

lineage, as opposed to Th2 or Th17 differentiation.

### Acknowledgments

This work is funded by an Australian National Health and Medical Research Council (NHMRC) CDF Fellowship and a Monash Fellowship to D.Y.

<sup>1</sup>Molecular Immunomodulation Laboratory, Department of Biochemistry and Molecular Biology, School of Biomedical Sciences, Monash University, Victoria 3800, Australia

<sup>2</sup>Centre for Inflammatory Diseases, School of Clinical Sciences, Monash University, Victoria 3800, Australia

\*Correspondence: Di.Yu@monash.edu (D. Yu).

<http://dx.doi.org/10.1016/j.it.2015.11.001>

### References

1. Crotty, S. (2014) T follicular helper cell differentiation, function, and roles in disease. *Immunity* 41, 529–542
2. Nakayama, S. *et al.* (2011) Early Th1 cell differentiation is marked by a Tfh cell-like transition. *Immunity* 35, 919–931
3. Oestreich, K.J. *et al.* (2012) Molecular mechanisms that control the expression and activity of Bcl-6 in TH1 cells to regulate flexibility with a TFH-like gene profile. *Nat. Immunol.* 13, 405–411
4. Choi, Y.S. *et al.* (2015) LEF-1 and TCF-1 orchestrate TFH differentiation by regulating differentiation circuits upstream of the transcriptional repressor Bcl6. *Nat. Immunol.* 16, 980–990
5. Wu, T. *et al.* (2015) TCF1 is required for the T follicular helper cell response to viral infection. *Cell Rep.*
6. Xu, L. *et al.* (2015) The transcription factor TCF-1 initiates the differentiation of TFH cells during acute viral infection. *Nat. Immunol.* 16, 991–999
7. De Obaldia, M.E. and Bhandoola, A. (2015) Transcriptional regulation of innate and adaptive lymphocyte lineages. *Annu. Rev. Immunol.* 33, 607–642
8. Zhou, X. *et al.* (2010) Differentiation and persistence of memory CD8<sup>+</sup> T cells depend on T cell factor 1. *Immunity* 33, 229–240
9. Yu, Q. *et al.* (2009) T cell factor 1 initiates the T helper type 2 fate by inducing the transcription factor GATA-3 and repressing interferon-gamma. *Nat. Immunol.* 10, 992–999
10. Auderset, F. *et al.* (2013) Notch signaling regulates follicular helper T cell differentiation. *J. Immunol.* 191, 2344–2350

## Spotlight CTLA-4 and Autoimmunity: New Twists in the Tale

Lucy S.K. Walker<sup>1,\*</sup>

**CTLA-4 has long been associated with control of autoimmunity. A recent study by Sharpe and**

**colleagues explores this relationship in a model that enables conditional deletion of CTLA-4 in adult mice, with some surprising new conclusions.**

CTLA-4 is one of a small number of immune genes whose deletion triggers fatal lymphoproliferative disease. Its importance in immune regulation is unquestioned; however, its precise mechanism of action has been hotly debated. In a recent paper in *The Journal of Experimental Medicine* [1] the Sharpe laboratory, who originally reported the phenotype of CTLA-4 null mice, brings us another intriguing installment in the CTLA-4 story. By generating mice with a floxed CTLA-4 allele, and crossing them to mice expressing a tamoxifen inducible Cre-recombinase, the authors constructed an experimental system in which CTLA-4 could be ablated ‘at will’ by tamoxifen administration. This permits for the first time the deletion of the CTLA-4 gene only in adult mice.

The first surprise in the data is that punctual ablation of the CTLA-4 gene in 7-week old mice failed to trigger spontaneous autoimmunity, contrasting with the situation following germline deletion. Furthermore, the CTLA-4-deleted mice exhibited reduced susceptibility to MOG-induced experimental autoimmune encephalomyelitis (EAE) and did not exhibit an increased capacity to clear tumors, both opposite to expectations based on the concept that CTLA-4 inhibits autoimmunity and limits anti-tumor responses.

So what is the basis for these striking findings? By breeding the tamoxifen deletable CTLA-4 mice to a MOG-specific TCR transgenic mouse (2D2) the authors first homed in on the role of CTLA-4 in conventional T (Tconv) cells. Adoptively transferred 2D2 T cells induced EAE in RAG-deficient mice with similar onset and severity regardless of whether these cells expressed CTLA-4 or not. An elegant approach in which CTLA-4 was deleted

from IL-17 expressing cells T cells (IL-17F-Cre) also revealed no difference in EAE incidence. Next, adoptive transfer experiments were performed in which both T regulatory (Treg) cells and Tconv cells were injected into recipient mice, but CTLA-4 deficiency was restricted to the latter: these experiments revealed a subtle increase in Tconv cell number in the absence of CTLA-4, although proliferative responses to MOG peptide or anti-CD3 and cytokine differentiation *in vitro* were not altered. Thus, CTLA-4 deficiency in the Tconv compartment had small but discernable biological consequences; however, it did not alter EAE induction.

By contrast, deletion of CTLA-4 from the Treg compartment was sufficient to recapitulate the effects of systemic CTLA-4 deletion, with similar protection from EAE being observed. Thus in keeping with many other experimental settings, the major role of CTLA-4 was shown to be in Treg cells; however, unexpectedly this resulted in disease protection rather than disease exacerbation.

In keeping with previous work, the authors observed a dramatic expansion of the Treg cell population in CTLA-4-deleted mice, reflecting a marked increase in their proliferation (~60% Treg cells were Ki67<sup>+</sup> in CTLA-4-deleted mice as compared to ~30% in controls) [1]. This nicely parallels the increased Treg cell proliferation seen in mice lacking CTLA-4 from birth (~62% Treg were Ki67<sup>+</sup> in CTLA-4<sup>-/-</sup> mice compared with ~28% in littermate controls) [2]. By ablating CTLA-4 specifically in the Treg compartment, using a tamoxifen-inducible cre under the control of the Foxp3 promoter, the authors were able to confirm that the Treg cell expansion was a direct effect of CTLA-4 in the Treg cells themselves. As the authors surmise, this Treg cell expansion following CTLA-4 ablation likely reflects unrestrained CD28 signaling, since CTLA-4 is known to compete with CD28 for access to their shared ligands CD80 and CD86.

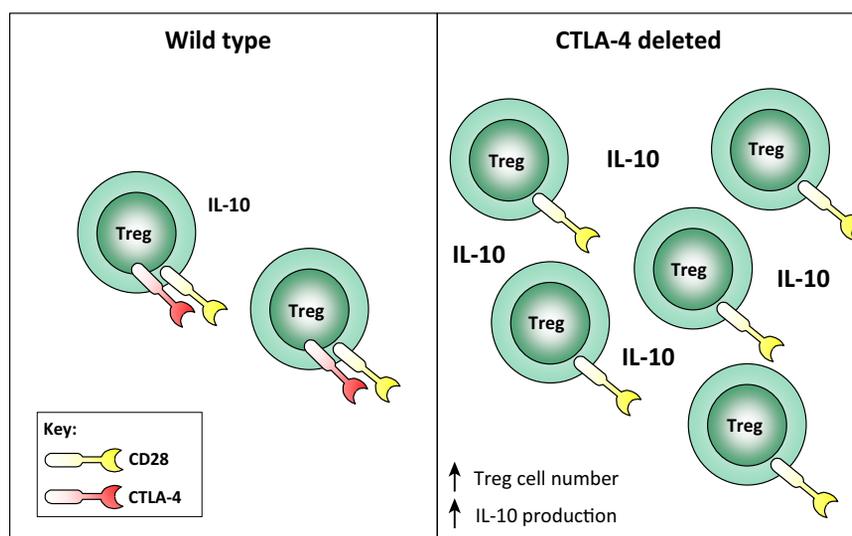
It has been shown previously that CTLA-4<sup>-/-</sup> Treg cells can exhibit a compensatory overproduction of both IL-10 and TGF- $\beta$  [3]. Indeed, the Powrie group demonstrated that while suppression of colitis by wild type Treg cells was CTLA-4 dependent, CTLA-4<sup>-/-</sup> Treg cells were able to use IL-10 to elicit suppression [4], providing a precedent for the compensation of CTLA-4 function by the IL-10 pathway. Data presented by Paterson and colleagues suggest that a similar situation may arise following inducible deletion of CTLA-4: In the context of the EAE model, CTLA-4 ablation was associated with a marked increase in the proportion of Treg cells producing IL-10. The functional impact of this could be potentiated by the greater than 3 fold increase in the absolute numbers of Treg cells. Thus, although most of these Treg cells could no longer use CTLA-4, the population was expanded in number and had an enhanced capacity to produce IL-10 (Figure 1). NanoString transcriptional analysis confirmed that the Treg cells adopted a gene expression signature associated with high IL-10 production, including increased expression of the transcription

factor Blimp-1. Therefore, one explanation for the paradoxical disease protection following CTLA-4 ablation could be the altered cytokine environment imposed by an expanded population of Treg cells that overproduce IL-10.

The authors next set out to test whether CTLA-4- ablated Treg cells had the capacity to elicit suppressive function. They first employed *in vitro* suppression assays, and demonstrated equivalent suppressive function regardless of whether the Treg cells derived from tamoxifen-treated (hence CTLA-4-ablated) animals. This result is not entirely unexpected, since several groups have found that CTLA-4<sup>-/-</sup> Treg cells exhibit intact suppression in such *in vitro* assays [2,3,5]. Moving to an *in vivo* setting, the authors went on to show that CTLA-4-ablated Treg cells retained the capacity to control the homeostatic proliferation of T cells adoptively transferred to Rag-deficient recipients. This contrasts with the findings of Sojka *et al.* [5] who showed that CTLA-4<sup>-/-</sup> Treg were impaired in their capacity to control lymphopenia-induced T cell expansion. This could reflect a

differential requirement for CTLA-4 in particular Treg cell subsets – Sojka *et al.* focused on CD62L<sup>+</sup>Treg cells. Alternatively, it might reflect residual CTLA-4 activity resulting from incomplete gene deletion in the tamoxifen system.

Curiously, despite conferring protection from EAE, loss of CTLA-4 from Treg cells triggered a significant expansion of Tconv cells in the cervical lymph nodes, with more cells expressing ki67 and bearing a CD44<sup>hi</sup>CD62L<sup>lo</sup> phenotype, consistent with a role for Treg cells in suppressing Tconv cell activation via CTLA-4. However, if Tconv cell responses are unleashed following Treg cell CTLA-4 ablation, why then is this associated with disease protection? One possibility is that although Tconv cells become activated, they differentiate in a manner that leads them to be non-pathogenic, at least in EAE. Alterations in T cell cytokine production are known to affect the disease course in the EAE model, sometimes in unexpected ways. For example increases in one cytokine (e.g., IFN $\gamma$ ) can lead to decreases in another (e.g., IL-17) and consequent disease suppression. Paterson *et al.* showed that these Tconv cells exhibited higher levels



Trends in Immunology

**Figure 1. Punctual CTLA-4 Ablation Triggers an Expansion of Treg Cells that Overproduce IL-10.** Deletion of CTLA-4 in adult mice, in all cells or specifically in Treg cells, is associated with an increase in Treg cell number and IL-10 production. The overabundance of IL-10 may suppress autoimmunity, perhaps via the induction of Tr1 cells.

of Ebi3, a subunit of the cytokine IL-27, which is known to inhibit Th17 differentiation and suppress EAE [6]. IL-27 is also recognized for its role in promoting the formation of type 1 regulatory T (Tr1) cells [6], and it is notable that amounts of several typical Tr1 products are increased in the Tconv cells (e.g., IL-10, Ahr, ICOS, LAG3). Thus one could envisage the expanded population of IL-10-producing Treg cells providing a favorable environment for induction of Tr1 cells that might in turn contribute to disease protection.

When the authors crossed the mice expressing a floxed CTLA-4 allele with mice bearing Foxp3-cre, this recapitulated the lethal phenotype reported by others [7], illustrating that CTLA-4 deficiency restricted to the Treg compartment is sufficient to cause fatal disease. Given that inducible CTLA-4 deletion in adult mice (in all cells or only in Treg cells) did not elicit disease, the authors suggest there may be a critical window of time developmentally during which CTLA-4 expression in Treg cells is essential. This could, for example, reflect a need for CTLA-4 during the neonatal period that diminishes once the peripheral immune compartment is fully seeded. Arguing against the need for CTLA-4 solely in the neonatal period is the observation that mixed bone marrow chimeras, comprising CTLA-4<sup>-/-</sup> and wild type cells, rapidly become sick if the wild type cells are deleted during adulthood [8]. Such a hypothesis is also hard to reconcile with the studies that demonstrate defective function of CTLA-4<sup>-/-</sup> Treg cells in

adoptive transfer models, which typically use adult donors and recipients. A comprehensive comparison between conditional CTLA-4 deletion in neonates versus adult mice will be important in exploring this hypothesis further.

An alternative explanation for the lack of disease following ablation of CTLA-4 in adult mice could relate to the efficiency of tamoxifen-driven gene excision in Treg cells, Tconv cells and their precursors. For example, ~ 7% of Treg cells still expressed CTLA-4 after tamoxifen treatment, and it is possible that these contribute to the observed lack of autoimmune disease. Incomplete deletion of CTLA-4 could potentially mirror the situation in humans with heterozygous CTLA-4 deficiency [9,10], some of whom are asymptomatic. Intriguingly, quantitative defects in CTLA-4 in humans are associated with significant expansion of the Treg cell compartment [9], and it is tempting to speculate that protection from autoimmunity in asymptomatic individuals results from compensatory mechanisms such as the overproduction of IL-10 reported here. Notably the autoimmune phenotype in symptomatic individuals with CTLA-4 heterozygosity manifests relatively late in life, contrasting with the early onset associated with Foxp3 deficiency in IPEX. It will be of great interest to track the status of CTLA-4-ablated mice during aging and following a variety of immunological challenges to gain a broader perspective on the role of CTLA-4 in different contexts. Meanwhile,

the increasingly refined tools being used to unravel the biology of this critical immune regulator will doubtless continue to turn up new surprises.

#### Acknowledgments

L.S.K.W. holds a Medical Research Council Senior Fellowship. I am grateful to Vitalijs Ovcinnikovs for helpful discussion and critical reading of the manuscript.

<sup>1</sup>Institute for Immunity & Transplantation, University College London Division of Infection & Immunity, Royal Free Campus, London, NW3 2PF, UK

\*Correspondence: lucy.walker@ucl.ac.uk (Lucy S.K. Walker).

<http://dx.doi.org/10.1016/j.it.2015.11.002>

#### References

1. Paterson, A.M. *et al.* (2015) Deletion of CTLA-4 on regulatory T cells during adulthood leads to resistance to autoimmunity. *J. Exp. Med.* 212, 1603–1621
2. Schmidt, E.M. *et al.* (2009) CTLA-4 controls regulatory T cell peripheral homeostasis and is required for suppression of pancreatic islet autoimmunity. *J. Immunol.* 182, 274–282
3. Tang, Q. *et al.* (2004) Distinct roles of CTLA-4 and TGF-beta in CD4+CD25+ regulatory T cell function. *Eur. J. Immunol.* 34, 2996–3005
4. Read, S. *et al.* (2006) Blockade of CTLA-4 on CD4+CD25+ regulatory T cells abrogates their function in vivo. *J. Immunol.* 177, 4376–4383
5. Sojka, D.K. *et al.* (2009) CTLA-4 is required by CD4+CD25+ Treg to control CD4+ T-cell lymphopenia-induced proliferation. *Eur. J. Immunol.* 39, 1544–1551
6. Pot, C. *et al.* (2011) Induction of regulatory Tr1 cells and inhibition of T(H)17 cells by IL-27. *Semin. Immunol.* 23, 438–445
7. Wing, K. *et al.* (2008) CTLA-4 control over Foxp3+ regulatory T cell function. *Science* 322, 271–275
8. Friedline, R.H. *et al.* (2009) CD4+ regulatory T cells require CTLA-4 for the maintenance of systemic tolerance. *J. Exp. Med.* 206, 421–434
9. Schubert, D. *et al.* (2014) Autosomal dominant immune dysregulation syndrome in humans with CTLA-4 mutations. *Nat. Med.* 20, 1410–1416
10. Kuehn, H.S. *et al.* (2014) Immune dysregulation in human subjects with heterozygous germline mutations in CTLA-4. *Science* 345, 1623–1627