Relationship between SARC-F score and Peripheral Blood Lymphocytes in Juvenile Idiopathic Arthritis: An Observational Pilot Study

Authors

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Acknowledgements

VR received funding support for this work from MRC-CARP Fellowship (MR/T024968/1), Versus Arthritis and UCL Division of Medicine Fellowship (2024). DS and SS received funding support for this work from the MRC CAPE grant.

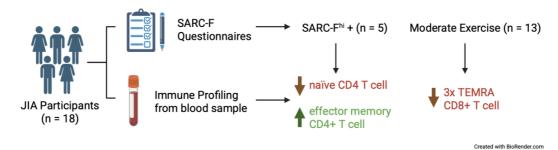
Disclosure of conflicts of interest

None

Key messages

- SARC-F score was used as a clinically relevant tool to identify musculoskeletal function in JIA
- 2. SARC-F^{hi}+ group had fewer naïve CD4+ T cells and greater effector memory CD4+ T cells
- 3. Moderate exercise group had 3-fold lower frequency of CD8+ TEMRA T cells

Graphical Abstract



Abstract

Background

Given the relatively high prevalence of sarcopenia and immune profile abnormalities in inflammatory arthritis such as rheumatoid arthritis (RA), we investigated the relationship between exercise habits, SARC-F score and peripheral blood lymphocyte profiles in patients with Juvenile idiopathic arthritis (JIA).

Methods

All participants with JIA completed the SARC-F survey and self-reported exercise habits through online questionnaires. We defined two groups based on SARC-F scores as musculoskeletal function impaired (SARC-F^{hi}+) and musculoskeletal function not impaired (SARC-F^{hi}-). We used flow cytometry of peripheral blood mononuclear cells to define immune cell subpopulations.

Statistical analyses were performed using GraphPad Prism software version10.

Results

Of the 18 participants with a median age of 24 years – five belonged to the SARC-F^{hi}+ group and 13 to the SARC-F^{hi}- group, of whom 80% and 38% were female, respectively. The SARC-F^{hi} + group had fewer CD45RA+CD27+ naïve CD4+ T cells (26.60% vs. 45.30%, p=0.0264) and greater CD45RA-CD27- effector memory CD4+ T cells (11.90% vs. 7.00%, p=0.0350). The SARC-F^{hi}+ group also had fewer CD56-CD16+ NK cells compared to the SARC-F^{hi}- group (2.60% v. 4.28%, p = 0.0194). The moderate exercise group (n = 13) had increased numbers of total CD4+ T cells (54.06% vs. 37.30%, p=0.0140) and 3-fold fewer CD45RA+CD27- EMRA CD8+ T cells (3.17% vs. 10.40%, p=0.0350). There were no other significant differences for other immune cells / subpopulations including B cells.

Conclusions

Our study findings suggest notable differences in immune profile relative to SARC-F score. Furthermore, the 3-fold lower frequency of CD8+ TEMRA subpopulation of T cells, reflective of immunosenescence, suggest a potential inverse relationship between moderate exercise and peripheral immune cell profile in JIA.

Introduction

A complex, bidirectional relationship between the skeletal muscle and the immune system influences chronic inflammation in ageing (1) and autoimmunity (2). The progressive loss of skeletal muscle mass, strength and function reflecting a generalised disorder of skeletal muscle is referred to as sarcopenia (3, 4). Sarcopenia is associated with poorer quality of life and increased risk of falls, fractures and overall mortality (5, 6). Chronic inflammation is implicated in sarcopenia as it interferes with the balance between muscle protein synthesis and breakdown (7).

Serum concentrations of IL-6, TNF α , and CRP have been shown to be elevated in sarcopenic elderly adults (14). A systematic review estimated that the prevalence of sarcopenia in RA is 31% (8). In RA, the presence of systemic, pro-inflammatory cytokines such as TNF α , IL-1, IL-6 and IFN γ promote skeletal muscle proteolysis and atrophy (9). Alterations in immune profile secondary to the autoimmune disease can influence skeletal muscle function (10). Furthermore, chronic inflammation, ageing, exercise and dietary habits also contribute to the development of sarcopenia in adults with inflammatory diseases such as rheumatoid arthritis (RA) (8, 9, 11, 12).

Juvenile idiopathic arthritis (JIA) is a chronic, autoimmune inflammatory disease (13) with poor outcomes in young adults even in the era of biologics (14). Current evidence suggests that similar to RA and ageing, sarcopenia and reduced bone mass are features of JIA in association with high disease activity (15-19). A recent study reported a significantly higher prevalence of severe sarcopenia of 33% in JIA when compared to 10% in ankylosing spondylitis and 18% in rheumatoid arthritis (20). Furthermore, specific subpopulations of lymphocytes such as memory B cells and effector memory T cells that re-express CD45RA (TEMRA), known for contributing to nonspecific inflammation, are notable at an increased frequency in chronic inflammatory conditions such as RA and ageing (21-23) and JIA (24).

However, our understanding of the relationship between skeletal muscle function, exercise

and peripheral blood immune profile in young adults with JIA is not known.

Given the high prevalence of sarcopenia in the elderly, the SARC-F questionnaire, was developed and validated as a screening tool (25-28) and used pragmatically in people with musculoskeletal symptoms (29). We have recently described the potential utility of the SARC-F questionnaire in identifying people with JIA with sarcopenia and risk of falls (Manuscript in preparation). However, in JIA, the relationship between impaired muscle function, as

assessed using SARC-F, and peripheral blood immune profile remains limited.

Furthermore, dietary and exercise habits are known to influence muscle function and/or immune profiles (11, 12, 30-37). Of direct relevance, a systematic review reported on the benefits of exercise on peripheral blood immune profile including B cells, T cells, NK cells and monocytes (36, 38-44) in addition to improving musculoskeletal function (45). Thus, exercise interventions and habits have the potential to restore peripheral blood immune profile and

sarcopenia.

To this end, we explored the relationship between musculoskeletal function and immune profile, we compared the frequency of peripheral blood lymphocytes and monocytes in people with JIA categorised based on SARC-F score and exercise habits. We demonstrate that in people with JIA SARC-F scores relate to alterations in CD4 T cells and moderate exercise is associated with significantly lower frequency of TEMRA cells.

exercise is associated with significantly lower frequency of TEININA cells

Materials and Methods

Participants

A total of 18 patients under the care of the Young Adult and Adolescent Rheumatology team at University College London Hospital participated in the study. All participants satisfied diagnostic criteria for JIA as defined by the International League of Associations for Rheumatology (46). Patients under the age of 18 years were excluded from the study. Data about current medications including Disease Modifying Anti Rheumatic Drugs, biologics and glucocorticoid use in the last six months was collected from electronic patient records.

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Musculoskeletal function questionnaire

Participants were invited to complete a short online questionnaire. The questionnaire comprised the SARC-F questionnaire and additional questions that included self-reported exercise habits. The SARC-F questionnaire (supplementary table 1) is a validated screening tool to identify the risk of sarcopenia (26). In this study, SARC-F scores were used as a surrogate measure of musculoskeletal function. Participants with a SARC-F score of four or more were considered to have musculoskeletal function impairment (SARC-F^{hi} +) or not impaired (SARC-F^{hi} -). Participants were also categorised according to the frequency of self-reported moderate exercise. Those performing moderate exercise more than once a week were categorised as the 'exercise' group (moderate exercise +) and the remaining were categorised as the 'no exercise' group (moderate exercise -).

Flow cytometry

Peripheral blood samples were collected in VACUETTE tubes anticoagulated with lithium heparin and at least 10 million PBMCs were stored in liquid nitrogen before flow cytometry assessment using the BD LSRFortessa™ X-20 Cell Analyser. The flow cytometry data was analysed using the FlowJo™version 10.10.0 Software.

Statistical analysis

Statistical analysis and data visualisation was conducted using GraphPad Prism 10.2.3. Participant JIA10 was receiving treatment with belimumab, a monoclonal antibody targeting BAFF-R, contributing to low B cell counts. For this reason, JIA10 was excluded from all CD19+B cell analysis. The Mann-Whitney U test was performed to test differences between lymphocyte subsets between the SARC-Fhi+ and SARC-Fhi- groups and the moderate exercise and no moderate exercise groups.

Results

Patient demographics and musculoskeletal function

Of the 18 participants, five were in the SARC-F^{hi}+ group and 13 in the SARC-F^{hi}- group. The median age of both the SARC-F^{hi}+ and SARC-F^{hi}- groups was 24 years. There were four females (80%) in the SARC-F^{hi}+ group compared to five (38.46%) in the SARC-F^{hi}-. SARC-F^{hi}+ and SARC-F^{hi}- subgroup demographics are detailed in Table 1.

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Table 1. Demographic data of the SARC-Fhi+ and SARC-Fhi- subgroups

Demographics		SARC-Fhi+ (n=5)	SARC-Fhi- (n=13)
A	Mean ± SD	23±2.07	24.08±4.61
Age, years	Median	24	24
Sex (%)	Female 4 (80.00%)		5 (38.46%)
Height, cm	Mean ± SD	167.75±8.54	170.17±13.53
	Median	165	168
Weight, kg	Mean ±SD	69.72±5.90	69.92±14.32
	Median	67	64.50
JIA subtype (%)	Systemic arthritis	0	2 (15.38%)
	Oligoarthritis	1 (20.00%)	3 (23.08%)
	Extended oligoarthritis	0	2 (5.38%)
	Polyarthritis	3 (60.00%)	2 (15.38%)
	Psoriatic arthritis	0	1 (7.69%)
	Enthesis-related arthritis	0	2 (15.38%)
	Undifferentiated arthritis	0	1 (7.69%)
	Undifferentiated JIA with lupus	1 (20.00%)	0
DMARD/Biologic therapy, %	Abatacept	1 (20.00%)	0
	Etanercept	0	1 (7.69%)
	Methotrexate	1 (20.00%)	1 (7.69%)
	Infliximab	2 (40.00%)	4 (30.77%)
	Tocilizumab	1 (20.00%)	4 (30.77%)
	Adalimumab	0	1 (7.69%)
	Tofacinib	0	1 (7.69%)
	Belimumab	0	1 (7.69%)
Oral or intravenous steroid use in last six months		3 (60.00%)	3 (23.08%)

Peripheral blood immune profile

Flow cytometry gating strategy is shown in Supplementary Figure 1. There were no significant differences in the frequency of CD19+CD3- B cells, CD19-CD3+ T cells and CD19-CD3- cells between the SARC-F^{hi}+ and SARC-F^{hi}- groups, shown in Figure 1 and Supplemental Table 4.

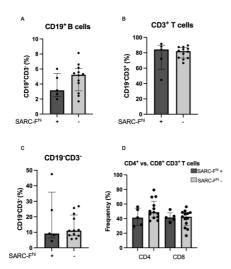


Figure 1. Peripheral blood B cells, T cells and NK cells based on SARC-F scores. Bar graphs represent the frequency of (A) CD19⁺CD3⁻ B cells, (B) CD19⁻CD3⁺ T cells and (C) CD19⁻CD3⁻ cells in the SARC-Fhi⁺ and SARC-Fhi⁻ groups. (D) shows the frequency of CD4⁺ and CD8⁺ cells within the CD19⁻CD3⁺ T-cell gate.

B cells

JIA and exercise habits can influence the frequency of peripheral blood B cell subpopulations (47, 48). However, this study did not find any significant differences in B cell subpopulations (Figure 2 A - D).

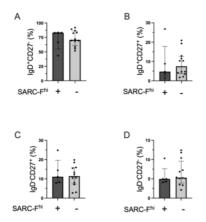


Figure 2. *B* cell subpopulations based on SARC-F scores. (A), IgD+CD27- B cells, (B) IgD+CD27+ B cells, (C) IgD-CD27+ B cells and D) IgD-CD27- B cells in SARC-F^{hi}+ and SARC-F^{hi}- groups.

T cells

T cell profile changes with exercise habits and chronic inflammation in JIA (24, 40). Both the SARC-F^{hi}+ and SARC-F^{hi}- groups had similar frequencies of CD4+ and CD8+ T cells. The SARC-F^{hi}+ group had 41.4% CD4+ and 41.5% CD8+ T cells. Similarly, the SARC-F^{hi}- group have 48.80% CD4+ T cells and 42.6% CD8+ T cells (Figure 1D).

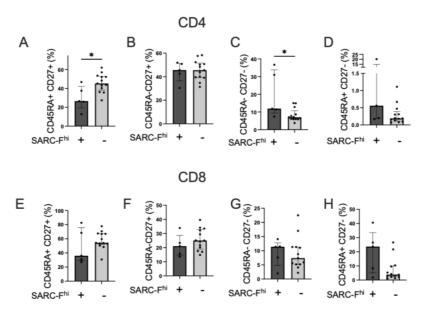


Figure 3. CD4 and CD8 T cell subpopulations in patients with JIA grouped based on SARC-F scores. Bar graphs represent the frequency of CD4⁺ T cells (A), naïve CD45RA⁺CD27⁺ T cells (B), central memory (CM) CD45RA⁻CD27⁺ T cells (C), effector memory (EM) CD45RA⁻CD27⁻ T cells (D) and EMRA CD45RA⁺CD27⁻ T cells in the SARC-Fhi⁺ and SARC-Fhi⁻ groups. * p < 0.05.

In the CD4+ population, the frequency of CD45RA-CD27+ central memory (CM) T cells was 45.6% in the SARC- F^{hi} + group and 45.5% in the SARC- F^{hi} - group and the frequency of CD45A+CD27- TEMRA cells was low at 0.56% in the SARC- F^{hi} + and 0.19% in the SARC- F^{hi} - groups. However, the SARC- F^{hi} + group had significantly lower CD45RA+CD27+ naïve T cells (26.6% v. 45.3%%, p = 0.0264) (Figure 2B) and higher CD45RA-CD27- effector memory (EM) T cells (11.90 v. 7.00%, p = 0.0350) (Figure 3D) (Supplementary table 4). There were no significant differences in the frequency of CD8+ T cell populations (figure 2E-H).

NK cells

Chronic inflammation in JIA (49) and exercise habits influence NK cell frequency in peripheral blood (37). The frequency of CD56dimCD16+ NK cells was 31.5% in the SARC- F^{hi} + group and 57.2% in the SARC- F^{hi} - group. There was a significant difference in the frequency of CD56-CD16+ NK cells with the SARC- F^{hi} - group having a greater proportion (4.28%) as compared to the SARC- F^{hi} + group (2.6%) (p = 0.0194) (Figure 4A). The frequencies of the remaining CD56brightCD16+, CD56brightCD16-, CD56dimCD16+ and CD56dimCD16-groups were similar (Supplementary Table 5).

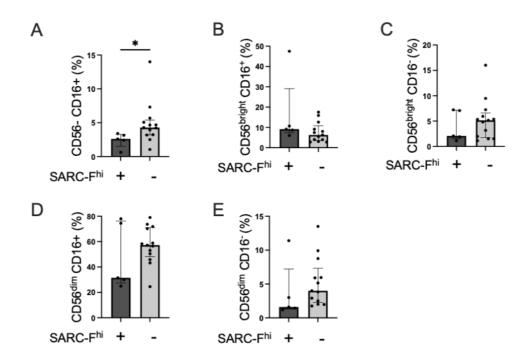


Figure 4. *NK cell subpopulations based on SARC-F scores*. Bar graphs represent the frequency of CD56⁻CD16⁺ (A), CD56brightCD16⁺ (B), CD56brightCD16⁻ (C), CD56dimCD16⁻ (D) and CD56dimCD16⁺ (E) NK cells in the SARC-Fhi⁺ and SARC-Fhi⁻ groups. * p < 0.05.

Moderate exercise

There were 13 participants in the exercise group and five in the no exercise group. The median ages in the exercise and no exercise groups were 23 years and 25 years, respectively. There were six females (46.15%) in the exercise group and three (60%) in the no exercise group (Table 4).

Table 2. Demographic data of the moderate exercise and no moderate exercise groups.

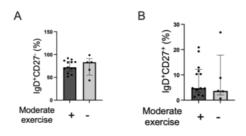
Demographics		Moderate exercise (n=13)	No moderate exercise (n=5)	
Age, years	Mean ± SD	24±4.50	24±2.77	
	Median	23	25.00	
Sex (%)	Female	6 (46.15%)	3.00 (60.00%)	
Height,	Mean ± SD	170.67±12.66	167±10.23	
	Median	166.5	166.50	
Weight, kg	Mean ±SD	72.28±13.03	63.58±8.49	
	Median	68.60	65.00	

Peripheral blood immune profile and exercise

There were similar frequencies of CD19+CD3- B cells, CD19-CD3+ T cells and CD19-CD3-cells (Supplementary Table 6).

B cells

Alterations in B cell profile can occur in JIA and with exercise habits (47, 48). However, we did not find any significant differences in B cell subpopulations between the two groups categorised based on self-reported moderate exercise (Figure 5 A- D).



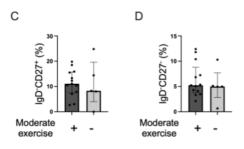


Figure 5. B cell subpopulations in patients with JIA grouped by self-reported exercise habits. Bar graphs represent the frequency of IgD+CD27- B cells (A), IgD+CD27+ B cells (B), IgD-CD27+ B cells (C) and IgD-CD27- B cells (D) in the Moderate + group (participants who perform moderate exercise) and Moderate - group (participants with no moderate exercise).

T cells

JIA and exercise habits can alter T cell profile (24, 40). The exercise group had significantly greater number of peripheral CD4+ T cells (52.30%) compared to the no exercise group (37.30%), (p = 0.0140), whereas no such differences between the groups were noted for CD8+ T cells, Supplementary Table 7.

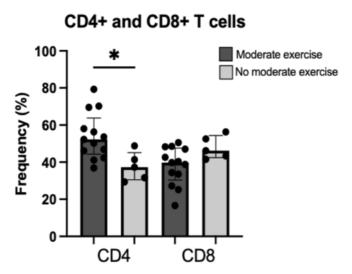


Figure 6. Frequency of CD4+ and CD8+ T cells affected by Moderate Exercise. Bar graphs represent the frequency of CD4⁺ and CD8⁺ T cells in the Moderate⁺ (exercise) and Moderate⁻ (no exercise) groups. * p < 0.05.

There were no significant differences between the two groups in CD4+ T cell subpopulations (Figure 7 A – D). In the CD8+ T cell population, however, CD45RA+CD27- CD8+ TEMRA cells in the exercise group were three-fold lower compared to the no exercise group (10.40% vs. 3.17%, p = 0.0350). The frequencies of the remaining CD45RA+CD27+ naïve, CD45A-CD27+ CM, and CD45RA-CD27- EM CD8+ T cells were similar across groups (Figure 7 A- H, supplementary table 8). Therefore, the exercise group had a significantly lower frequency of senescence related CD8+ TEMRA cells.

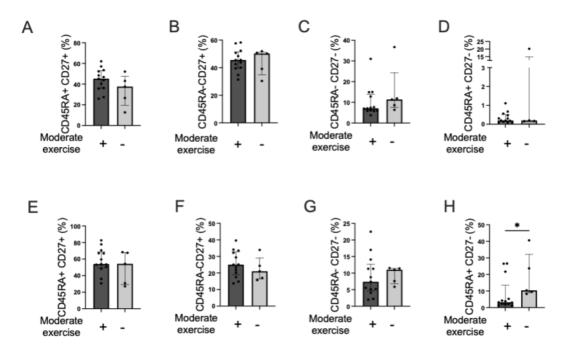


Figure 7. CD4 and CD8 T cell subpopulations in patients with JIA grouped based on self-reported exercise habits. Bar graphs represent the frequency of CD8⁺ EMRA CD45RA⁺CD27⁻ T cells (A), naïve CD45RA⁺CD27⁺ T cells (B), central memory (CM) CD45RA⁻CD27⁺ T cells (C), and effector memory (EM) CD45RA⁻CD27⁻ T cells (D) in the Moderate⁺ and Moderate⁻ groups. * p < 0.05.

NK cells

Previous reports significant alterations in NK cells in both JIA and exercise (37, 50). However, we found no significant differences in NK cell subpopulations based on self-reported exercise.

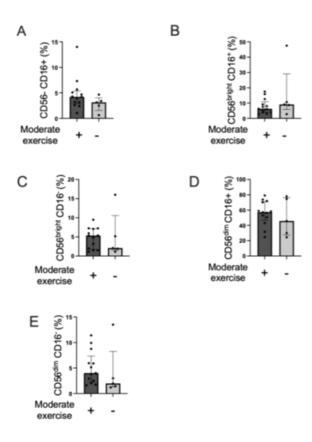


Figure 8. NK cell subpopulations in patients with JIA grouped based on self-reported exercise habits. Bar graphs represent the frequency of CD56⁻CD16⁻ NK cells (A), CD56brightCD16⁺ NK cells (B), CD56brightCD16⁻ NK cells (C), CD56dimCD16⁺ NK cells (D) and CD56dimCD16⁻ NK cells (E) in the Moderate + group (participants who perform moderate exercise) and Moderate - group (participants with no moderate exercise).

Discussion

Considering the dynamic interactions between the skeletal muscle and the immune system in health and disease (1, 10, 51), a key aim of this study was to initiate the exploration of the relationship between musculoskeletal function and peripheral blood immune profile in young people with JIA.

Limitations

Our study has several limitations. Fist, this was a single centre observational study that was not powered to account for the potential impact of factors other than the diagnosis of JIA and

exercise habits such as medications including corticosteroids and biologics that could influence peripheral blood immune profile. Second, participants were recruited in outpatient settings and did not have high disease activity, therefore, these findings may not apply to patients with acute high disease activity. Third, we have not evaluated immune profile of internal organs such as the joint and muscle. Fourth, SARC-F score, but not comprehensive measurements of muscle mass, strength etc., was used as a measure of musculoskeletal function. Finally, the cryopreservation process of PBMCs may differentially affect cell viability and enrich for certain cell types. The discussion that follows considers these limitations.

Alterations in immune profile during ageing (52) and exercise are well described (40). In this study, the SARC-F^{hi}+ group had fewer peripheral naïve CD4+ T cells as compared to the SARC-F^{hi}- group and trends towards an association between musculoskeletal function and increased frequency of CD4+ and CD8+ TEMRA cells were notable. TEMRA cells are terminally differentiated effector memory T cells with the capacity to secrete cytotoxic and inflammatory molecules and cause nonspecific tissue damage accumulate in ageing (52) and JIA (24) and exercise is associated with reduced frequency (38, 40). These findings support previous reports of T cell immunosenescence in JIA (24) with add evidence of a relationship with poor musculoskeletal function, thereby providing the rationale for modifying lifestyle such as exercise (40).

Our study is the first to report, in JIA on the relationship between peripheral CD4+ T cells and musculoskeletal functional impairment. Participants with musculoskeletal functional impairment had fewer circulating CD56-CD16+ NK cells compared to those without. CD56-CD16+ NK cells have primarily been studied in the context of chronic infectious diseases (53). There is a lack of literature on the role of CD56-CD16+ NK cells in JIA in the context of interactions with skeletal muscle. This study raises the question of how the bidirectional relationship between skeletal muscle and NK cells may change with skeletal muscle functionality in chronic inflammation.

This study has suggested that exercise can impact peripheral T cells in JIA. The exercise group had significantly increased total CD4+ T cells, however, CD4+ T cell subpopulations did not differ between the groups. The difference in total CD8+ T cell proportion was insignificant, however, the exercise group had three-fold lower CD8+ TEMRA cells compared to the non-exercise group. Few studies have examined the impact of exercise on the peripheral immune profile in JIA patients, especially those with poor musculoskeletal function. Our findings indicate that participants with poor musculoskeletal function may exhibit immune profile features similar to ageing adults, particularly in naïve and TEMRA cell

frequencies (54). Collectively, these observations suggest that exercise may influence the peripheral T cell pool in a manner that slows or prevents TEMRA cell accumulation in JIA..

Future perspectives

This study highlights significant variations in immune profiles related to musculoskeletal function and exercise in young adults with JIA. Future studies overcoming the limitations of our study are warranted to improve understanding of sarcopenia and peripheral blood immune profile in JIA, particularly focusing on moderate exercise's impact on total CD4⁺ populations and CD8⁺ TEMRA cells. Furthermore, investigating other factors such as dietary habits and vitamin D status may inform strategies to promote skeletal muscle function and restore a favourable immune profile.

Conclusions

Sarcopenia is a disorder of skeletal muscle affecting not only older adults but also young adults with inflammatory arthritis. This study demonstrates that JIA patients with musculoskeletal functional impairment have fewer naïve and greater effector memory CD4⁺ T cells, suggesting a relationship between musculoskeletal dysfunction and immunosenescence in this cohort. Further research into immune–muscle interactions in JIA may advance efforts to reverse sarcopenia and restore immune homeostasis.

Reference:

- 1. Nelke C, Dziewas R, Minnerup J, Meuth SG, Ruck T. Skeletal muscle as potential central link between sarcopenia and immune senescence. eBioMedicine. 2019;49:381-8.
- 2. Afzali AM, Müntefering T, Wiendl H, Meuth SG, Ruck T. Skeletal muscle cells actively shape (auto)immune responses. Autoimmunity Reviews. 2018.
- 3. Cruz-Jentoft AJ, Baeyens JP, Bauer JM, Boirie Y, Cederholm T, Landi F, et al. Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. Age and Ageing. 2010;39(4):412-23.
- 4. Cruz-Jentoft A, Bahat G, Bauer J, Boirie Y, Bruyère O, Cederholm T, et al. Sarcopenia: revised European consensus on definition and diagnosis. Age and Ageing. 2019.
- 5. Anker SD, Morley JE, von Haehling S. Welcome to the ICD-10 code for sarcopenia. J Cachexia Sarcopenia Muscle. 2016;7(5):512-4.

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- 6. Veronese N, Demurtas J, Soysal P, Smith L, Torbahn G, Schoene D, et al. Sarcopenia and health-related outcomes: an umbrella review of observational studies. Eur Geriatr Med. 2019;10(6):853-62.
- 7. Bano G, Trevisan C, Carraro S, Solmi M, Luchini C, Stubbs B, et al. Inflammation and sarcopenia: A systematic review and meta-analysis. Maturitas. 2017.
- 8. Li TH, Chang YS, Liu CW, Su CF, Tsai HC, Tsao YP, et al. The prevalence and risk factors of sarcopenia in rheumatoid arthritis patients: A systematic review and meta-regression analysis. Semin Arthritis Rheum. 2021;51(1):236-45.
- 9. Bennett JL, Pratt A, Dodds R, Sayer A, Isaacs J. Rheumatoid sarcopenia: loss of skeletal muscle strength and mass in rheumatoid arthritis. Nature Reviews Rheumatology. 2023.
- 10. Nelke C, Kleefeld F, Preusse C, Ruck T, Stenzel W, Nelke C, et al. Inclusion body myositis and associated diseases: an argument for shared immune pathologies. 2022.
- 11. Shen Y, Shi Q, Nong K, Li S, Yue J, Huang J, et al. Exercise for sarcopenia in older people: A systematic review and network meta-analysis. J Cachexia Sarcopenia Muscle. 2023;14(3):1199-211.
- 12. Bilski J, Pierzchalski P, Szczepanik M, Bonior J, Zoladz JA. Multifactorial Mechanism of Sarcopenia and Sarcopenic Obesity. Role of Physical Exercise, Microbiota and Myokines. Cells. 2022;11(1).
- 13. Ravelli A, Martini A. Juvenile idiopathic arthritis. Lancet. 2007;369(9563):767-78.
- 14. Ostring G, Singh-Grewal D. Juvenile Idiopathic Arthritis in the New World of Biologics. Journal of Paediatrics and Child Health. 2013.
- 15. Brabnikova Maresova K, Jarosova K, Pavelka K, Stepan JJ. The association between lean mass and bone mineral content in the high disease activity group of adult patients with juvenile idiopathic arthritis. BMC Musculoskeletal Disorders. 2014;15(1):51.
- 16. Marešová KB, Jarošová K, Pavelka K, Stepan JJ. The Association Between Lean Mass and Bone Mineral Content in the High Disease Activity Group of Adult Patients With Juvenile Idiopathic Arthritis. BMC Musculoskeletal Disorders. 2014.
- 17. Stagi S, Masi L, Capannini S, Cimaz R, Tonini G, Matucci-Cerinic M, et al. Cross-sectional and Longitudinal Evaluation of Bone Mass in Children and Young Adults with Juvenile Idiopathic Arthritis: The Role of Bone Mass Determinants in a Large Cohort of Patients. The Journal of Rheumatology. 2010.
- 18. Lien G, Flatø B, Haugen M, Vinje O, Sørskaar D, Dale K, et al. Frequency of osteopenia in adolescents with early-onset juvenile idiopathic arthritis: a long-term outcome study of one hundred five patients. Arthritis and rheumatism. 2003.

- 19. Maresova KB. Secondary Osteoporosis in Patients with Juvenile Idiopathic Arthritis. Journal of osteoporosis. 2011.
- 20. Kulyk M, Dzhus M. Assessment of sarcopenia in young patients with inflammatory arthritis: a cross-sectional study. Scientific Reports. 2025;15(1):8236.
- 21. Goronzy JJ, Weyand CM. Aging, autoimmunity and arthritis: T-cell senescence and contraction of T-cell repertoire diversity catalysts of autoimmunity and chronic inflammation. Arthritis research & therapy. 2003;5(5):225-34.
- 22. Weyand CM, Goronzy JJ. Immunometabolism in the development of rheumatoid arthritis. Immunol Rev. 2020;294(1):177-87.
- 23. Akbar AN, Henson SM, Lanna A. Senescence of T Lymphocytes: Implications for Enhancing Human Immunity. Trends Immunol. 2016;37(12):866-76.
- 24. Dvergsten JA, Mueller RG, Griffin P, Abedin S, Pishko A, Michel JJ, et al. Premature cell senescence and T cell receptor-independent activation of CD8+ T cells in juvenile idiopathic arthritis. Arthritis Rheum. 2013;65(8):2201-10.
- 25. Malmstrom TK, Miller DK, Simonsick EM, Ferrucci L, Morley JE. SARC-F: a symptom score to predict persons with sarcopenia at risk for poor functional outcomes. J Cachexia Sarcopenia Muscle. 2016;7(1):28-36.
- 26. Malmstrom TK, Morley JE. SARC-F: a simple questionnaire to rapidly diagnose sarcopenia. J Am Med Dir Assoc. 2013;14(8):531-2.
- 27. Voelker SN, Michalopoulos N, Maier AB, Reijnierse EM. Reliability and Concurrent Validity of the SARC-F and Its Modified Versions: A Systematic Review and Meta-Analysis. Journal of the American Medical Directors Association. 2021.
- 28. Woo J, Leung J, Morley JE. Validating the SARC-F: a suitable community screening tool for sarcopenia? J Am Med Dir Assoc. 2014;15(9):630-4.
- 29. Tsuji H, Tetsunaga T, Tetsunaga T, Misawa H, Oda Y, Takao S, et al. Evaluation of SARC-F and SARC-CalF for Sarcopenia Screening in Patients With Chronic Musculoskeletal Pain: A Prospective Cross-Sectional Study. Medicine. 2022.
- 30. Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. Nature Reviews Endocrinology. 2012.
- 31. McLean RR, Mangano KM, Hannan MT, Kiel DP, Sahni S. Dietary Protein Intake Is Protective Against Loss of Grip Strength Among Older Adults in the Framingham Offspring Cohort. J Gerontol A Biol Sci Med Sci. 2016;71(3):356-61.
- 32. Scott D, Blizzard L, Fell J, Giles G, Jones G. Associations between dietary nutrient intake and muscle mass and strength in community-dwelling older adults: the Tasmanian Older Adult Cohort Study. J Am Geriatr Soc. 2010;58(11):2129-34.

- 33. Welch AA, Skinner J, Hickson M. Dietary Magnesium May Be Protective for Aging of Bone and Skeletal Muscle in Middle and Younger Older Age Men and Women: Cross-Sectional Findings from the UK Biobank Cohort. Nutrients. 2017;9(11).
- 34. Boßlau TK, Wasserfurth P, Reichel T, Weyh C, Palmowski J, Nebl J, et al. 12-week combined strength and endurance exercise attenuates CD8(+) T-cell differentiation and affects the kynurenine pathway in the elderly: a randomized controlled trial. Immun Ageing. 2023;20(1):19.
- 35. Benatti FB, Pedersen BK. Exercise as an anti-inflammatory therapy for rheumatic diseases-myokine regulation. Nat Rev Rheumatol. 2015;11(2):86-97.
- 36. Hunt RM, Elzayat MT, Markofski MM, Laughlin M, LaVoy EC. Characterization of transitional memory CD4+ and CD8+ T-cell mobilization during and after an acute bout of exercise. Front Sports Act Living. 2023;5:1120454.
- 37. Woods JA, Evans JK, Wolters BW, Ceddia MA, McAuley E. Effects of maximal exercise on natural killer (NK) cell cytotoxicity and responsiveness to interferon-alpha in the young and old. J Gerontol A Biol Sci Med Sci. 1998;53(6):B430-7.
- 38. Donovan T, Bain AL, Tu W, Pyne DB, Rao S. Influence of Exercise on Exhausted and Senescent T Cells: A Systematic Review. Front Physiol. 2021;12:668327.
- 39. de Souza Teixeira AA, Minuzzi LG, Lira FS, Goncalves A, Martinho A, Rosa Neto JC, et al. Improvement in the anti-inflammatory profile with lifelong physical exercise is related to clock genes expression in effector-memory CD4+ T cells in master athletes. Exerc Immunol Rev. 2021;27:67-83.
- 40. Hanson ED, Bates LC, Bartlett DB, Campbell JP. Does exercise attenuate age- and disease-associated dysfunction in unconventional T cells? Shining a light on overlooked cells in exercise immunology. Eur J Appl Physiol. 2021;121(7):1815-34.
- 41. Arnold L, Henry A, Poron F, Baba-Amer Y, Van Rooijen N, Plonquet A, et al. Inflammatory monocytes recruited after skeletal muscle injury switch into antiinflammatory macrophages to support myogenesis. The Journal of experimental medicine. 2007.
- 42. Avolio E, Spinetti G, Madeddu P. Training monocytes by physical exercise: good practice for improving collateral artery development and postischemic outcomes. Arterioscler Thromb Vasc Biol. 2015;35(8):1733-5.
- 43. Steppich B, Dayyani F, Gruber R, Lorenz R, Mack M, Ziegler-Heitbrock HW. Selective mobilization of CD14(+)CD16(+) monocytes by exercise. Am J Physiol Cell Physiol. 2000;279(3):C578-86.
- 44. Timmerman KL, Flynn MG, Coen PM, Markofski MM, Pence BD. Exercise training-induced lowering of inflammatory (CD14+CD16+) monocytes: a role in the anti-inflammatory influence of exercise? J Leukoc Biol. 2008;84(5):1271-8.

- 45. Langston PK, Sun Y, Ryback BA, Mueller AL, Spiegelman BM, Benoist C, et al. Regulatory T cells shield muscle mitochondria from interferon-gamma-mediated damage to promote the beneficial effects of exercise. Sci Immunol. 2023;8(89):eadi5377.
- 46. Petty RE, Southwood TR, Manners P, Baum J, Glass DN, Goldenberg J, et al. International League of Associations for Rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton, 2001. J Rheumatol. 2004;31(2):390-2.
- 47. Fragala MS, Kraemer WJ, Mastro AM, Denegar CR, Volek JS, Kupchak BR, et al. Glucocorticoid receptor expression on human B cells in response to acute heavy resistance exercise. Neuroimmunomodulation. 2011;18(3):156-64.
- 48. Morbach H, Girschick HJ. Do B cells play a role in the pathogenesis of juvenile idiopathic arthritis? Autoimmunity. 2009;42(4):373-5.
- 49. de Almeida-Neto PF, Goncalves CAM, Wilde P, Jaggers JR, Junior GBC, de Farias Sales VS, et al. Influence of age and fitness level on immune responses of T and NK cells in healthy physically active subjects after strenuous aerobic exercise: a cross-sectional study. Front Immunol. 2023;14:1252506.
- 50. Dobloug JH, Forre O, Kvien TK, Egeland T, Degre M. Natural killer (NK) cell activity of peripheral blood, synovial fluid, and synovial tissue lymphocytes from patients with rheumatoid arthritis and juvenile rheumatoid arthritis. Ann Rheum Dis. 1982;41(5):490-4.
- 51. Dodds RM, Granic A, Robinson SM, Sayer AA. Sarcopenia, long-term conditions, and multimorbidity: findings from UK Biobank participants. J Cachexia Sarcopenia Muscle. 2020;11(1):62-8.
- 52. Lai W, Feng Q, Lei W, Xiao C, Wang J, Zhu Y, et al. Deciphering Immunosenescence From Child to Frailty: Transcriptional Changes, Inflammation Dynamics, and Adaptive Immune Alterations. Aging Cell. 2025:e70082.
- 53. Cocker ATH, Guethlein LA, Parham P. The CD56-CD16+ NK cell subset in chronic infections. Biochem Soc Trans. 2023;51(3):1201-12.
- 54. Xu Y, Wang Z, Li S, Su J, Gao L, Ou J, et al. An in-depth understanding of the role and mechanisms of T cells in immune organ aging and age-related diseases. Sci China Life Sci. 2025;68(2):328-53.