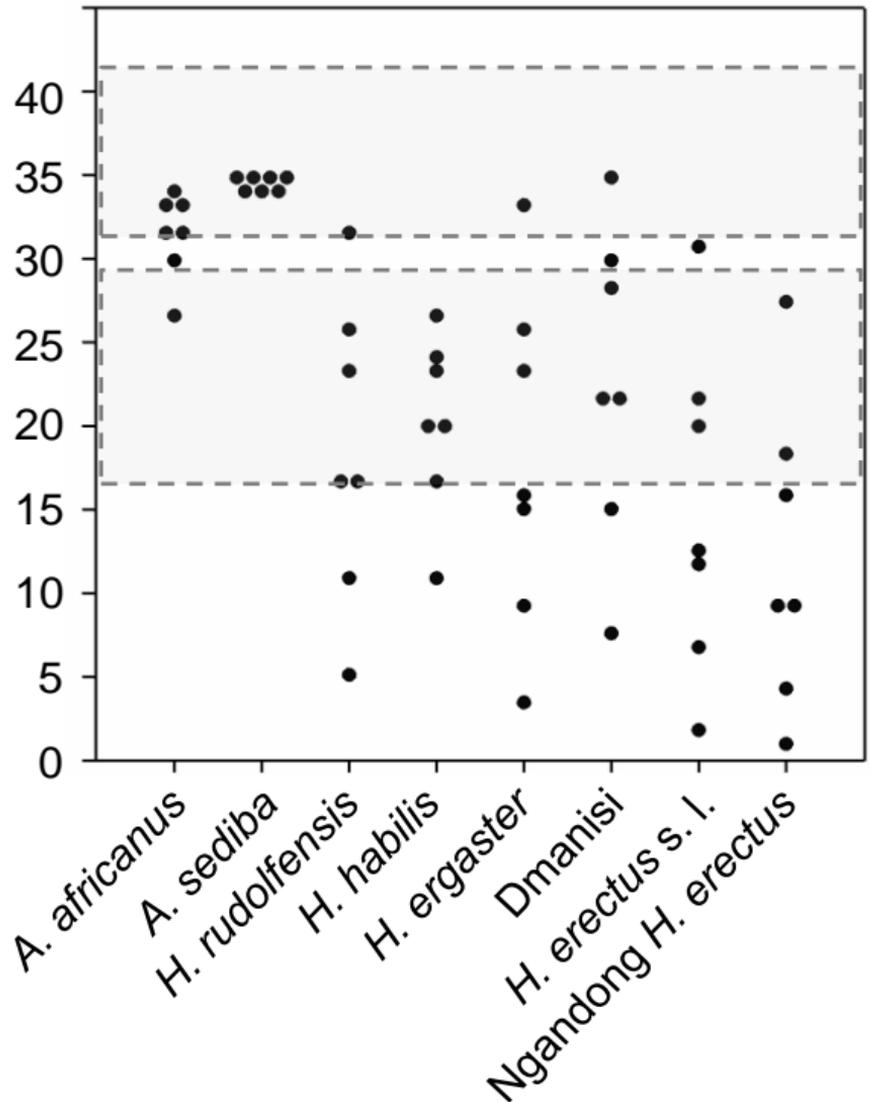
	<i>H. floresiensis</i> body mass		
	Lower limit of range, $D_i < 0.5$	Upper limit of range, $D_i < 0.5$	Body mass at minimum $D_i$
<i>Australopithecus africanus</i>	22	42	35
<i>Australopithecus sediba</i>	22	43	38
<i>Homo rudolfensis</i>	8	>45	15
<i>Homo habilis</i>	12	38	25
<i>Homo ergaster</i>	<1	19	10
Dmanisi homoids	14	>45	22
<i>Homo erectus s. l.</i>	<1	11	2
<i>Homo erectus</i> (Ngandong)	<1	3	1

**Table 5. Predicted *H. floresiensis* body masses that based on intraspecific scaling components and a range of ancestors**

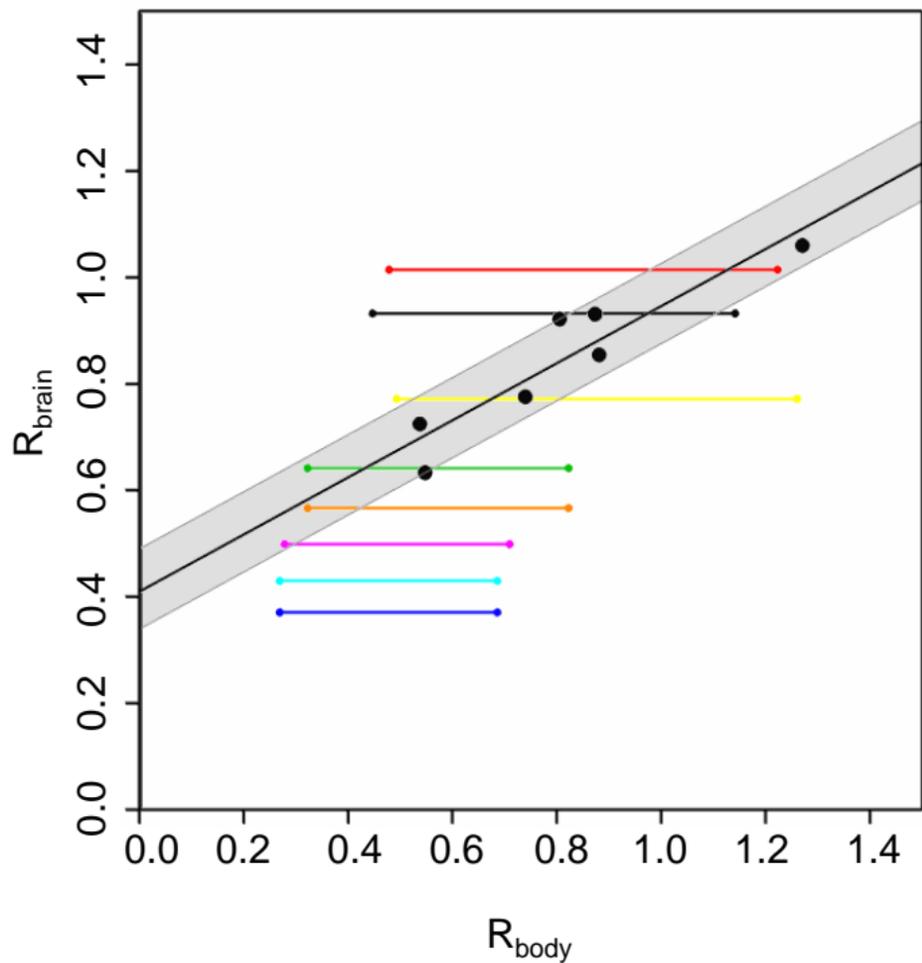
	Range of estimated body mass for <i>H. floresiensis</i> derived from scaling components (b)		
	Typical mammalian intraspecific scaling (b = 0.2-0.4)	Primate intraspecific scaling (b = 0.12-0.33)	Martin et al.'s human intraspecific scaling (0.03-0.17)
<i>Australopithecus africanus</i>	25.4 - 30.2	20.4 - 29.4	3.5 - 23.8
<i>Australopithecus sediba</i>	34.8 - 36.1	35.0-37.7	36.5 - 53.9
<i>Homo rudolfensis</i>	2.9 - 12.1	0.5 - 9.8	0.0 - 1.8
<i>Homo habilis</i>	8.9 - 17.1	4.0 - 15.5	0.0 - 7.1
<i>Homo ergaster</i>	1.8 - 10.2	0.2 - 7.9	0.0 - 1.0
Dmanisi homoids	5.4 - 16.5	1.4 - 14.0	0.0 - 3.7
<i>Homo erectus s. l.</i>	0.9 - 7.3	0.1 - 5.3	0.0 - 0.4
<i>Homo erectus</i> (Ngandong)	0.4 - 5.0	0.0 - 3.5	0.0 - 0.2



*H. floresiensis* body mass (kg)



A



B

