

STUDY PROTOCOL

Open Access



ARFID Genes and Environment (ARFID-GEN): study protocol

Cynthia M. Bulik^{1,2,3*}, Nadia Micali^{4,5,6}, Casey M. MacDermod¹, Baiyu Qi⁷, Melissa A. Munn-Chernoff^{1,8}, Laura M. Thornton¹, Jennifer White¹, Lisa Dinkler², Emily M. Pisetsky¹, Jessica Johnson^{1,9}, Katelin R. Devine¹, Shelby N. Ortiz¹, Ava E. Silverman^{1,10}, Natasha Berthold^{1,11,12}, Alexis Dumain^{1,13}, Jerry Guintivano^{1,9}, Matthew Halvorsen^{9,14} and James J. Crowley^{1,9,14}

Abstract

Background The Avoidant Restrictive Food Intake Disorder – Genes and Environment (ARFID-GEN) study is a study of genetic and environmental factors that contribute to risk for developing ARFID in children and adults.

Methods A total of 3,000 children and adults with ARFID from the United States will be included. Parents/guardians and their children with ARFID (ages 7 to 17) and adults with ARFID (ages 18+) will complete comprehensive online consent, parent verification of child assent (when applicable), and phenotyping. Enrolled participants with ARFID will submit a saliva sample for genotyping. A genome-wide association study of ARFID will be conducted.

Discussion ARFID-GEN, a large-scale genetic study of ARFID, is designed to rapidly advance the study of the genetics of eating disorders. We will explicate the genetic architecture of ARFID relative to other eating disorders and to other psychiatric, neurodevelopmental, and metabolic disorders and traits. Our goal is for ARFID to deliver “actionable” findings that can be transformed into clinically meaningful insights.

Trial registration ARFID-GEN is a registered clinical trial: clinicaltrials.gov NCT05605067.

Keywords Avoidant restrictive food intake disorder, Picky eating, Selective eating, Eating disorders, Genome-wide association, Psychiatric genetics, Psychiatric genomics consortium, Social media

*Correspondence:

Cynthia M. Bulik
cynthia_bulik@med.unc.edu

¹ Department of Psychiatry, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA

² Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, PO Box 281, 171 77 Stockholm, Sweden

³ Department of Nutrition, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, US

⁴ Center for Eating and Feeding Disorders Research, Psychiatric Center Ballerup, Mental Health Services of the Capital Region of Denmark, Ballerup, Denmark

⁵ Institute of Biological Psychiatry, Psykiatrisk Center Sct. Hans, Boserupvej 2, 4000 Roskilde, Denmark

⁶ Great Ormond Street Hospital Institute of Child Health, University College London, London, UK

⁷ Department of Epidemiology, University of North Carolina at Chapel Hill, Chapel Hill, NC 27516, USA

⁸ Texas Tech University, Lubbock, TX 79409, USA

⁹ Department of Genetics, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA

¹⁰ Smith College, Northampton, MA 01063, USA

¹¹ University of Western Australia, Crawley, WA 6009, Australia

¹² Perron Institute for Neurological and Translational Science, Nedlands, WA 6009, Australia

¹³ Department of Psychology and Neuroscience, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, US

¹⁴ Department of Clinical Neuroscience, Karolinska Institutet, Stockholm, Sweden



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

We describe the Avoidant Restrictive Food Intake Disorder – Genes and Environment (ARFID-GEN) study, which is designed to expand the discovery of genetic and environmental contributions to ARFID risk. ARFID-GEN builds on previous and ongoing genome-wide association studies (GWAS) by the Eating Disorders Working Group of the Psychiatric Genomics Consortium (PGC-ED) as part of a global effort to fully characterize the genetic architecture of all eating disorders (EDs) and explore their relation to each other and to other psychiatric, neurodevelopmental, and metabolic/anthropometric traits.

ARFID-GEN will ascertain, phenotype, and genotype a large sample of children and adults with ARFID. We will apply advanced analytic strategies to test and refine an etiological model of ARFID, explicate heterogeneity, and simultaneously document environmental risk factors for ARFID.

ARFID is associated with high personal and family emotional and financial cost. ARFID, present in 2–5% of the population [1], is marked by the avoidance and/or restriction of food intake resulting in significant weight loss or nutritional deficiency, dependence on feeding supplements, and/or interference with psychosocial functioning. Unlike other EDs, food restriction is not driven by weight and shape concerns, and ARFID may be equally common in males and females [1]. The Diagnostic and Statistical Manual of Mental Health Disorders 5th edition (DSM-5) [2] describes three predominant ARFID presentations that likely overlap [3]: (1) sensory sensitivity (i.e., rejection of food based on sensory qualities such as texture), (2) phobic avoidance of food (i.e., concern about aversive consequences of eating, such as fear of choking), and (3) low interest/appetite. Psychiatric, neurodevelopmental, and somatic medical comorbidities are common, with ~50% of ARFID cases having a co-occurring diagnosis [4].

Using existing Swedish twin data, we demonstrated that an ARFID phenotype is highly heritable (twin heritability estimate [h^2_{twin}] = 0.79; CI: 0.71, 0.86), with the remaining variance attributable to nonshared environmental factors [5]. This places ARFID amongst the most heritable psychiatric disorders and on par or higher than anorexia nervosa (AN) (0.50–0.60) [6], bulimia nervosa (BN) (0.50–0.60) [6], and binge-eating disorder (BED) (0.39–0.57) [7, 8]. These results support a GWAS for ARFID.

The three key dimensions of ARFID identified are viewed as symptom clusters rather than distinct presentations. The sensory dimension encompasses rejection of food based on sensory qualities (e.g., taste, smell, texture, temperature, appearance) and is the most common reason for referral [9]. When phobic avoidance is

the dominant presentation, conditioned aversion may play a role [10, 11]. A genetic predisposition to anxiety may increase the risk of children developing ARFID after an aversive feeding experience like gagging or witnessing someone choke or vomit [12]. The low interest/appetite dimension likely captures symptoms previously described as infantile anorexia and food avoidance emotional disorder [13].

Methods

Specific aims

Aim 1a. Ascertainment of 3,000 ARFID cases

Leveraging the existing Eating Disorders Genetics Initiative (EDGI) [14] infrastructure at the University of North Carolina at Chapel Hill (UNC), we will ascertain 3,000 children and adults with ARFID. Appropriate controls will be sourced from archived, genotyped repositories such as the National Institute of Mental Health (NIMH) Genomics Repository, database of Genotypes and Phenotypes (dbGaP). We will conduct efficient online phenotyping of children and adults with ARFID including environmental exposures and at-home saliva sampling for deoxyribose nucleic acid (DNA). We will genotype new samples using contemporary methodology.

Aim 1b. Validation substudy

We will interview parents/guardians of 25 children with ARFID and 25 adults with ARFID with the Pica, ARFID and Rumination Disorder Interview (PARDI) [15] to further validate our online ARFID diagnostic battery.

Aim 2. Within-disorder ARFID GWAS

We will conduct comprehensive phenotypic and genomic analyses: single nucleotide polymorphism (SNP)-based heritability, GWAS (imputable to minor allele frequency [MAF] ≥ 0.005), genetic correlations (r_g s), polygenic risk scores (PRS), standard post-GWAS analyses of the non-mutually exclusive ARFID presentations, and rare copy number variants (CNVs) and CNV burden. Hypotheses: We will identify genome-wide significant loci for ARFID, informative r_g s, implicated CNVs, and environmental precipitants.

Aim 3. Genetic relation of ARFID to other eating disorders

We will test if ARFID shares a core set of genetic factors with other EDs yet is differentiated by disorder-specific genetic factors. We will conduct a set of cross-disorder genomic analyses to map genetic interrelations between ARFID and other EDs including: (a) cross-disorder GWAS meta-analysis to identify loci with pleiotropic effects, (b) if indicated, calculate r_g s and conduct Mendelian Randomization (MR), multi-trait conditional

and joint analysis (mtCOJO) [16], disorder-specific SNP associations, Multi-PRS [17], and, (c) genomic structural equation modelling (GSEM) [18] to examine genome-wide architecture of ARFID relative to other EDs. Based on our preliminary data, we predict that ARFID will show the strongest genetic association with AN.

Aim 4. Genetic relation of ARFID with psychiatric, metabolic/anthropometric, neurodevelopmental, and other relevant phenotypes

To test a conceptualization that ARFID has specific genetic associations with psychiatric, metabolic/anthropometric, and neurodevelopmental phenotypes we will apply: (a) approaches as in Aim 3b, and (b) GSEM to examine genetic, psychiatric, neurodevelopmental, and metabolic/anthropometric factors associated with ARFID. We predict that high sensory sensitivity will be related to autism spectrum disorder (ASD), high levels of phobic avoidance to anxiety and obsessive-compulsive disorder (OCD), and high levels of low interest/appetite to AN. Deliverables: (a) dissection of converging and diverging relations among ARFID and other traits informing and refining its etiology; (b) genetic assessment of ARFID's relation to other phenotypes, informing nosology.

Participants

Objective

We will engage the infrastructure utilized in EDGI [14] to ascertain 3,000 children and adults with ARFID. Controls will be ascertained from data repositories from other genomic studies.

Case definition (ARFID)

Inclusion criteria Individuals ages 7+ who meet DSM-5 criteria for ARFID. An age 7+ cut-off allows adequate phenotyping with a range of valid instruments available for relevant constructs for this age and older. We anticipate most cases will be pediatric as ARFID typically onsets in childhood. Following a brief online eligibility pre-screen, participants/parents/guardians will be screened for eligibility (for themselves or, in the case of parents/guardians, for their children) and for diagnostic purposes using the questionnaires listed in *Supplement* and Table 1.

Exclusion criteria Current Eating Disorder Examination-Questionnaire (EDE-Q) global score greater than 4.0 or self-induced vomiting, laxative use, or more than four episodes of loss of control eating/binge eating in the past 28 days (DSM-5 ARFID diagnosis requires rule-out of other eating disorders). However, we will carefully

monitor those screening eligible and ineligible for the study to evaluate this exclusion criterion, because adequate longitudinal data do not yet exist documenting the frequency with which ARFID may transition to other eating disorders over time.

Recruitment

We will use a multi-pronged recruitment approach including: outreach to ED clinicians and programs across the country, traditional media (press releases and newspaper announcements), and social media and other online platforms (websites, Facebook, Twitter, Instagram, and podcasts). This includes use of social media ads on Facebook and Instagram, which have proven successful for other similar studies [19]. We enrolled a group of ARFID-GEN parent stakeholders who have consulted with us on study design prior to finalizing our methods.

Procedure

Self-report measures

Table 1 presents the age-appropriate assessment instruments that are completed by parents/guardians of children with ARFID, children with ARFID (ages 7–17 with variable age-appropriate formats), and adults with ARFID (ages 18+). The battery includes validated instruments that capture: ARFID diagnosis and symptoms; other eating disorder diagnoses and symptoms; co-occurring psychiatric disorders and symptoms; general health and neurodevelopmental disorders; impairment; and environmental exposures. Complete information on all self-report questionnaires is available in the *Supplement*.

Parents/guardians of children ages 7–17 and their children

Parents/guardians of children with ARFID who are interested in participating in the study will visit the website (arfidgen.org) and select “Take Our Survey” with the child available. The first step requires the parent/guardian to answer prescreen questions and to consent to the study which includes answering questionnaires and having their child provide a saliva sample for DNA extraction. Parents also have the option to consent to be recontacted for future research. To determine eligibility of the child, parents/guardians then complete a parent report version of the Nine Item ARFID Screen (NIAS-PR) [20]; a parent version of the ARFID self-report version of the Pica, ARFID, and Rumination Disorder Interview ARFID questionnaire (PARDI-AR-Q) [21]; and Version 2.0 of the Parent Version of the Eating Disorder Examination-Questionnaire (PEDE-Qv2.0) [22], developed by KL Loeb based on the Eating Disorder Examination-Questionnaire Version 6 (EDE-Qv6) [23]. If the child is eligible based on parent/guardian report, the parent/guardian provides contact information, the

Table 1 ARFID-GEN Assessment Battery

Domain	Assessment	Version	Parent- or self-report	Age
ARFID symptoms and diagnosis				
ARFID symptoms	Nine Item ARFID Screen (NIAS)	NIAS [20]	Self	14+
		NIAS-Parent Report (NIAS-PR) [20, 49]	Parent	7–17
ARFID diagnosis	Pica, ARFID, and Rumination Disorder Interview-ARFID-Questionnaire (PARDI-AR-Q)	PARDI-AR-Q [21]	Self	14+
		PARDI-AR-Q parent [21]	Parent	7–17
Other eating disorder pathology				
Lifetime eating disorder diagnoses	ED100K Version 3 (ED100Kv3)	ED100Kv3 [26]	Self	15+
Current eating disorder symptoms (last 28 days)	Eating Disorder Examination-Questionnaire (EDE-Q)	EDE-Qv6 [23]	Self	15+
		Child Version of the Eight Item EDE-Q (ChEDE-Q8) [24]	Self	7–14
		Version 2.0 of the Parent Version of the EDE-Qv6 (PEDE-Qv2.0) [22]	Parent	7–17
Depression, anxiety, and neurodevelopment				
Current depressive symptoms	Patient Health Questionnaire (PHQ)	PHQ-9 [50]	Self	18+
		PHQ-Adolescents (PHQ-A) ^a [51]	Self	12–17
Current depressive symptoms (last two weeks)	Short Mood and Feelings Questionnaire (MFQ) [52]	MFQ [52, 53]	Self	18+
		MFQ child version [52, 53]	Self	7–17
		MFQ parent version [52, 53]	Parent	7–11
Current anxiety symptoms	Generalized Anxiety Disorder-7 (GAD-7)	GAD-7 [54]	Self	12+
Major depression and anxiety disorders	Revised Child Anxiety and Depression Scale (RCADS)	RCADS-Child Version (RCADS-C) ^b [55, 56]	Self	7–11
		RCADS-Parent Version (RCADS-P) ^b [57, 58]	Parent	7–11
Obsessive–compulsive symptoms	Obsessive–Compulsive Inventory-Revised (OCI-R)	OCI-R [59]	Self	12+
Intellectual/ developmental disability	Three intellectual/ developmental questions (developed for this study)	N/A	Parent	7–17
Temperament	One Item Temperament Scale	Temperament [60]	Parent	7–17
Detailed assessment of major depressive disorder and generalized anxiety disorder	Items from the mood and anxiety questionnaire for the Genetic Links to Anxiety and Depression Study (GLAD) [61]	GLAD mood & anxiety items (GLAD) ^c	Self	15+
Impairment				
Eating disorder-specific health-related quality of life	Eating Disorders-Quality of Life (EDQOL)	EDQOL [62, 63]	Self	18+
General health-related quality of life	Short Form Health Survey-12 (SF-12)	SF-12 [64]	Self	15+
Health-related quality of life	Pediatric Quality of Life Inventory 4.0 (Peds-QL 4.0)	Peds-QL 4.0 for children [65]	Self	7–12
		Peds-QL 4.0 for adolescents [65]	Self	13–17
		Peds-QL 4.0 for parents of children [65]	Parent	7–12
		Peds-QL 4.0 for parents of adolescents [65]	Parent	13–17
General health	General health questions (developed for this study)	Adult version	Self	18+
		Parent version	Parent	7–17
Environmental exposures				
Negative life events	Avon Longitudinal Study of Parents and Children (ALSPAC) life events checklists [66]	Major Life Changes (MLC) ^c [67]	Self	12+
		Upsetting Events (UE) ^c [67]	Parent	7–11
Mother's pregnancy history	Pregnancy History Questionnaire (PH) [68]	PH	Self	18+
		PH parent version	Parent	7–17
Psychological health	Strengths and Difficulties Questionnaire (SDQ) [69]	SDQ adult single-sided version [70]	Self	18+
		SDQ child single-sided version [70]	Self	12–17
		SDQ parent single-sided version [70]	Parent	7–11
Other				
Experiences with ARFID	Free response question for other comments on experiences with ARFID (developed for this study)	Adult version	Self	18+
		Parent version	Parent	7–17

^a Item 9: "Thoughts that you would be better off dead, or of hurting yourself in some way?" was not included^b Items about major depressive disorder (MDD) were not included^c Modified for this study

child provides assent to participate, and the parent/guardian acknowledges that the child provided verbal assent as well.

Children ages 7–13 then complete the ChEDE-Q8 [24], a child version of the 8-item short form of the Eating Disorders Examination-Questionnaire [25]. Children age 14 complete the NIAS [20], PARDI-AR-Q [21], and ChEDE-Q8. Children ages 15–17 complete the NIAS, PARDI-AR-Q, EDE-Qv6, and the ED100Kv3 [26]. Embedded algorithms determine if the child is eligible for the study. If they meet criteria, they are considered enrolled. All consents, assents, and study questionnaires are completed online using Research Electronic Data Capture (REDCap) [27]. Table 1 contains the series of assessments and which version is used. If the child is determined eligible by parent/guardian- and self-report, they are asked to complete additional questionnaires and provide a saliva sample. A saliva sample collection kit for the child with return packaging is mailed to the parent/guardian with directions addressed to the parents/guardian to oversee the saliva collection from the child. Parents/guardians mail back the completed saliva sample. Once the kit is received by the study team and all questionnaires are finished, participation in the study is complete. For a flowchart of study procedures, see Fig. 1.

Adults (age 18+) with ARFID

Adults visit the website (arfidgen.org) and click the “Take Our Survey” link. Participants complete a brief pre-screen, providing online informed consent for the entire study, have the option to consent to be recontacted for future studies, and provide contact information. Next, they complete the NIAS and the adult version of the PARDI-AR-Q to confirm the presence of ARFID, the EDE-Qv6 to rule out other current eating disorder symptoms, and the ED100Kv3. Surveys are presented in REDCap [27]. Embedded algorithms determine if participants meet inclusion criteria (DSM-5 criteria for ARFID and no other current eating disorder symptoms that warrant exclusion). If they meet criteria, they are considered enrolled. Enrolled participants are asked to complete additional questionnaires (Table 1) and provide a saliva sample. A saliva collection kit is mailed to the participant’s home and the completed sample is returned. Participation in the study is complete once the kit is received by the study team and all questionnaires are finished. For a flowchart of study procedures, see Fig. 1.

Saliva sampling

Saliva samples are collected with Isohelix saliva collection kits and returned to the Center for Psychiatric Genetics (CPG) Biorepository at UNC.

Gift cards

Parents and adult participants are sent a gift card (\$25) once all required questionnaires are complete and their spit kit is received by the study team.

DNA extraction and genotyping

DNA extraction and GWAS genotyping are standard. We will use the most contemporary chip appropriate for diverse ancestry populations when genotyping occurs.

Planned data analysis

Aim 1a. Ascertainment of 3,000 ARFID cases

We will conduct descriptive analyses to characterize the sample stratified by pediatric and adult cases. We will report demographics, symptom patterns, onset, course of illness, comorbid psychiatric conditions, neurodevelopmental characteristics, environmental exposures, and health-related quality of life in child (parent/guardian and child report) and adult ARFID cases.

Aim 1b. Validation substudy

Diagnoses obtained with the PARDI interview will be compared with those obtained with the online ARFID-GEN battery. We will calculate positive predictive value (PPV) to confirm diagnostic properties of our ARFID-GEN battery. Data on weight, height, body mass index (BMI), and BMI percentile (for those < 18) will be verified during remote interviews.

Aim 2. Within-disorder ARFID GWAS

We will conduct a comprehensive set of genomic analyses including SNP-based heritability, GWAS, r_g s, and PRS, and standard post-GWAS analyses of ARFID and the non-mutually exclusive ARFID presentations, and rare CNVs and CNV burden. The following sections briefly summarize the Aim 2 analyses; a more detailed description can be found in the *Supplement*.

- a) *PGC Ricopili pipeline supports rapid analysis.* As described in our previous PGC-ED publications [28], and the *Supplement*, we will use “Ricapili” software [29] for pre-imputation quality control, principal components analysis (PCA), imputation, and meta-analysis. Briefly, we will first follow standard methods to retain high quality SNPs and subjects. Next, we will perform imputation using the largest available resources (currently Haplotype Reference Consortium [HRC]) [30] updating to Trans-omics for Precision Medicine (TOPMed) [31] (N=65K 30x whole genome sequencing [WGS]). Ancestry will be assessed using PCA for each subject, mapped relative to reference samples of known ancestry. Consistent

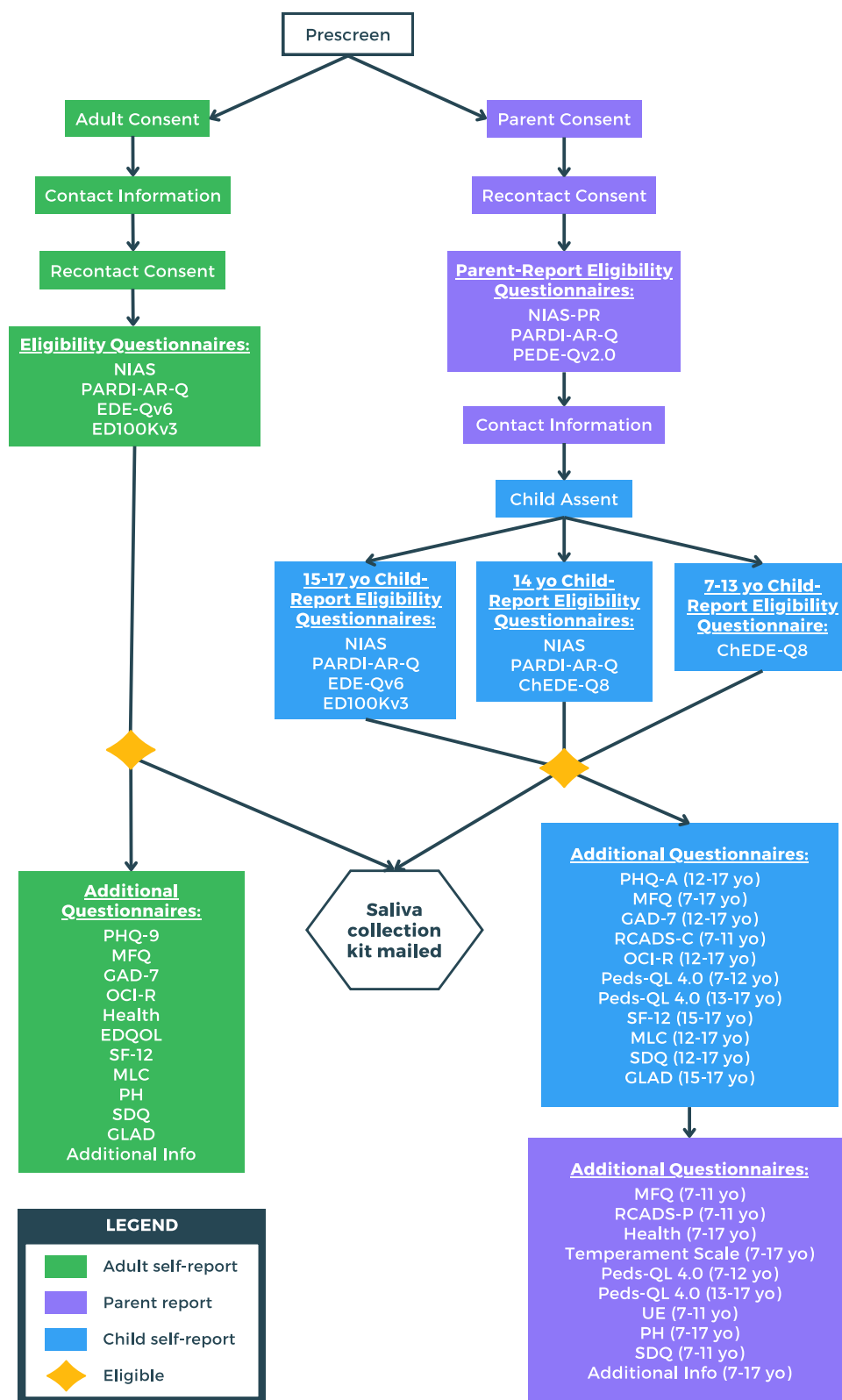


Fig. 1 Flow diagram of ARFID-GEN study

with our intention to include non-European ancestries, we will use established Psychiatric Genomics Consortium (PGC) cross-ancestry analytical approaches.

- b) *Post-GWAS analyses.* The following outlines standard analytic strategy for post-GWAS analysis to maximize information yield and interpretability. The field and methodology evolve rapidly. Below represents what we would do today; however, novel proven methods may emerge before analyses are conducted. Greater detail can be found in the *Supplement*.

Analysis of chrX X chromosome (chrX) variants in the pseudo-autosomal regions will be handled separately. SNPs with MAF > 0.01 and INFO > 0.70 will be retained.

Females and males We will conduct secondary GWAS analysis separately on females and males to determine similarity of the results to the primary combined GWAS.

Clumping GWAS results implicate genomic regions (“loci”). To define a locus, SNPs with $P < 5 \times 10^{-8}$ will be identified and “clumping” will be used to convert significant SNPs to regions.

Conditional and joint analysis Conditional and joint analysis will be conducted using genome-wide complex trait analysis-conditional and joint analysis (GCTA-COJO) [32]. GCTA-COJO investigates every locus with a joint combination of independent markers via a genome-wide SNP selection procedure.

Functional genomic integration We now routinely use functional genomic results from CommonMind, PsychENCODE, and other efforts to understand GWAS results [33]. Much of this is automated in the Functional Mapping and Annotation of Genome-Wide Association Studies platform (FUMA) [34]. We also integrate brain single cell ribonucleic acid sequencing (RNA-seq) data to identify the cell types implied by the GWAS results. See the PGC major depressive disorder (MDD) paper for examples [35].

SNP-based heritability Linkage disequilibrium score regression (LDSC) will be used to estimate SNP-based heritabilities for ARFID and related presentations [36, 37].

Polygenic risk scores (PRS) PRS aggregate risk alleles across the genome weighted by effect sizes. We will use both the classical p-value thresholding method and a summary statistics version of Bayesian multiple regression (SBayesR) [38].

ARFID presentation analysis ARFID presentations are overlapping. We will examine the association between ARFID PRS and: a) PARDI-AR-Q subscale scores (sensory sensitivity, phobic avoidance, and low interest/appetite) as continuous scores; and b) a variable categorizing individuals as having a predominant

ARFID presentation (e.g., sensory vs. phobic vs. low interest/appetite) defined as highest subscale score.

Gene-wise analysis Multi-marker analysis of genomic annotation (MAGMA) [39] will be used to perform gene-wise tests of association with ARFID based on GWAS summary statistics. MAGMA generates gene-based p-values by combining SNP-based p-values within a gene while accounting for linkage disequilibrium (LD).

Partitioned heritability Partitioned heritability will be investigated using stratified LDSC [40], which estimates the per-SNP contribution to overall SNP-heritability (SNP-h²) across functional annotation categories.

Gene expression We will investigate whether ARFID heritability is enriched in tissue/cell type specifically expressed genes using publicly available gene expression data (e.g., the Genotype-Tissue Expression project [GTEx]).

Predicted tissue-specific gene expression We will predict differential gene expression using S-PrediXcan v1.0 [41] and genomic and transcriptomic reference data from the brain regions assayed in CommonMind, GTEx and other resources.

Pathway analyses We will evaluate whether genes associated with ARFID are enriched in specific pathways, tissues, or cell types. To do this, we will use FUMA to annotate SNPs, identify independent loci, perform pathway analysis, and integrate with a wide array of functional genomic data including gene expression, single cell gene expression, and all available brain epigenetic information.

- c) *Discover structural variation associated with ARFID.* The PGC CNV working group is actively optimizing CNV calling using global screening array (GSA) data and we will follow these developments closely. We will maximize comparability between cases and controls by applying rigorous quality control (QC) [42–44]. Following QC, we will fit a series of linear models with CNV burden (at different size thresholds) as the dependent variable to investigate both disease status and experimental biases that can potentially confound CNV detection.

Aim 3. Genetic relation of ARFID to other eating disorders

Greater detail on Aim 3 analyses can be found in the *Supplement*.

- a) *Disorder-specific GWAS.* We will conduct disorder-specific GWAS for ARFID (and AN, BN, and BED as part of other projects) combining ARFID-GEN with EDGI and existing PGC-ED data using imputed variant dosages and an additive model. We anticipate

aggregate sample sizes for the PGC-ED by ~2025 (when our collection would be complete) of: (Projected: ARFID=3,000, AN=50,000, BN=20,000, BED=20,000, ARFID-GEN controls [sourced from existing data repositories]=42,439, PGC-ED controls=543,967). GWAS meta-analysis will be conducted with ARFID-GEN and any other ARFID samples that may be introduced to the PGC-ED by time of analysis.

Combined eating disorder GWAS. Combining ARFID-GEN with PGC-ED GWAS of AN, BN, and BED (which will be available by the time our data are analysis-ready), conduct GWAS meta-analysis of all EDs and of component behaviors that cross-cut disorders (e.g., binge eating, restriction), increasing power to identify genetic risk factors that are common across the four disorders.

- b) *Genetic correlations.* Common variant-based genetic correlations (SNP- r_g) measure the extent to which two traits or disorders share common genetic variation. We will calculate SNP- r_g for ARFID, and selected traits using GWAS summary statistics via an analytical extension of LDSC [36, 37], as well as explore genome-based restricted maximum likelihood (GREML), which was recently shown to have higher accuracy than LDSC [45]. Risk factors classically considered as environmental, such as exercise for AN, are now known to be complex traits underpinned by genetic and non-genetic factors. Large community cohorts like United Kingdom (UK) Biobank, generate GWAS summary statistics for a plethora of phenotypes (e.g., diet, medication, blood metabolites). It is very fast to estimate r_g from GWAS summary statistics, and this screening process has the potential to identify unknown associations which will guide the hypotheses of down-stream analyses.

Generalized summary data-based Mendelian randomization (GSMR). Guided by the r_g estimated above and our hypotheses, we will perform bidirectional MR analyses to investigate causal relationships between correlated traits and ARFID. For example, it is reasonable to test hypotheses of causality using anxiety as an exposure for ARFID. Significant MR results must be reviewed with caution with respect to strong conclusions about causality as unmeasured confounders may exist. Nonetheless, these analyses are potentially exceptionally informative. MR analyses take SNPs that are genome-wide significant for one trait (the exposure) and test the correlation in effect sizes in a second trait (the outcome). Under pleiotropy, there is an expectation that the mean

effect size in the outcome trait is different from zero, but under causality a directional relationship in effect sizes is expected. Different versions of MR analysis are highly related; at the time of analysis, we will implement best practices for MR.

Multi-trait-based conditional and joint analysis. Results from a) and b) will inform these analyses. For example, we expect to detect genetic correlations of BMI with EDs, and MR analyses will aid interpretation of correlation by causality. We will conduct conditional GWAS analyses to determine if the detected SNP associations for EDs can be explained through their relationship with correlated traits. We will perform a multi-trait-based conditional and joint analysis (GCTA-mtCOJO) [16] using an extension of GCTA [46]. This method uses summary-level data to perform conditional analyses. Based on our previous work, we expect to condition the results of our ARFID GWAS on the best available GWAS results for relevant traits including intelligence quotient (IQ), education years, type 2 diabetes, high density lipid cholesterol, BMI, schizophrenia (SCZ), MDD, ASD, OCD, and neuroticism. Comparing these results to those generated from unconditional GWAS results (i.e., from a) above) provides insights into the forces shaping the shared genetic relationships between disorders.

Disorder-specific SNP associations. A key question in ED research is to understand the differences as well as the similarities among the EDs. This question is very similar to the one posed by PGC colleagues interested in understanding the similarity/differences between SCZ and bipolar disorder (BIP) [47]. Guided by their analyses, we will conduct case vs. case (e.g., ARFID-AN) GWAS. This approach is powerful if case samples can be grouped (e.g., if genotyped together so that technical confounding factors are not present), as sampling errors associated with control allele frequency estimates are avoided.

Multi-polygenic score (MPS). To gain insight into factors underlying ED heterogeneity, we will use the largest available GWAS summary statistics for psychiatric and somatic disorders/traits and combine derived PRS into MPS to predict target outcomes (ARFID, AN, BN, or BED diagnosis), and then more granular phenotypes (e.g., age of onset, severity, low weight). Combining ARFID-GEN with EDGI and existing AN, BN, and BED cohorts in the PGC-ED samples should yield a sample size of ~95,000 ED cases and >500,000 controls. We will increase power by using MPS to combine the predictive power of several PRS in one regression model. Training data

will combine the best available GWAS summary statistics on psychiatric, metabolic, BMI, anthropometric, personality, physical activity, and educational phenotypes. These PRS will be used as genetic predictors in models of ARFID, AN, BN, BED, and more granular phenotypes. This approach is ideal for our overarching intentions of predicting outcomes rather than discovering their etiology (i.e., using, not finding genes). MPS is valuable when trait prediction is a priority.

- c) *GSEM*. We will use lifetime ARFID, AN, BN, and BED GWAS summary statistics. We will employ *GSEM* [18] to identify genetic factors for ARFID and associated EDs. *GSEM* is a multivariate method for analyzing the joint genetic architecture of complex traits. By modeling covariance structure, *GSEM* synthesizes genetic correlations and SNP heritabilities inferred from GWAS summary statistics of individual traits from samples with varying and unknown degrees of overlap. *GSEM* analyses will include several steps, including a factor analysis of correlated traits, estimating SNP effects and computation of factor-level PRS.

Aim 4. Genetic relation of ARFID with psychiatric, metabolic/ anthropometric, neurodevelopmental, and other relevant phenotypes

Analyses will parallel Aim 3 only with an outward focus on traits other than EDs. For r_g s, we will follow methods applied in the Anorexia Nervosa Genetics Initiative-PGC-ED (ANGI-PGC-ED) Freeze 2 analysis [28], adding additional traits as GWAS summary statistics become available. Only GWAS summary statistics are needed for *GSEM*, many of which are publicly available. Phenotypes of interest are not directly evaluated in the target sample. Related traits for *GSEM* may include: lifetime anxiety disorder (ANX), neuroticism, BMI, fat mass, and fat-free mass (available in the UK Biobank); ASD, attention-deficit/hyperactivity disorder (ADHD), post-traumatic stress disorder (PTSD), MDD, and OCD GWAS summary statistics (available from the PGC).

Discussion

In addition to the science, we will create rich data and sample resources for the pursuit of related research questions. Our analytic aims are dense, but results will inform follow-on research questions such as: 1) How do genetic and environmental exposures act and co-act to influence risk for ARFID? 2) If detected, how will carriers of CNVs differ on PRS and environmental exposures? 3) Can we identify genetic factors that influence course of illness (e.g., predict who is at risk for developing persistent vs. transient ARFID)? 4) Can we answer precision-medicine

questions regarding identification of optimal interventions informed by genotype and environmental exposures? Given the paucity of effective interventions for ARFID (and other EDs) [48], ultimately, we hope our work will yield information on critical biological pathways that may point toward drug discovery or repurposing that could aid in reversing the tenacity and lethality of these illnesses.

Abbreviations

ADHD	Attention-deficit/hyperactivity disorder
ALSPAC	Avon Longitudinal Study of Parents and Children
AN	Anorexia nervosa
ANGI	Anorexia Nervosa Genetics Initiative
ANX	Anxiety disorder
ARFID	Avoidant/restrictive food intake disorder
ARFID-GEN	Avoidant Restrictive Food Intake Disorder: Genes and Environment Study
ASD	Autism spectrum disorder
BED	Binge-eating disorder
BIP	Bipolar disorder
BMI	Body mass index
BN	Bulimia nervosa
ChEDE-Q8	Child version of the 8-item short form of the Eating Disorders Examination-Questionnaire
chrX	X chromosome
CNV	Copy number variants
dbGaP	Data base of Genotypes and Phenotypes
DNA	Deoxyribonucleic acid
DSM-5	Diagnostic and Statistical Manual of Mental Disorders 5th Edition
ED	Eating disorder
ED100Kv3	ED100K Version 3
EDE-Q	Eating Disorders Examination-Questionnaire
EDE-Qv6	Eating Disorders Examination-Questionnaire Version 6
EDGI	Eating Disorders Genetics Initiative
EDQOL	Eating Disorders Quality of Life
FUMA	Functional Mapping and Annotation of Genome-Wide Association Studies platform
GAD-7	Generalized Anxiety Disorder-7
GCTA	Genome-wide complex trait analysis
GTCA-COJO	Genome-wide complex trait analysis-conditional and joint analysis
GLAD	Genetic Links to Anxiety and Depression Study
GREML	Genome-based restricted maximum likelihood
GSA	Global screening array
GSEM	Genomic structural equation modelling
GSMR	Generalized summary data-based Mendelian randomization
GTE _x	Genotype-Tissue Expression project
GWAS	Genome-wide association study
h^2	Heritability estimate
HRC	Haplotype Reference Consortium
IQ	Intelligence quotient
LD	Linkage disequilibrium
LDSC	Linkage disequilibrium score regression
MAF	Minor allele frequency
MAGMA	Multi-marker analysis of genomic annotation
MDD	Major depressive disorder
MFQ	Short Mood and Feelings Questionnaire
MLC	ALSPAC Major Life Changes
MPS	Multi-polygenic score
MR	Mendelian randomization
mtCOJO	Multi trait conditional and joint analysis
NIAS	Nine Item ARFID Screen
NIAS-PR	Nine Item ARFID Screen-Parent Report
NIMH	National Institute of Mental Health
OCD	Obsessive compulsive disorder

OCI-R	Obsessive Compulsive Inventory-Revised
PARDI	Pica, ARFID, and Rumination Disorder Interview
PARDI-AR-Q	Pica, ARFID, and Rumination Disorder Interview-ARFID-Questionnaire
PCA	Principal components analysis
PEDE-Qv2.0	Version 2.0 of the Parent Version of the Eating Disorders Examination-Questionnaire Version 6
Peds-QL 4.0	Pediatric Quality of Life Inventory 4.0
PGC	Psychiatric Genomics Consortium
PGC-ED	Eating Disorders Working Group of the Psychiatric Genomics Consortium
PH	Pregnancy History Questionnaire
PHQ	Patient Health Questionnaire
PHQ-9	Patient Health Questionnaire-9
PHQ-A	Patient Health Questionnaire-Adolescents
PPV	Positive predictive value
PRS	Polygenic risk scores
P _T	P-value thresholds
PTSD	Post-traumatic stress disorder
QC	Quality control
RCADS	Revised Child Anxiety and Depression Scale
RCADS-C	Revised Child Anxiety and Depression Scale-Child Version
RCADS-P	Revised Child Anxiety and Depression Scale-Parent Version
REDCap	Research Electronic Data Capture
r _g	Genetic correlation
RNA-seq	Ribonucleic acid sequencing
SBayesR	A summary statistics version of Bayesian multiple regression
SCZ	Schizophrenia
SDQ	Strengths and Difficulties Questionnaire
SF-12	Short Form Health Survey-12
SNP	Single nucleotide polymorphism
SNP-r _g	Common variant based genetic correlations
TOPMed	Trans-omics for Precision Medicine
UE	ALSPAC Upsetting Events
UK	United Kingdom
UNC	University of North Carolina at Chapel Hill
WGS	Whole genome sequencing

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12888-023-05266-x>.

Additional file 1.

Acknowledgements

The ARFID-GEN team acknowledges contributions of Ana Dobrot, Evelyn Watkins, and Ava Iwashita for their input on aspects of ARFID-GEN design. UNC Center of Excellence for Eating Disorder 2023 summer fellows Wesley R Barnhart, Kayla Costello, and Hannah Norling provided input and contributed to participant recruitment methodology. Study data are collected and managed using REDCap electronic data capture tools provided by the NC Translational and Clinical Sciences (NC TraCS) Institute, which is supported by the National Center for Advancing Translational Sciences (NCATS), National Institutes of Health, through Grant Award Number UL1TR001111. We also acknowledge the NCTraCS Community and Stakeholder Engagement (CaSE) team for assistance with community feedback sessions and the parent group F.E.A.S.T for assisting with making parents of children with ARFID aware of the ARFID-GEN study.

Authors' contributions

Conception/design of the work: CMB, NM, CMM, MAM-C, LMT, LD, JG, MH, JJC Data acquisition/analysis: CMB, NM, CMM, BQ, MAM-C, LMT, JW, EMP, JJ, KR, SNO, JG, BQ Creation of software: JG Drafted/ revised the work: CMB, NM, CMM, BQ, MAM-C, LMT, JW, LD, EMP, JJ, KR, SNO, AES, NB, AD, JG, MH, JJC Approved submitted version: CMB, NM, CMM, BQ, MAM-C, LMT, JW, LD, EMP, JJ, KR, SNO, AES, NB, AD, JG, MH, JJC Agreed both to be personally accountable for the author's own contributions and to ensure that questions related to the accuracy or integrity of any part of the work, even ones in which the author

was not personally involved, are appropriately investigated, resolved, and the resolution documented in the literature: CMB, NM, CMM, BQ, MAM-C, LMT, JW, LD, EMP, JJ, KR, SNO, AES, NB, AD, JG, MH, JJC.

Funding

Open access funding provided by Karolinska Institute. National Institute of Mental Health (R56MH129437 [Bulik/Micali (MPI)]; R01 MH124871 [Sullivan/Bulik, MPI]). No funding bodies were involved in the design of the study and collection, analysis, interpretation of data, or writing the manuscript. The NIMH peer-reviewed the study protocol.

Availability of data and materials

Our liberal data and analysis sharing principles will make phenotypic and genotype data and scripts widely available for access by other scientists to maximize utility of our investigation. The datasets generated and/or analyzed will be available in the National Data Archive (<https://nda.nih.gov/>). Genomic data access is made possible by the Psychiatric Genomics Consortium Data Access Committee. Step-by-step procedures are thoroughly described here (<https://www.med.unc.edu/pgc/shared-methods/open-source-philosophy/>). DNA samples will be available from the NIMH Repository and Genomics Resource (<https://www.nimhgenetics.org/order-biosamples/how-to-order-biosamples>).

Declarations

Ethics approvals and consent to participate

ARFID-GEN was approved by the University of North Carolina Biomedical Institutional Review Board (IRB) Protocol # 22–1524. Informed consent will be obtained from all participants and/or their legal guardian(s). Child assent will also be obtained. All methods were carried out in accordance with relevant guidelines and regulations.

Consent for publication

Not applicable.

Competing interests

CMB: Pearson (author, royalty recipient); Equip Health Inc. (Stakeholder Advisory Board). NM: receives an honorarium for services as Associate Editor for the European Eating Disorders Review. LD: reports speaker fees from Baxter Medical AB and Fresenius Kabi AB. CMM, BQ, MAM-C, LMT, JW, EMP, JJ, KR, SNO, AES, NB, AD, JG, MH, JJC: no competing interests to declare.

Received: 19 July 2023 Accepted: 9 October 2023

Published online: 21 November 2023

References

- Dinkler L, Bryant-Waugh R. Assessment of avoidant restrictive food intake disorder, pica and rumination disorder: interview and questionnaire measures. *Curr Opin Psychiatry*. 2021;34(6):532–42.
- American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders* (5th ed.). Arlington, VA: American Psychiatric Publishing; 2013.
- Thomas JJ, Lawson EA, Micali N, Misra M, Deckersbach T, Eddy KT. Avoidant/restrictive food intake disorder: a three-dimensional model of neurobiology with implications for etiology and treatment. *Curr Psychiatry Rep*. 2017;19(8):54.
- Kambanis PE, Kuhnle MC, Wons OB, Jo JH, Keshishian AC, Hauser K, et al. Prevalence and correlates of psychiatric comorbidities in children and adolescents with full and subthreshold avoidant/restrictive food intake disorder. *Int J Eat Disord*. 2020;53(2):256–65.
- Dinkler L, Lichtenstein P, Birgegard A, Bulik CM. Etiology of the broad avoidant restrictive food intake disorder phenotype in Swedish twins aged 6–12 years. *JAMA Psychiat*. 2023;80:260–9.
- Yilmaz Z, Hardaway J, Bulik C. Genetics and epigenetics of eating disorders. *Adv Genom Genet*. 2015;5:131–50.

7. Javaras KN, Laird NM, Reichborn-Kjennerud T, Bulik CM, Pope HG Jr, Hudson JL. Familiality and heritability of binge eating disorder: results of a case-control family study and a twin study. *Int J Eat Disord*. 2008;41(2):174–9.
8. Reichborn-Kjennerud T, Bulik C, Tambs K, Harris J. Genetic and environmental influences on binge eating in the absence of compensatory behaviours: a population-based twin study. *Int J Eat Disord*. 2004;36:307–14.
9. Reilly EE, Brown TA, Gray EK, Kaye WH, Menzel JE. Exploring the co-occurrence of behavioural phenotypes for avoidant/restrictive food intake disorder in a partial hospitalization sample. *Eur Eat Disord Rev*. 2019;27(4):429–35.
10. Fisher MM, Rosen DS, Ornstein RM, Mammel KA, Katzman DK, Rome ES, et al. Characteristics of avoidant/restrictive food intake disorder in children and adolescents: a “new disorder” in DSM-5. *J Adol Health*. 2014;55(1):49–52.
11. Nicely TA, Lane-Loney S, Masciulli E, Hollenbeak CS, Ornstein RM. Prevalence and characteristics of avoidant/restrictive food intake disorder in a cohort of young patients in day treatment for eating disorders. *J Eat Disord*. 2014;2(1):1–8.
12. Pliner P, Loewen ER. Temperament and food neophobia in children and their mothers. *Appetite*. 1997;28(3):239–54.
13. Lucarelli L, Sechi C, Cimino S, Chatoor I. Avoidant/Restrictive Food Intake Disorder: A longitudinal study of malnutrition and psychopathological risk factors from 2 to 11 years of age. *Front Psychol*. 2018;9:1608.
14. Bulik CM, Thornton LM, Parker R, Kennedy H, Baker JH, MacDermod C, et al. The Eating Disorders Genetics Initiative (EDGI): study protocol. *BMC Psychiatry*. 2021;21(1):234.
15. Bryant-Waugh R, Micali N, Cooke L, Lawson EA, Eddy KT, Thomas JJ. Development of the Pica, ARFID, and Rumination Disorder Interview, a multi-informant, semi-structured interview of feeding disorders across the lifespan: A pilot study for ages 10–22. *Int J Eat Disord*. 2019;52(4):378–87.
16. Zhu Z, Zheng Z, Zhang F, Wu Y, Trzaskowski M, Maier R, et al. Causal associations between risk factors and common diseases inferred from GWAS summary data. *Nat Commun*. 2018;9(1):224.
17. Krapohl E, Patel H, Newhouse S, Curtis CJ, von Stumm S, Dale PS, et al. Multi-polygenic score approach to trait prediction. *Mol Psychiatry*. 2018;23(5):1368–74.
18. Grotzinger AD, Rhemtulla M, de Vlaming R, Ritchie SJ, Mallard TT, Hill WD, et al. Genomic structural equation modelling provides insights into the multivariate genetic architecture of complex traits. *Nat Hum Behav*. 2019;3(5):513–25.
19. MacDermod C, Pettie MA, Carrino EA, Garcia SC, Padalecki S, Finch JE, et al. Recommendations to encourage participation of individuals from diverse backgrounds in psychiatric genetic studies. *Am J Med Genet B Neuropsychiatr Genet*. 2022;189(5):163–73.
20. Burton Murray H, Dreier MJ, Zickgraf HF, Becker KR, Breithaupt L, Eddy KT, et al. Validation of the nine item ARFID screen (NIAS) subscales for distinguishing ARFID presentations and screening for ARFID. *Int J Eat Disord*. 2021;54(10):1782–92.
21. Bryant-Waugh R, Stern CM, Dreier MJ, Micali N, Cooke LJ, Kuhnle MC, et al. Preliminary validation of the pica, ARFID and rumination disorder interview ARFID questionnaire (PARDI-AR-Q). *J Eat Disord*. 2022;10(1):179.
22. Drury CR, Hail L, Rienecke RD, Accurso EC, Coelho JS, Lock J, et al. Psychometric properties of the Parent Eating Disorder Examination Questionnaire. *Int J Eat Disord*. 2023.
23. Fairburn C, Beglin S. Eating Disorder Examination Questionnaire (6.0). In: Fairburn C, editor. *Cognitive behavior therapy for eating disorders*. New York: Guilford; 2008.
24. Kliem S, Schmidt R, Vogel M, Hiemisch A, Kiess W, Hilbert A. An 8-item short form of the Eating Disorder Examination-Questionnaire adapted for children (ChEDE-Q8). *Int J Eat Disord*. 2017;50(6):679–86.
25. Kliem S, Mossle T, Zenger M, Strauss B, Brahler E, Hilbert A. The eating disorder examination-questionnaire 8: A brief measure of eating disorder psychopathology (EDE-Q8). *Int J Eat Disord*. 2016;49(6):613–6.
26. Thornton L, Munn-Chernoff M, Baker J, Juréus A, Parker R, Henders A, et al. The Anorexia Nervosa Genetics Initiative (ANGI): Overview and methods. *Contemp Clin Trials*. 2018;74:61–9.
27. Harris P, Taylor R, Thielke R, Payne J, Gonzalez N, Conde J. Research electronic data capture (REDCap)—a metadata-driven methodology and workflow process for providing translational research informatics support. *J Biomed Inform*. 2009;42:377–81.
28. Watson H, Yilmaz Z, Thornton L, Hübel C, Coleman J, Gaspar H, et al. Genome-wide association study identifies eight risk loci and implicates metabolic-psychiatric origins for anorexia nervosa. *Nat Genet*. 2019;51:1207–14.
29. Lam M, Awasthi S, Watson HJ, Goldstein J, Panagiotaropoulou G, Trubetskoy V, et al. RICOPILI: Rapid Imputation for Consortiums Pipeline. *Bioinform*. 2020;36(3):930–3.
30. Consortium THR. A reference panel of 64,976 haplotypes for genotype imputation. *Nat Genet*. 2016;48(10):1279–83.
31. Brody J, Morrison A, Bis J, O’Connell J, Brown M, Huffman J, et al. NHLBI Trans-Omics for Precision Medicine (TOPMed) Consortium, Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Consortium, TOPMed Hematology and Hemostasis Working Group, CHARGE Analysis and Bioinformatics Working Group. Analysis commons, a team approach to discovery in a big-data environment for genetic epidemiology. *Nat Genet*. 2017;49:1560–3.
32. Yang J, Loos RJF, Powell JE, Medland SE, Speliotes EK, Chasman DI, et al. FTO genotype is associated with phenotypic variability of body mass index. *Nature*. 2012;490(7419):267–72.
33. Fromer M, Roussos P, Sieberts SK, Johnson JS, Kavanagh DH, Perumal TM, et al. Gene expression elucidates functional impact of polygenic risk for schizophrenia. *Nat Neurosci*. 2016;19(11):1442–53.
34. Watanabe K, Taskesen E, Van Bochoven A, Posthuma D. Functional mapping and annotation of genetic associations with FUMA. *Nat Commun*. 2017;8(1):1–11.
35. Wray N, Ripke S, Mattheisen M, Trzaskowski M, Byrne E, Abdellaoui A, et al. Genome-wide association analyses identify 44 risk variants and refine the genetic architecture of major depression. *Nat Genet*. 2018;50(5):668.
36. Bulik-Sullivan B, Finucane H, Anttila V, Gusev A, Day F, ReproGen Consortium, et al. An atlas of genetic correlations across human diseases and traits. *Nat Genet*. 2015;47:1236–41.
37. Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J, Schizophrenia Working Group of the Psychiatric Genomics C, et al. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet*. 2015;47:291–5.
38. Lloyd-Jones LR, Zeng J, Sidorenko J, Yengo L, Moser G, Kemper KE, et al. Improved polygenic prediction by Bayesian multiple regression on summary statistics. *Nat Commun*. 2019;10(1):5086.
39. de Leeuw C, Mooij J, Heskes T, Posthuma D. MAGMA: generalized gene-set analysis of GWAS data. *PLoS Comput Biol*. 2015;11(4):e1004219.
40. Finucane H, Bulik-Sullivan B, Gusev A, Trynka G, Reshef Y, Loh P-R, et al. Partitioning heritability by functional annotation using genome-wide association summary statistics. *Nat Genet*. 2015;47(11):1228–35.
41. Barbeira AN, Dickinson SP, Bonazzola R, Zheng J, Wheeler HE, Torres JM, et al. Exploring the phenotypic consequences of tissue specific gene expression variation inferred from GWAS summary statistics. *Nat Commun*. 2018;9(1):1825.
42. Szatkiewicz JP, Wang W, Sullivan PF, Wang W, Sun W. Improving detection of copy-number variation by simultaneous bias correction and read-depth segmentation. *Nucleic Acids Res*. 2013;41(3):1519–32.
43. Szatkiewicz JP, Neale BM, O’Dushlaine C, Fromer M, Goldstein JL, Moran JL, et al. Detecting large copy number variants using exome genotyping arrays in a large Swedish schizophrenia sample. *Mol Psychiatry*. 2013;18(11):1178–84.
44. Ruderfer DM, Chambert K, Moran J, Talkowski M, Chen ES, Gigek C, et al. Mosaic copy number variation in schizophrenia. *Eur J Hum Genet*. 2013;21(9):1007–11.
45. Ni G, Zeng J, Revez JA, Wang Y, Zheng Z, Ge T, et al. A comparison of ten polygenic score methods for psychiatric disorders applied across multiple cohorts. *Biol Psychiatry*. 2021;90(9):611–20.
46. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet*. 2011;88(1):76–82.
47. Ruderfer DM, Ripke S, McQuillan A, Boocock J, Stahl EA, Pavlides JMW, et al. Genomic dissection of bipolar disorder and schizophrenia, including 28 subphenotypes. *Cell*. 2018;173(7):1705–15.e16.
48. Kaye WH, Bulik CM. Treatment of patients with anorexia nervosa in the US-A crisis in care. *JAMA Psychiat*. 2021;78:591–2.
49. Ziolkowska B, Ocalewski J, Zickgraf H, Brytek-Matera A. The Polish Version of the Avoidant/Restrictive Food Intake Disorder Questionnaire-Parents Report (ARFID-Q-PR) and the Nine Items Avoidant/Restrictive Food

- Intake Disorder Screen-Parents Report (NIAS-PR): Maternal Perspective. *Nutrients*. 2022;14(15).
50. Kroencke K, Spitzer R, Williams J. The PHQ-9: validity of a brief depression severity measure [Electronic version]. *J Gen Intern Med*. 2001;16(9):606–13.
 51. Johnson JG, Harris ES, Spitzer RL, Williams JB. The patient health questionnaire for adolescents: validation of an instrument for the assessment of mental disorders among adolescent primary care patients. *J Adolesc Health*. 2002;30(3):196–204.
 52. Angold A, Costello EJ, Messer S, Pickles A, Winder F, Silver D. Mood and Feelings Questionnaire: the development of a short questionnaire for use in epidemiological studies of depression in children and adolescents. *Int J Methods Psychiatr Res*. 1995;5:237–49.
 53. Duke University Psychiatry and Behavioral Sciences. Mood and Feelings Questionnaire (MFQ). [<https://psychiatry.duke.edu/research/research-programs-areas/assessment-intervention/developmental-epidemiology-instruments-0>] Accessed 18 Jul 2023.
 54. Lowe B, Decker O, Muller S, Brahler E, Schellberg D, Herzog W, et al. Validation and standardization of the Generalized Anxiety Disorder Screener (GAD-7) in the general population. *Med Care*. 2008;46(3):266–74.
 55. Chorpita BF, Yim L, Moffitt C, Umemoto LA, Francis SE. Assessment of symptoms of DSM-IV anxiety and depression in children: a revised child anxiety and depression scale. *Behav Res Ther*. 2000;38(8):835–55.
 56. Chorpita BF, Moffitt CE, Gray J. Psychometric properties of the Revised Child Anxiety and Depression Scale in a clinical sample. *Behav Res Ther*. 2005;43(3):309–22.
 57. Ebesutani C, Chorpita BF, Higa-McMillan CK, Nakamura BJ, Regan J, Lynch RE. A psychometric analysis of the Revised Child Anxiety and Depression Scales—parent version in a school sample. *J Abnorm Child Psychol*. 2011;39(2):173–85.
 58. Ebesutani C, Bernstein A, Nakamura BJ, Chorpita BF, Weisz JR. Research Network on Youth Mental H. A psychometric analysis of the revised child anxiety and depression scale—parent version in a clinical sample. *J Abnorm Child Psychol*. 2010;38(2):249–60.
 59. Foa EB, Huppert JD, Leiberg S, Langner R, Kichic R, Hajcak G, et al. The Obsessive-Compulsive Inventory: development and validation of a short version. *Psychol Assess*. 2002;14(4):485.
 60. Sladdens EF, Hughes SO, O'Connor TM, Beltran A, Baranowski JC, Nicklas TA, et al. The Children's Behavior Questionnaire very short scale: psychometric properties and development of a one-item temperament scale. *Psychol Rep*. 2012;110(1):197–217.
 61. GLAD Study [Available from: <https://gladstudy.org.uk>]. Accessed 18 Jul 2023.
 62. Engel S. Health Related Quality of Life and Disordered Eating: Development and Validation of the Eating Disorders Quality of Life Instrument. [Unpublished Dissertation]. Fargo, ND: North Dakota State University; 2003.
 63. Engel S, Wittrock D, Crosby R, Wonderlich S, Mitchell J, Kolotkin R. Development and psychometric validation of an eating disorder-specific health-related quality of life instrument. *Int J Eat Disord*. 2006;39(1):62–71.
 64. Ware J Jr, Kosinski M, Keller S. A 12-Item Short-Form Health Survey: construction of scales and preliminary tests of reliability and validity. *Med Care*. 1996;34:220–33.
 65. Varni JW, Seid M, Kurtin PS. PedsQL 4.0: reliability and validity of the Pediatric Quality of Life Inventory version 4.0 generic core scales in healthy and patient populations. *Med Care*. 2001;39(8):800–12.
 66. Golding J, Pembrey M, Jones R, Team AS. ALSPAC—the Avon Longitudinal Study of Parents and Children. I. Study methodology. *Paediatr Perinat Epidemiol*. 2001;15(1):74–87.
 67. ALSPAC Child completed questionnaires [Available from: <https://www.bristol.ac.uk/alspac/researchers/our-data/questionnaires/child-completed-questionnaires/>]. Accessed 18 Jul 2023.
 68. Mills-Koonce WR, Willoughby MT, Short SJ, Propper CB. The Brain and Early Experience Study: Protocol for a prospective observational study. *JMIR Res Protoc*. 2022;11(6):e34854.
 69. Goodman R. The strengths and difficulties questionnaire: a research note. *J Child Psychol Psychiatry*. 1997;38:581–6.
 70. youthinmind English (USA) [Available from: [https://www.sdqinfo.org/py/sdqinfo/b3.py?language=Englishqz\(USA\)](https://www.sdqinfo.org/py/sdqinfo/b3.py?language=Englishqz(USA))]. Accessed 18 Jul 2023.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

