

ScienceDirect



Interactions between homeostatic plasticity and statistical learning: A role for inhibition

Elisa Galliano¹ and Tara Keck²



Statistical learning, sensory-driven unsupervised learning of repeating patterns, must coexist with ongoing homeostatic plasticity that is responsible for the necessary balance of activity in the brain; however, the mechanisms that facilitate these interactions are not clear. While models of both statistical learning, a form of associative plasticity, and homeostatic plasticity have primarily focused on excitatory cells and their synaptic changes, inhibition may play a key role in facilitating the balance between homeostatic plasticity and statistical learning. Here, we review the inhibitory synaptic, cellular, and network mechanisms underlying homeostatic and associative plasticity in rodents and propose a model in which localized inhibition, provided by diverse interneuron types, supports both statistical learning and homeostatic plasticity, as well as the interactions between them.

Addresses

- Department of Physiology, Development and Neuroscience, University of Cambridge, UK
- Department of Neuroscience, Physiology and Pharmacology, University College London, London WC1E 6DE, UK

Corresponding authors: Galliano, Elisa (eg542@cam.ac.uk); Keck, Tara (t.keck@ucl.ac.uk)

Current Opinion in Neurobiology 2025, 93:103065

This review comes from a themed issue on **Systems Neuroscience** 2025

Edited by Robert Froemke, Máté Lengyel, József Fiser and Livia de Hoz

For a complete overview see the Issue and the Editorial

Available online xxx

https://doi.org/10.1016/j.conb.2025.103065

0959-4388/© 2025 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Homeostatic plasticity ensures neural activity remains within a functional range over extended periods of time, a requirement of all circuits which is perhaps even more critical in sensory areas, where environmental stimuli constantly change across large orders of magnitude [1]. Without homeostatic compensatory mechanisms, systems become inherently unstable, resulting in either runaway activity or a quiescent circuit [2]. These essential homeostatic mechanisms must also act in

conjunction with forms of associative and Hebbian plasticity that underlie learning, memory formation, and functional reorganization after changes in the sensory periphery. The combination of these plasticity mechanisms allows for flexibility in the circuit to encode novel experiences (associative plasticity) while avoiding extreme activity levels (homeostatic plasticity). One important form of associative learning is statistical learning-sensory-driven, unsupervised learning of statistical patterns of sensory inputs—which, by definition, relies on repeated presentations of the same stimuli. Consequently, it is likely slower than reinforcement or one-shot learning, suggesting there may be a convergence of timescales between statistical learning and the learning-induced homeostatic plasticity that it must coexist with. However, how statistical learning and homeostatic plasticity integrate to enable changes to the system while maintaining activity balance without hindering one another is not fully clear.

Experiments and models of both statistical learning and homeostatic plasticity in adult rodents have traditionally emphasized changes at excitatory synapses, particularly through mechanisms like synaptic scaling, and Hebbian long-term potentiation (LTP) and long-term depression (LTD) [1,3,4]. However, methodological advances in genetically targeting specific inhibitory interneuron subtypes for activity manipulation and plasticity readouts have enabled novel experiments revealing their role in these forms of plasticity. Focusing on the rodent brain, here we review inhibitory plasticity mechanisms that are thought to support homeostasis, as well as associative and statistical learning, and propose that differential inhibitory plasticity processes may help mediate their interactions.

Inhibitory plasticity mechanisms: homeostasis

Plasticity of inhibitory interneurons and inhibitory synapses has been shown to play a crucial, yet often underemphasized, role in homeostatic plasticity [5–7]. Traditionally induced by transient loss or overrepresentation of inputs (e.g. via surgical, chemical, or mechanical manipulation of the peripheral sensor; environmental enrichment; or the pharmacological or chemogenetic manipulation of local circuits) and studied as part of maintaining the critical balance between

excitation and inhibition at the subcellular, cellular, and network levels [5,8], inhibitory plasticity can occur in a wide variety of ways.

At the level of synapses, the strength and number of inhibitory synapses onto excitatory neurons can undergo homeostatic changes in response to changes in sensory input via synaptic scaling-like mechanisms or Hebbian mechanisms such as shifting the LTP/LTD threshold for plasticity induction [4,9-12]. Either of these mechanisms could result in altered levels of inhibition onto these principal cells. Additionally, the excitatory and inhibitory synapses onto inhibitory neurons themselves can change as a result of changes to sensory input [13-15], which could also alter inhibitory cell activity due to increases or decreases in synaptic drive. Network-level activity is influenced by these changes in activity levels of individual inhibitory neurons or in the number of inhibitory neurons in olfactory and hippocampal circuits via adult neurogenesis [14,16-18]. Previous studies have shown that inhibitory neurons regulate small, local cortical networks through inhibition stabilization. where strong reciprocal excitatory-inhibitory maintain connections help balanced activity levels and provide a fast-acting form of homeostatic regulation [19–21].

A nonsynaptic mechanism that may underlie changes in activity levels in inhibitory neurons themselves is the modulation of intrinsic excitability, which is well suited to rapidly change activity levels [22]. Changes in excitability in inhibitory neurons has been shown to occur as a homeostatic response to decreased input in somatosensory [23], auditory [24], visual [25], and olfactory [26] cortical and subcortical areas. Because of its speed of implementation, sensitivity to changes in inputs including neuromodulatory inputs [27,28], and effectiveness in adjusting the output of the entire neuron, plasticity of intrinsic excitability can be highly effective at controlling homeostatic inhibition and disinhibition in individual inhibitory neurons and therefore also in the network.

Moreover, there are a number of different inhibitory subtypes, including (but not limited to) parvalbumin (PV), somatostatin (SOM), vasoactive intestinal peptide (VIP)-, calretinin- (CR), and calbindin- (CB) positive neurons, as well as dual releasers of GABA and monoamines or neuromodulators [29,30]. Different inhibitory subtypes are known to target other inhibitory subtypes, which can facilitate disinhibitory activity, as well as different parts of the dendritic tree, the soma, or the axon initial segment (AIS) of principal cells [31–34]. Given that homeostatic regulation can occur at the AIS [26,35], within dendrites [13,36], at cells [37], and in small networks [19], activity changes in inhibitory subtypes targeting any of these spatial scales could facilitate homeostatic plasticity. Because of their different postsynaptic targets, activity changes in different subtypes would have drastically different effects on overall activity, depending on the nature of their connections.

Inhibitory plasticity mechanisms: learning and associative plasticity

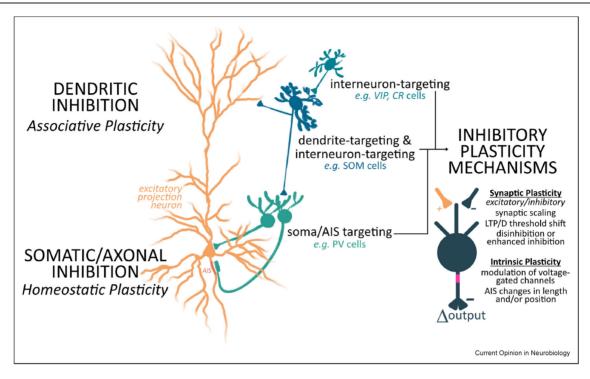
Inhibition is also proposed to play an important role in plasticity induction associated with learning. In both human [38] and animal models [39], inhibition has been proposed to gate learning, with a reduction in inhibition levels being associated with increased levels of learning and associative plasticity at excitatory synapses. Similar observations have been made for homeostatic plasticity occurring following the loss of peripheral input, with a reduction of inhibitory levels preceding excitatory synaptic plasticity proposed to be associated with fast compensation and functional remapping [6,40]. These permissive homeostatic changes in inhibition include reduced excitatory input onto inhibitory neurons [40,41], decreased turnover of adult-born inhibitory neurons [42], and reduced inhibitory inputs onto excitatory neurons [40,43,44], all of which lower inhibitory synaptic drive onto excitatory neurons. While historically there have been fewer studies mechanistically linking inhibitory plasticity with behavioral-level associative learning, more recent work has started to strongly link changes in plasticity in inhibitory cells with disinhibition of principal neurons and the overall network [45–48]. Supporting this idea, it has been widely demonstrated that following sensory changes and learning induction, inhibitory plasticity precedes excitatory plasticity, with inhibitory changes acting on a faster time course [25,49].

The consistency of these results across different plasticity induction paradigms suggests that this may be a general principle. Therefore, reducing inhibition, whether homeostatically or not, through changes to inhibitory synapses and neurons may also facilitate statistical learning, though this remains untested. Furthermore, different inhibitory subtypes have been shown to have unique plasticity profiles and play different roles in circuit computation and reinforcement learning [50-52]; however, the specificity of their plasticity in statistical learning remains unexplored.

A potential role for inhibition in homeostasis and statistical learning

While inhibitory homeostatic plasticity is well studied at the molecular and cellular level, little is known about how inhibition mechanistically supports statistical learning. Thus, we venture to extrapolate from the existing literature and propose a deliberately speculative hypothesis on how statistical learning, inhibition, and homeostasis may intersect. One possibility is that different inhibitory subtypes mediate homeostatic

Figure 1



Schematic representation of the working hypothesis that localized inhibitory plasticity, mediated by diverse plasticity mechanisms, may enable the synergistic coexistence of associative plasticity and statistical learning with maintaining network homeostasis. The indicated examples of neuronal subtypes are found in the cortex and hippocampus, but a similar circuit design—albeit with different cell names and makers—applies broadly across the brain. AIS, axon initial segment; VIP, vasoactive intestinal peptide; CR, calretinin; SOM: somatostatin; PV, parvalbumin; LTP/LTD, long-term potentiation/ long-term depression.

plasticity and statistical learning respectively (Figure 1), which would support these two complementary forms of plasticity occurring in tandem, while minimizing conflict. For example in the cortex, cell- and network-level homeostatic balance could be regulated through changes to interneurons, such as PV cells, that typically target the excitatory cells' soma [32]. PV cells have been shown to increase their selectivity to match that of excitatory pyramidal cells during visual reinforcement learning [50], with PV-pyramidal cell coupling potentially having a stabilizing effect on network activity. Similar changes in other brain areas could also entrain soma-targeting interneurons to strengthen coupling with excitatory cells during learning-related increases in activity, which could in theory help balance network activity via inhibitory—excitatory reciprocal interactions. Additionally, a subset of PV cells, *i.e.* corticohippocampal chandelier cells and cerebellar basket cells, are known to target the AIS of pyramidal/Purkinje cells, which affects the action potential threshold and as a result, cell and network activity [53]. Changing inhibition levels onto the AIS would provide another PV-specific way in which cellular and network activity can be homeostatically regulated [54]. Complementing PV plasticity, a reduction of inhibition from interneurons targeting the

dendrites of excitatory neurons, for example, bulbar granule cells and cortical SOM cells [32,55], could be achieved either directly or indirectly through increased inhibitory drive from VIP cells onto SOM cells [56]. This reduction could gate the associative or Hebbian synaptic plasticity that is proposed to underlie statistical learning, similar to what has been observed for other forms of learning and plasticity [40,50].

We propose this spatial compartmentalization of regulation of excitatory cells by different inhibitory subtypes may facilitate complementary statistical learning and homeostatic plasticity, with dendrite-targeting inhibitory neurons gating excitatory plasticity in local compartments, as has been observed in learning and functional reorganization [39,40,43,44], and PV-like inhibitory neurons homeostatically regulating activity levels at the soma or AIS, affecting cellular and network activity [52,54]. While these two forms of plasticity are unlikely to be strictly restricted to these spatial compartments, this framework may generally provide a mechanism by which these forms of plasticity coexist. Furthermore, these homeostatic changes in inhibition are unlikely to be operating in isolation and likely occur in tandem with other homeostatic mechanisms in excitatory neurons, such as intrinsic excitability modulation, synaptic scaling, and shifting the threshold for LTP/LTD induction. These mechanisms have been discussed elsewhere and have been shown to globally regulate synaptic strength while allowing for localized associative or Hebbian plasticity that are thought to underlie learning [1,3].

While relatively few experimental studies have examined the role of inhibition in statistical learning, many associative learning and statistical learning models do contain inhibitory layers [57,58]. Furthermore, numerous models have demonstrated that inhibition stabilization is a key feature in the cortex, with recurrent networks of excitatory and inhibitory cells showing strong coupling that creates stable activity levels independent of the level of sensory stimulation [20,59,60]. To date, models of statistical learning or homeostasis have not typically incorporated spatially compartmentalized inhibitory inputs that would be associated with particular inhibitory subtypes. A model of statistical learning that includes localized inhibition, along with homeostatic compensatory components including direct modulation of intrinsic excitability, could be a critical first step for testing the role of inhibitory subtypes in statistical learning, homeostasis, and the interactions between the two.

Funding

E.G. was supported by URKI Biotechnology and Biological Sciences Research Council (BB\W014688\1) and Wellcome Trust (301427/Z/23/Z); T.K. was supported by Wellcome Trust (212264/Z/18/Z), BIRAX and The Royal Society. This research was also supported by the National Science Foundation under Grant NSF PHY-1748958.

Declaration of competing interest

The authors have none to declare.

Data availability

No data was used for the research described in the article.

References

Papers of particular interest, published within the period of review, have been highlighted as:

- * of special interest
- ** of outstanding interest
- Wen W, Turrigiano GG: Keeping your brain in balance: homeostatic regulation of network function. Annu Rev Neurosci 2024, 47:41–61, https://doi.org/10.1146/annurev-neuro-092523-110001.
- Litwin-Kumar A, Doiron B: Formation and maintenance of neuronal assemblies through synaptic plasticity. Nat Commun 2014, 5:5319, https://doi.org/10.1038/ncomms6319.
- 3. Keck T, Toyoizumi T, Chen L, Doiron B, Feldman DE, Fox K, Gerstner W, Haydon PG, Hübener M, Lee H-K, Lisman JE,

- Rose T, Sengpiel F, Stellwagen D, Stryker MP, Turrigiano GG, van Rossum MC: Integrating Hebbian and homeostatic plasticity: the current state of the field and future research directions. *Philos Trans R Soc Lond B Biol Sci* 2017, **372**, https://doi.org/10.1098/rstb.2016.0158.
- Keck T, Hübener M, Bonhoeffer T: Interactions between synaptic homeostatic mechanisms: an attempt to reconcile BCM theory, synaptic scaling, and changing excitation/inhibition balance. Curr Opin Neurobiol 2017, 43:87–93, https://doi.org/10.1016/j.conb.2017.02.003. Neurobiology of Learning and Plasticity.
- Chen L, Li X, Tjia M, Thapliyal S: Homeostatic plasticity and excitation-inhibition balance: the good, the bad, and the ugly. *Curr Opin Neurobiol* 2022, 75:102553, https://doi.org/10.1016/ j.conb.2022.102553.
- Huang L, Hardyman F, Edwards M, Galliano E: Deprivationinduced plasticity in the early central circuits of the rodent visual, auditory, and olfactory systems. eNeuro 2024, 11, https://doi.org/10.1523/ENEURO.0435-23.2023. ENEURO.0435-23.2023.
- McFarlan AR, Chou CYC, Watanabe A, Cherepacha N, Haddad M, Owens H, Sjöström PJ: The plasticitome of cortical interneurons. Nat Rev Neurosci 2023, 24:80–97, https://doi.org/ 10.1038/s41583-022-00663-9.
- lascone DM, Li Y, Sümbül U, Doron M, Chen H, Andreu V, Goudy F, Blockus H, Abbott LF, Segev I, Peng H, Polleux F: Whole-neuron synaptic mapping reveals spatially precise excitatory/inhibitory balance limiting dendritic and somatic spiking. Neuron 2020, 106:566–578.e8, https://doi.org/10.1016/ i.neuron.2020.02.015.
- Bienenstock EL, Cooper LN, Munro PW: Theory for the development of neuron selectivity: orientation specificity and binocular interaction in visual cortex. J Neurosci 1982, 2: 32–48, https://doi.org/10.1523/JNEUROSCI.02-01-00032.1982.
- Keck T, Keller GB, Jacobsen RI, Eysel UT, Bonhoeffer T, Hübener M: Synaptic scaling and homeostatic plasticity in the mouse visual cortex in vivo. Neuron 2013, 80:327–334, https:// doi.org/10.1016/j.neuron.2013.08.018
- Lau CG, Murthy VN: Activity-dependent regulation of inhibition via GAD67. J Neurosci 2012, 32:8521–8531, https://doi.org/10.1523/JNEUROSCI.1245-12.2012.
- Whiting B, Moiseff A, Rubio ME: Cochlear nucleus neurons redistribute synaptic AMPA and glycine receptors in response to monaural conductive hearing loss. Neuroscience 2009, 163:1264-1276, https://doi.org/10.1016/ j.neuroscience.2009.07.049.
- Barnes SJ, Franzoni E, Jacobsen RI, Erdelyi F, Szabo G, Clopath C, Keller GB, Keck T: Deprivation-induced homeostatic spine scaling in vivo is localized to dendritic branches that have undergone recent spine loss. Neuron 2017, 96: 871–882.e5, https://doi.org/10.1016/j.neuron.2017.09.052.
- Denizet M, Cotter L, Lledo P-M, Lazarini F: Sensory deprivation increases phagocytosis of adult-born neurons by activated microglia in the olfactory bulb. Brain Behav Immun 2017, 60: 38–43, https://doi.org/10.1016/j.bbi.2016.09.015.
- Hartman KN, Pal SK, Burrone J, Murthy VN: Activity-dependent regulation of inhibitory synaptic transmission in hippocampal neurons. *Nat Neurosci* 2006, 9:642–649, https://doi.org/ 10.1038/nn1677.
- Li H, Tamura R, Hayashi D, Asai H, Koga J, Ando S, Yokota S,
 Kaneko J, Sakurai K, Sumiyoshi A, Yamamoto T, Hikishima K, Tanaka KZ, McHugh TJ, Hisatsune T: Silencing dentate newborn neurons alters excitatory/inhibitory balance and impairs behavioral inhibition and flexibility. Sci Adv 2024, 10, eadk4741.

This study shows that, in the rodent hippocampus, silencing adult-born granule cells with genetically-engineered tetanus toxin results in impaired behavioral learning and reveals their essential role in maintaining E/I balance in the network.

 Resnik J, Polley DB: Fast-spiking GABA circuit dynamics in the auditory cortex predict recovery of sensory processing following peripheral nerve damage. eLife 2017, 6, e21452, https://doi.org/10.7554/eLife.21452.

- 18. Rochefort C, Gheusi G, Vincent J-D, Lledo P-M: Enriched odor exposure increases the number of newborn neurons in the adult olfactory bulb and improves odor memory. J Neurosci 2002, 22:2679-2689.
- 19. Barnes SJ, Sammons RP, Jacobsen RI, Mackie J, Keller GB, Keck T: Subnetwork-specific homeostatic plasticity in mouse visual cortex in vivo. Neuron 2015, 86:1290-1303, https:// doi.org/10.1016/j.neuron.2015.05.010.
- Sanzeni A, Akitake B, Goldbach HC, Leedy CE, Brunel N, Histed MH: Inhibition stabilization is a widespread property of cortical networks. eLife 2020, 9, e54875, https://doi.org/10.75 eLife.54875.
- 21. Waitzmann F, Wu YK, Gjorgjieva J: Top-down modulation in canonical cortical circuits with short-term plasticity. Proc Natl Acad Sci U S A 2024, 121, e2311040121.

This study uses a combination of computer simulations and mathematical analyses to demonstrate a novel role for somatostatin inhibitory neurons in regulating network dynamics via short-term plasticity.

- Zeldenrust F, Calcini N, Yan X, Bijlsma A, Celikel T: The tuning of tuning: how adaptation influences single cell information transfer. PLoS Comput Biol 2024, 20, e1012043, https://doi.org/ 10.1371/journal.pcbi.1012043
- 23. Gainey MA, Aman JW, Feldman DE: Rapid disinhibition by adjustment of PV intrinsic excitability during whisker map plasticity in mouse S1. *J Neurosci* 2018, 38:4749–4761, https://doi.org/10.1523/JNEUROSCI.3628-17.2018.
- 24. Takesian AE, Kotak VC, Sanes DH: Age-dependent effect of hearing loss on cortical inhibitory synapse function. J Neurophysiol 2012, 107:937-947, https://doi.org/10.1152/ in.00515.2011.
- 25. Hengen KB, Lambo ME, Van Hooser SD, Katz DB, Turrigiano GG: Firing rate homeostasis in visual cortex of freely behaving rodents. Neuron 2013, 80:335-342, https:// doi.org/10.1016/j.neuron.2013.08.038
- 26. Galliano E, Hahn C, Browne LP, R Villamayor P, Tufo C, Crespo A, Grubb MS: Brief sensory deprivation triggers cell type-specific structural and functional plasticity in olfactory bulb neurons. *J Neurosci* 2021, 41:2135–2151, https://doi.org/ 10.1523/JNEUROSCI.1606-20.2020.
- Swanson OK, Maffei A: From hiring to firing: activation of inhibitory neurons and their recruitment in behavior. Front Mol Neurosci 2019, 12, https://doi.org/10.3389/fnmol.2019.00168.
- 28. Wester JC, McBain CJ: Behavioral state-dependent modulation of distinct interneuron subtypes and consequences for circuit function. Curr Opin Neurobiol 2014, 29:118-125, https:// doi.org/10.1016/j.conb.2014.07.007.
- Lim L, Mi D, Llorca A, Marín O: Development and functional diversification of cortical interneurons. Neuron 2018, 100: 294-313, https://doi.org/10.1016/j.neuron.2018.10.009.
- 30. Vaaga CE, Borisovska M, Westbrook GL: Dual-transmitter neurons: functional implications of co-release and co-transmission. Curr Opin Neurobiol 2014:25, https://doi.org/10.1016/ i.conb.2014.04.010 0.
- 31. Brown AM, Arancillo M, Lin T, Catt DR, Zhou J, Lackey EP, Stay TL, Zuo Z, White JJ, Sillitoe RV: Molecular layer interneurons shape the spike activity of cerebellar Purkinje cells. Sci Rep 2019, 9: 1742, https://doi.org/10.1038/s41598-018-38264-1.
- Callaway EM: Inhibitory cell types, circuits and receptive fields in mouse visual cortex. In Micro-, meso- and macroconnectomics of the brain. Edited by Kennedy H, Van Essen DC, Christen Y, Cham (CH): Springer; 2016.
- Pan-Vazquez A, Wefelmeyer W, Gonzalez Sabater V, Neves G, Burrone J: Activity-Dependent plasticity of axo-axonic synapses at the axon initial segment. Neuron 2020, https://doi.org/ 10.1016/i.neuron.2020.01.037.
- Zhao R, Ren B, Xiao Y, Tian J, Zou Y, Wei J, Qi Y, Hu A, Xie X, Huang ZJ, Shu Y, He M, Lu J, Tai Y: Axo-axonic synaptic input drives homeostatic plasticity by tuning the axon initial segment structurally and functionally. Sci Adv 2024, 10, eadk4331, https://doi.org/10.1126/sciadv.adk4331.

- This study demonstrates that homeostatic changes to the axon initial segment in the medial prefrontal cortex are initiated following chronic changes to inhibitory inputs from chandelier cells. These homeostatic changes are associated with recovery of social behaviors that are perturbed in the absence of homeostatic compensation.
- Jamann N, Dannehl D, Lehmann N, Wagener R, Thielemann C, Schultz C, Staiger J, Kole MHP, Engelhardt M: Sensory input drives rapid homeostatic scaling of the axon initial segment in mouse barrel cortex. Nat Commun 2021, 12:23, https:// doi.org/10.1038/s41467-020-20232-x.
- 36. Sutton MA, Schuman EM: Dendritic protein synthesis, synaptic plasticity, and memory. *Cell* 2006, **127**:49–58, https://doi.org/10.1016/j.cell.2006.09.014.
- 37. Burrone J, O'Byrne M, Murthy VN: Multiple forms of synaptic plasticity triggered by selective suppression of activity in individual neurons. Nature 2002, 420:414-418, https://doi.org/ 0.1038/nature01242.
- 38. Barron HC: Neural inhibition for continual learning and memory. Curr Opin Neurobiol 2021, 67:85-94, https://doi.org/ 10.1016/j.conb.2020.09.007. Neurobiology of Learning and Plasticity
- 39. Jeong N, Singer AC: Learning from inhibition: functional roles of hippocampal CA1 inhibition in spatial learning and memory. Curr Opin Neurobiol 2022, 76:102604, https://doi.org/ 10.1016/i.conb.2022.102604.
- Keck T, Scheuss V, Jacobsen RI, Wierenga CJ, Eysel UT, Bonhoeffer T, Hübener M: Loss of sensory input causes rapid structural changes of inhibitory neurons in adult mouse visual cortex. Neuron 2011, 71:869-882, https://doi.org/ 10.1016/i.neuron.2011.06.034.
- 41. Hamilton KA, Parrish-Aungst S, Margolis FL, Erdélyi F, Szabó G, Puche AC: Sensory deafferentation transsynaptically alters neuronal GluR1 expression in the external plexiform layer of the adult mouse main olfactory bulb. Chem Senses 2008, 33: 201-210, https://doi.org/10.1093/chemse/bjm079.
- Sawada M, Kaneko N, Inada H, Wake H, Kato Y, Yanagawa Y, Kobayashi K, Nemoto T, Nabekura J, Sawamoto K: **Sensory** input regulates spatial and subtype-specific patterns of neuronal turnover in the adult olfactory bulb. J Neurosci 2011, 31:11587-11596, https://doi.org/10.1523/JNEUROSCI.0614-11.2011.
- 43. Chen JL, Villa KL, Cha JW, So PTC, Kubota Y, Nedivi E: Clustered dynamics of inhibitory synapses and dendritic spines in the adult neocortex. Neuron 2012, 74:361-373, https:// doi.org/10.1016/j.neuron.2012.02.030.
- 44. van Versendaal D, Rajendran R, Saiepour MH, Klooster J, Smit-Rigter L, Sommeijer J-P, De Zeeuw CI, Hofer SB, Heimel JA, Levelt CN: Elimination of inhibitory synapses is a major component of adult ocular dominance plasticity. Neuron 2012, 74:374-383, https://doi.org/10.1016/j.neuron.2012.03.015.
- 45. Donato F, Rompani SB, Caroni P: Parvalbumin-expressing basket-cell network plasticity induced by experience regulates adult learning. *Nature* 2013, 504:272–276, https://doi.org/ 10.1038/nature12866.
- 46. Krabbe S, Gründemann J, Lüthi A: Amygdala inhibitory circuits regulate associative fear conditioning. *Biological Psychiat Mol Sign Stress Posttraum Stress Disorder* 2018, **83**:800–809, https://doi.org/10.1016/j.biopsych.2017.10.006.
- 47. Letzkus JJ, Wolff SBE, Meyer EMM, Tovote P, Courtin J, Herry C, Lüthi A: A disinhibitory microcircuit for associative fear learning in the auditory cortex. Nature 2011, 480:331-335, https://doi.org/10.1038/nature10674.
- Tzilivaki A, Tukker JJ, Maier N, Poirazi P, Sammons RP Schmitz D: Hippocampal GABAergic interneurons and memory. Neuron 2023, 111:3154-3175, https://doi.org/10.1016/
- 49. Richter LMA, Gjorgjieva J: A circuit mechanism for independent modulation of excitatory and inhibitory firing rates after sensory deprivation. Proc Natl Acad Sci 2022, 119, e2116895119, https://doi.org/10.1073/pnas.2116895119.

- Khan AG, Poort J, Chadwick A, Blot A, Sahani M, Mrsic-Flogel TD, Hofer SB: Distinct learning-induced changes in stimulus selectivity and interactions of GABAergic interneuron classes in visual cortex. *Nat Neurosci* 2018, 21: 851–859, https://doi.org/10.1038/s41593-018-0143-z.
- Kuchibhotla KV, Gill JV, Lindsay GW, Papadoyannis ES, Field RE, Sten TAH, Miller KD, Froemke RC: Parallel processing by cortical inhibition enables context-dependent behavior. Nat Neurosci 2017, 20:62–71, https://doi.org/10.1038/nn.4436.
- Lackey EP, Moreira L, Norton A, Hemelt ME, Osorno T,
 Nguyen TM, Macosko EZ, Lee W-CA, Hull CA, Regehr WG: Specialized connectivity of molecular layer interneuron subtypes leads to disinhibition and synchronous inhibition of cerebellar Purkinje cells. Neuron 2024, 112:2333, https://doi.org/10.1016/j.neuron.2024.04.010.

This study revisits the traditional classification of cerebellar molecular layer interneurons (MLIs) using RNA seq. They show that the two MLIs groups are differentially connected via both electrical and chemical synapses with each other and with different compartments of Purkinje cells. These connectivity profiles make them differentially suitable for synchronizing network activity, or for gating learning.

- Bender KJ, Trussell LO: The physiology of the axon initial segment. Annu Rev Neurosci 2012, 35:249–265, https://doi.org/ 10.1146/annurev-neuro-062111-150339.
- 54. Seignette K, Jamann N, Papale P, Terra H, Porneso RO, de ** Kraker L, van der Togt C, van der Aa M, Neering P, Ruimschotel E, Roelfsema PR, Montijn JS, Self MW, Kole MHP, Levelt CN: Experience shapes chandelier cell function and structure in the visual cortex. eLife 2024, 12, RP91153, https:// doi.org/10.7554/eLife.91153.

Using a combination of viral tracing, calcium imaging, and computational analysis, this study demonstrates that AIS-targeting chandelier cells in primary visual cortex respond less strongly and undergo structural plasticity of their axonal cartridges after repeated visual

- exposure. These plastic changes in the PV-expressing chandelier cells may gate AIS plasticity in excitatory neurons.
- Tufo C, Poopalasundaram S, Dorrego-Rivas A, Ford MC, Graham A, Grubb MS: Development of the mammalian main olfactory bulb. Development 2022, 149:dev200210, https:// doi.org/10.1242/dev.200210.
- Canto-Bustos M, Friason FK, Bassi C, Oswald A-MM: Disinhibitory circuitry gates associative synaptic plasticity in olfactory cortex. J Neurosci 2022, 42:2942–2950, https://doi.org/10.1523/JNEUROSCI.1369-21.2021.
- 57. Asabuki T, Clopath C: Embedding stochastic dynamics of the environment in spontaneous activity by prediction-based plasticity. *eLife* 2024, **13**, https://doi.org/10.7554/eLife.95243.2.
- Endress AD, Johnson SP: When forgetting fosters learning: a neural network model for statistical learning. Cognition, Special Issue in Honour of Jacques Mehler, Cognition's founding editor 2021, 213:104621, https://doi.org/10.1016/ j.cognition.2021.104621.
- 59. Agnes EJ, Vogels TP: Co-dependent excitatory and inhibitory plasticity accounts for quick, stable and long-lasting memories in biological networks. Nat Neurosci 2024, 27:964–974, https://doi.org/10.1038/s41593-024-01597-4.

This study uses modeling approaches to examine the interactions between excitatory and inhibitory synapses. They show how inhibition can gate excitatory plasticity and how interactions between excitatory and inhibitory synapses play an important role in maintaining the stability of synaptic weights.

Soldado-Magraner S, Seay MJ, Laje R, Buonomano DV: Paradoxical self-sustained dynamics emerge from orchestrated excitatory and inhibitory homeostatic plasticity rules. Proc Natl Acad Sci U S A 2022, 119, e2200621119, https://doi.org/10.1073/pnas.2200621119.