Biological approaches to the repair and regeneration of the rotator cuff tendon-bone enthesis: a literature review

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Key Words:

enthesis; rotator cuff; stem cells; tissue engineering; tendon-bone enthesis

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ABSTRACT

Entheses are highly specialised organs connecting ligaments and tendons to bones, facilitating force transmission, and providing mechanical strengths to absorb forces encountered. Two types of entheses, fibrocartilaginous and fibrous, exist in interfaces. The gradual fibrocartilaginous type is in rotator cuff tendons and is more frequently injured due to the poor healing capacity that leads to loss of the original structural and biomechanical properties and is attributed to the high prevalence of retears. Fluctuating methodologies and outcomes of biological approaches are challenges to overcome for them to be routinely used in clinics. Therefore, stratifying the existing literature according to different categories (chronicity, extent of tear, and studied population) would effectively guide repair approaches. This literature review supports tissue engineering approaches to promote rotator cuff enthesis healing employing cells, growth factors, and scaffolds period. Outcomes suggest its promising role in animal studies as well as some clinical trials and that combination therapies are more beneficial than individualized ones. It then highlights the importance of tailoring interventions according to the tear extent, chronicity, and the population being treated. Contributing factors such as loading, deficiencies, and lifestyle habits should also be taken into consideration. Optimum results can be achieved if biological, mechanical, and environmental factors are approached. It is challenging to determine whether variations are due to the interventions themselves, the animal models, loading regimen, materials, or tear mechanisms. Future research should focus on tailoring interventions for different categories to formulate protocols, which would best guide regenerative medicine decision making.

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Introduction

Entheses are highly specialised interfaces between fibrous isotropic tendons and ligaments (elastic modulus = 450 MPa axially) that provide tensile strength, and rigid inelastic isotropic bones (elastic modulus = 20 GPa) that optimise compressive loads, facilitating joint motion. Entheses evolved highly specialised structures that effectively dampen the forces encountered in the region of peak strain, to facilitate seamless stress transfer and prevent tendon avulsion; whilst maintaining effective communication between tendons and bones to preserve homeostasis. ²⁻⁴

This heterogenous interface is unique and owing to its complexity and propensity to heal with biomechanically inferior scar tissue, presents significant challenges to repair and regeneration.

Rotator cuff (RC) tears are common with an increasing prevalence of partial and full thickness tears after the age of 50, and a 90% surgical repair failure. ⁵⁻⁷ The capacity to repair the damaged enthesis is insufficient, because of the tissue's lack of regeneration, leading to scar formation with inferior mechanical properties at the repair site. ⁸ Trials to enhance the healing progress of the RC enthesis have been directed towards

regenerative methods.⁷ To date, there are no records of a single successful clinical biological approach achieving an enhanced healing of the injured enthesis. This review investigates the potential use of tailored biological therapies in different RC tear mechanisms, chronicity, and populations treated.

Entheses types

Histologically, there are two types of entheses classified according to the presence or absence of fibrocartilage, Fibrocartilaginous and Fibrous. Fibrous entheses insert via collagen fibres at the metaphysis or diaphysis to dissipate load over a large area, such as the deltoid and the medial collateral ligament, while fibrocartilaginous are at the epiphyses of long bones, such as the RC and Achilles tendons.^{9, 10} The fibrocartilage of fibrocartilaginous entheses is not distributed evenly throughout the interface; appearing thickest in the deepest layers and absent superficially, consistent with the highest stress distribution.^{11, 12} Fibrocartilaginous entheses, such as the RC, are more commonly injured and will therefore be the focus of this review.¹³

Entheses microstructure

Fibrocartilaginous entheses comprise four layers: dense fibrous tendon, fibrocartilage, mineralised fibrocartilage, and bone. The layers gradually increase in stiffness from tendon to bone to distribute forces over wider surface areas. The first level is composed of organised parallel appearance of collagen I and low levels of Proteoglycans within interspaced elongated tendon fibroblasts. The non-mineralised layer is populated with rows of round fibrochondrocytes embedded in a collagen II- and III-rich extracellular matrix (ECM), and low levels of collagen X, proteoglycans, and glycosaminoglycans. The fibrocartilage and mineralised fibrocartilage regions are separated by a tide mark. The mineralised fibrocartilage merges into bone, which consists of osteoblasts, osteocytes, and osteoclasts, alongside collagen I and a 69% elevated mineral content of which 99% is hydroxyapatite.¹⁴

The progressive structure and composition of entheses allow minimisation of the mechanical vulnerability that would arise due to the large elastic modulus mismatch. The structural and mechanical properties of entheses are a result of mechanical loading that have an essential role in enthesis maturation, allowing load transmission and stress reduction.¹⁴ This highlights the importance of developing repair solutions that mimic the native biological and mechanical integration of entheses.

Entheses mechanical properties

Force transmission via the enthesis dynamically involves fibre realignment, crimp deformation, and sliding, which are essential for remodelling and turnover. Fibrocartilage enthesis acts like a stretch break that limits tendon narrowing at the insertion, which is a region of stress concentration, hence tendons cross-sectional area increases as they insert into the bone. ^{15, 16} The unmineralised region resists compression, while the mineralised resists shearing, creating a two-layered protective component. ¹⁷ Levels of unmineralised fibrocartilage are higher at entheses with more varied ranges of insertion angles, highlighting its functional role. The mineralised fibrocartilage attachment into bone is rather atypical, providing entheses with varied mechanical integrity. ¹⁴ Owing to the complex structure of the enthesis; when damaged, it is extremely challenging to repair surgically. The functionally gradual layers are not restored after repairs and the subsequent healing process. ^{2, 18, 19}

Entheses embryological development

Embryologically, it has been hypothesised that initially hyaline cartilage at the enthesis exists alone, and tendons attach to it. The cartilage erodes from the bone and is replaced by the fibrocartilaginous enthesis akin to a growth plate. This emphasises the fibrocartilaginous enthesis development within the tendon. ^{20,21} In utero, the enthesis is originally organised as an unmineralised cartilaginous unit that is mineralised postnatally by endochondral ossification. 22-24 Transcription factors, such as SRY-box 9, have been found essential for chondrogenesis, and others for tenogenesis such as mohawk and scleraxis. 25-27 Scleraxis knockout mice demonstrated a lower bone mineral density, decreased attachment strength, and disorganised collagen fibres. 23, 28 Growth factors (GFs), such as transforming growth factor-beta (TGF-B) and bone morphogenetic protein (BMP) are regulators in early enthesis formation, and molecules such as Indian hedgehog (Ihh) and parathyroid hormone-related protein regulate late mineralisation.^{22, 23, 29-31} During early development, enthesis progenitor cells proliferate and lengthen it, and express collagen I. Stimulated by BMP-4 and Ihh, at the base of the developing enthesis, the cells differentiate into unmineralised fibrochondrocytes, and express collagen I and II. By stimulation of Ihh at the postnatal stage, fibrochondrocytes undergo hypertrophic differentiation at the base of the enthesis. As the mineralised fibrochondrocytes form, collagen X and alkaline phosphate are expressed.32

Since tissue distortion and decreased mineralisation are seen in paralysed mice models, muscle loading is essential for the developing enthesis maturation alongside molecular components. Disordered cell patterns and lack of enthesis regional transition are effects of reduced muscular stress during development. A balance between GFs and mechanical stimulation gives rise to the mature complex enthesis structure and composition.

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Rotator cuff enthesis and scar tissue

High rates of RC retears are well documented, where various improved suture techniques have been ineffective in reducing them. Surgical repairs for RC tears are often performed via a single or double row suture technique with the aim of approximating the ruptured soft tissue segment from bone. However, due to factors such as age, tear chronicity, and poor vascularity, high retear rates persist with scar-mediated tissue of disorganised matrix and inferior mechanical properties forms. Scar formation is thought to be a result of TGF- β 1 production by macrophages during the inflammatory phase, leading to fibroblast recruitment. Since expression of TGF- β 1 is elevated at injured entheses, it increases collagen I and III deposition and inhibits chondrogenic differentiation.

Factors influencing healing

Despite better outcomes being linked to intact repairs, it does not guarantee patient satisfaction with functional outcomes. 44,45 Variables affecting outcomes of RC repairs include (I) Age: decrease in turnover, fatty infiltration and tear retraction are thought to be the cause for altering the enthesis healing in older populations regardless of the surgical procedure used. 46-51 (II) Muscle atrophy: severe pre-operative atrophy and fatty infiltration were linked to lower post-operative repair integrity. 45, 52, 53 (III) Smoking: nicotine has been linked to impairments in biomechanical properties and delayed healing.⁵⁴ (IV) Diabetes and hypercholesterolemia: they have shown to reduce RC enthesis mechanical properties.⁵⁵ Additionally, diabetes increased post-operative complications, such as infections and failures after RC repairs, while hypercholesterolemia increased tendon-related pathologies. 56,57 (V) Obesity demonstrated inferior mechanical properties and poorer histological outcomes with high fat diet.⁵⁸ This suggests that long-term maintenance of dietary intake to manage lipid levels and blood glucose prior to injury might have a positive influence on the healing process. (VI) Contrasting evidence exists regarding the effect of vitamin D levels on RC enthesis healing. Animal studies suggest a negative influence of low vitamin D on RC repair, while clinical trials found no correlation between vitamin D levels and retear rates.^{59,60} However, a large clinical study of arthroscopic RC repair found bone mineral density to be a reliable indicator of RC recovery.⁶¹ (VII) Non-steroidal anti-inflammatory drugs: they negatively affect healing in the acute post-operative stage for up to 6 weeks, while positively on remodelling of collagen matrix in chronic stages.^{62,63}

Methodology

An electronic database search was performed using UCL's Online Library Service and PubMed. Keywords used were "enthesis AND rotator cuff", "rotator cuff AND tear", "rotator cuff OR enthesis", "tissue engineering AND rotator cuff", "stem cells AND rotator cuff enthesis", and "growth factors AND rotator cuff enthesis". Outcomes of *in-vivo*, *in-vitro*, and clinical studies on RC enthesis repair involving various tear extent, mechanism, chronicity, and population studies were also included. Exclusion criteria included other types of entheses, and surgical and rehabilitative interventions.

A total of 59 relevant studies were found after exclusion criteria were applied. Of these, 45 presented positive, 10 negative, and 4 mixed findings.

Literature was then summarised based on three main interventions: stem cells, GFs, and scaffolds. Findings were then categorised into groups according to their findings and methodologies (acute-full, chronic-full, acute-partial, chronic-partial, in aging and younger populations, and *in-vitro* studies). **Figure 1** summarises the methodology process.

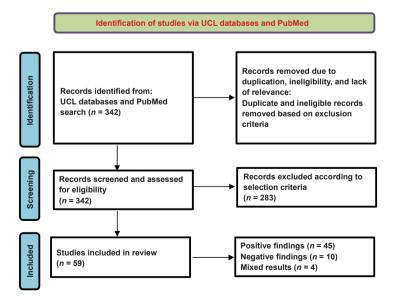


Figure 1. A flowchart representing selection stages of the studies for their inclusion in the literature review.

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Literature Review Findings Cells and rotator cuff enthesis healing

Various stem cells such as mesenchymal stem cells (MSCs), adipose-derived stem cells (ADSCs), and bone marrow MSCs (BMSCs) are under investigation for potential use in enthesis regeneration.⁶⁴

Mesenchymal stem cells

RC enthesis repairs could benefit from MSCs' capacity for self-renewal and multipotency, their availability, accessibility, and low immunogenicity. Moreover, MSCs' local synthesis of GFs may improve the environment for healing. Animal model studies have proven MSCs' potential for increasing ultimate load failure and resistance to mechanical deformation, enhancing bone quality, and attracting new fibrocartilage formation at the repaired site. Additionally, exogenous hyaluronic acid and MSCs have been showing promising potential to reduce retears.

Bone marrow mesenchymal stem cells

BMSCs enhanced structural integrity of the repair and decreased the risk of retears in a chronic full tear cohort that underwent arthroscopic RC surgery with repeated channelling.⁶⁹ Moreover, partial tear BMSC-enhanced repairs and prevented retears in 10-year follow-ups.⁷⁰ Additionally, arthroscopy surface holding with BMSC stimulation increased RC repair integrity in large chronic tears.⁷¹ BMSCs stimulated by bioactive factors were more effective in promoting tissue regeneration. Platelet-rich plasma (PRP)-infused BMSCs improved GF production, osteogenic differentiation capacity, cell death resistance in-vitro, and biomechanical properties of newly formed bone in-vivo,72 Conversely, structure, composition, and strength of the repaired enthesis remained unchanged, despite BMSCs being present and active metabolically.73 However, this may be due to the short time taken to conduct the study, since stem cells require a longer time to be effective.⁷⁴ Moreover, adenoviral MT1-matrix metalloproteinase (MMP)-augmented MSCs resulted in better fibrocartilage formation, and higher ultimate load and stress to failure after 4-week repair of acute full tears, while long-term effects require further research.⁷⁵

Adipose-derived stem cells

ADSCs have a comparable shape and expression of the cluster differentiation surface marker protein to BMSCs with greater colony-forming and adipogenic capacity. They demonstrated multipotency *in-vitro*; developing into adipose, osteogenic, chondrogenic, and myogenic cells. Four groups were compared in a chronic full rabbit repair model: saline + suture, suture + ADSCs, saline, and ADSCs. ADSC and saline groups failed to heal, while suture + ADSCs group had a larger load to failure and less fatty infiltration than saline + suture group. Moreover, ADSCs imbedded in a fibrin sealant resulted in superior histological and biomechanical outcomes, in acute full murine RC repairs. As for the ideal location to deliver cells, regeneration was found most effective when stem cell sheets were interposed at the enthesis, in chronic full tears.

In contrast, ADSCs in acute and chronic rat RC reconstruction models did not enhance biomechanical characteristics. However, inflammation was reduced, which could potentially lead to a more elastic repair and less scar formation after the healing process. Furthermore, TGF- $\beta 3$ supplementation did not enhance ADSCs' effect on healing, despite its known role in the development of the enthesis. The absence of other mediators that are present during development and the presence of inflammatory mediators may have affected the bioactivity of TGF- $\beta 3$ at the repair time. However, this study did not measure healing at different time points and ADSCs were not labeled to permit analysis of cell retention.

Due to contradictory evidence, it is unclear whether ADSCs constitute a good option to speed RC tendon-bone healing. To demonstrate its value in RC healing, more animal and clinical research is required. Additionally, challenges of availability, seeding, survival, and specificity of such therapies and the ethical regulatory barriers are yet to be overcome for this to reach a clinical trial. **Table 1** summarises the findings.

Cell pathways and rotator cuff enthesis healing *Ihh signaling molecule*

Ihh signaling is active during the initial phases of RC enthesis healing. ^{29,82,83} Fibrocartilage production in an acute full rat RC repair model had higher numbers of Ihh chondrocyte-like cells with MSC augmentation. Increased GLI family zinc finger 1 (Gli1) and Patched1 expression suggests that Ihh signaling pathway controls fibrocartilage production process brought on by stem cells. While both are expressed in fibroblasts of the tendon mid-substance, Ihh is primarily expressed in chondrocytes of the fibrocartilage region, which may indicate a coordinated interaction between chondrocytes and fibroblasts during healing. ⁸² Additionally, immature mice with acute partial tears had a better recovery than adult mice due to the increased density of Gli1⁺ cells near the injured enthesis, mimicking its natural developmental process. ⁸³

Parathyroid hormone related protein

Parathyroid hormone (PTH) has been demonstrated to enhance tissue repair via a chondrogenic pathway.^{84, 85} Osteoblasts speed up tendon-bone healing when PTH binds to its receptor in BMSCs during the tendon-bone healing process. Chen et al.⁸⁶ showed that PTH may influence the tendon-bone healing process by maintaining the proliferation of BMSCs.

Daily systemic PTH injections boosted fibrocartilage development, type-I procollagen-producing cells, and vascularity, which improved collagen fibre structure and mineralised fibrocartilage production in acute full RC tears. Biomechanically, Duchman et al reported higher load to failure in recombinant human PTH-treated acute full RC tear rat model, and expression of intracellular and extracellular vascular endothelial growth factor (VEGF) with controlled recombinant human PTH systemic injections. Clinical investigations of advanced cases with chronic large tears achieved similar outcomes, and reduced retear rates. Table 2 summarises the findings.

Table 1. Summary of cell-based therapies literature findings

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Author	Study	Model	Tear	Intervention	Outcome measure	Results			
Honda et al. ⁶⁸	In-vivo	Rabbit	Chronic full	MSC + HA	Biomechanical, histological, and immunohistochemical analyses	Positive: Improved ultimate load and faster healing			
Jo et al. ⁶⁹	Clinical	Human	Chronic full	BMSC + arthroscopic repeated channeling	Pain scale, ROM, muscle strength, patient satisfaction questionnaire, and functional scores. Structural integrity by MRI and CT	Positive: Enhanced structural integrity of the repair and decreased retears			
Hernigou et al. ⁷⁰	Clinical	Human	Chronic partial	BMSC + arthroscopy	MRI	Positive: Faster complete healing/retear prevention			
Taniguchi et al. ⁷¹	Clinical	Human	Chronic partial	ASH + BMSC	MRI	Positive: Reduced retear rates/better integrity			
Han et al. ⁷²	In-vitro, in-vivo	Rat	Acute full	PRP-infused BMSC	Expression of genes that related to tissue repair, bone formation, and tendon regeneration;	Positive: Stronger signals to angiogenesis, bone formation, and tendon generation <i>in-situ</i>			
					Biomechanical assessment	Promoted healing in-vivo			
Gulotta et al. ⁷³	In-vivo	Rat	Acute full	BMSC	Biomechanical & histological analyses	Negative: No change in structure, composition, or strength			
Gulotta et al. ⁷⁵	In-vivo	Rat	Acute full	MT1-MMP- transduced MSCs	Biomechanical & histological analyses	Positive: Better fibrocartilage formation, higher ultimate load and stress to failure, and higher stiffness			
Oh et al. ⁷⁷	In-vivo	Rabbit	Chronic full	ADSC + suture	Electromyographic, biomechanical & histological analyses	Positive: Larger load to failure and less fat infiltration			
Chen et al. ⁷⁸	In-vivo	Murine	Acute full	ADSC imbedded in fibrin sealant scaffold	Biomechanical & histological analyses	Positive: Better biomechanical strength and histological score			
Choi et al. ⁷⁹	In-vivo	Rat	Chronic full	ADSC sheets interposed at the enthesis	Biomechanical & histological analyses	Positive: Successful complete regeneration and biomechanical strength			
Valencia Mora et al. ⁸⁰	In-vivo	Rat	Chronic full	ADSC, ADSC + TGF-β3	Biomechanical & histological analyses	Positive: Reduced inflammation			
						Negative: Unchanged maximum load, elastic energy, mechanical deformation, and stiffness			

Note: ADSC: adipose-derived stem cell; ASH: arthroscopy surface holding; BMSC: bone marrow mesenchymal stem cell; CT: computed tomography; HA: hyaluronic acid; MMP: matrix metalloproteinase; MRI: magnetic resonance imaging; MSC: mesenchymal stem cell; PRP: platelet-rich plasma; ROM: range of motion; TGF: transforming growth factor.

Growth factors

GFs have been investigated to modulate stem cells in RC enthesis of small and large animal models. $^{90-96}$ Fibroblast growth factor 2 (FGF-2), growth differentiation factor, TGF- β 3, and platelet-derived growth factor are commonly used examples. $^{97-104}$ GFs such as BMP-12, -13, -14, basic fibroblast growth factor, cartilage oligomeric matrix protein, connective tissue growth factor, platelet-derived growth factor-B, and TGF- β 1 have been found to be involved in enthesis healing process, one week after acute full rat supraspinatus repair. By 16 weeks, GF upregulation was back to pre-injury levels, indicating exogenous supplementation of these substances may encourage successful healing in acute

stages. ¹⁰⁵ Angiogenesis and fibroblast proliferation are both powerfully stimulated by basic fibroblast growth factor. It continues to be strongly exhibited throughout the healing process, peaking at 7 and 9 days. ¹⁰⁶ Similar outcomes were attained in acute full sheep model RC tear, where BMP and VEGF induced angiogenesis and vasculogenesis, causing faster and better recovery. ^{43, 107}

Studies investigated TGF- β 3's potential to push scar-mediated healing to a regenerative one. ¹⁰⁸⁻¹¹¹ Sustained delivery of TGF- β 3 via a heparin/fibrin-based system in a rat model improved RC enthesis healing, while still inferior to an uninjured enthesis and disorganised scarring. ¹¹⁰ Conversely, studies reported no improvement in biomechanical properties

Table 2. Summary of signaling molecules therapies literature findings

Author	Study	Model	Tear	Intervention	Outcome measure	Study results
Zong et al. ⁸²	In-vivo	Rat	Acute full	Ihh + MSC	Immunohistochemical staining and proliferating cell nuclear antigen staining	Positive: Increased Gli1 and Patched1 expression. More organised and stronger staining for collagen II
Schwartz et al. ²⁹	In-vivo	Murine	Acute partial	Ihh	Lineage tracing	Positive: Gli1 lineage cells that originate in utero eventually populate the entire mature enthesis. Ablation of the Hh-responsive cells during the first week of postnatal development resulted in a loss of mineralised fibrocartilage
Schwartz et al. ⁸³	In-vivo	Mouse	Acute partial	Ihh	Lineage tracing	Positive: High levels of Gli1 expression in immature mice and mature entheses had fewer Gli1* cells
Hettrich et al. ⁸⁷	In-vivo	Rat	Acute full	Systemic PTH	Histologic, immunohistochemical, biomechanical analyses	Positive: Higher stiffness, bone volume and mineral content; More fibrocartilage, osteoblasts, and blood vessels formation; Better collagen orientation
Duchman et al.88	In-vivo	Rat	Acute full	Systemic rhPTH	Biomechanical and histologic analysis	Positive: Higher load to failure. Expression of intracellular and extracellular VEGF
Oh et al. ⁸⁹	Clinical	Human	Chronic full	Systemic rhPTH	MRI, ROM, American Shoulder and Elbow Surgeons and Constant scores, and simple shoulder test	Positive: Lower retear rate

Note: Gli1: GLI family zinc finger 1; Hh: hedgehog; Ihh: Indian hedgehog; MRI: magnetic resonance imaging; MSC: mesenchymal stem cell; PTH: parathyroid hormone; rhPTH: recombinant human parathyroid hormone; ROM: range of motion; VEGF: vascular endothelial growth factor.

or scar reduction, with TGF- $\beta 3$ and neutralising bodies for TGF- $\beta 1$ and - $\beta 2$ delivered via osmotic pumps, while inhibiting TGF- $\beta 1$ improved RC enthesis quality by reducing fibrosis, fatty infiltration, and muscle atrophy. This however may be due to the delivery method used in the study, as antibodies were not delivered directly to the interface, rather to the bursal surface. Additionally, TGF- $\beta 1$ release could compete for receptors with TGF- $\beta 3$, which leads to reduced effectiveness of TGF- $\beta 3$ treatment. A suggested solution is to combine TGF- $\beta 3$ with a cytokine antagonist and TGF- β -neutralising antibodies or MMP inhibitors for acute full tears.

Recombinant human FGF-18 was investigated for chondrogenic differentiation of BMSCs in a rat acute full tear model and enhanced regeneration in acute full RC tears. Furthermore, tumour necrosis factor (TNF) cytokine has been found to suppress chondrogenic stimulation via nuclear factor-kappa B by downregulating SRY-box 9 expression, whilst TGF- β 3 solely was not ideal. While TNF inhibitor pegylated soluble TNF-R1 increased fibrocartilage and enhanced load to failure and stiffness, no improvement was seen past 8 weeks. 115

Since loading after RC repairs is essential to prevent bone loss at the enthesis attachment, to achieve a mechanically resilient enthesis, collagen fibres within the tendon must attach to bone via mineralised fibrocartilage and remodel the underlying bone. $^{30,\,116}$ Hence, osteoinductive factors (BMP-2–7) are used to induce chondrogenic differentiation and stimulate ECM component synthesis such as proteoglycans and collagen-II. Additionally, BMP-7 improved enthesis matrix maturation in gelatin hydrogel sheets of acute full RC tears. $^{117,\,118}$ Conversely, Rodeo et al. 107 implanted TGF- β 1–3, BMP-2–7, and FGF in a collagen I matrix sponge and compared it to collagen sponge alone, in a sheep acute RC repair. While the study reported greater bone, fibrocartilage, and soft tissue formation, stiffness was less than collagen sponge only group. 107 Although frequently having superior mechanical properties compared to controls, GF-enhanced repairs lack identical biomechanical qualities as native entheses.

GF temporal expression

Despite being present in all stages of healing, GF signal different pathways with different cell types during each stage. BMP-12, -13, -14, bFGF, connective tissue growth factor, platelet-derived growth factor, TGF- β 1, and cartilage oligomeric matrix protein-1 are upregulated at the initial inflammatory stage, and subside by 16 weeks. ¹⁰⁵ The delay of TGF- β 1 upregulation at 8 weeks is correlated with scar formation process. ¹¹⁹ However, BMP-12 is expressed during all three phases, with a marked increase at eight weeks of remodelling stage. This highlights

GF temporal expression involved in various stages of healing, which is essential in developing GFs delivery devices that can be introduced to spatiotemporally target RC repairs. Since

healing process is regulated by multiple GFs, targeting one is not ideal functionally or mechanically.¹⁰⁵ **Table 3** summarises the findings.

Table 3. Summary of growth factor-based therapies literature findings

Author	Study	Model	Tear	Intervention	Outcome measure	Study results
Würgler-Hauri et al. ¹⁰⁵	In-vivo	Rat	Acute full	BMP-12–14, bFGF, COMP, CTGF, PDGFB, TGF-β1	Immunohistochemical staining	Positive: Increase in the expression of all GFs at 1 week, and followed by a return to control or undetectable levels by 16 weeks
Kobayashi et al. ¹⁰⁶	In-vivo	Rabbit	Acute full	BMP-12–14, bFGF, COMP, CTGF, PDGFB, TGF-β1	Light microscopy after staining with hematoxylin-eosin and Elastica-Masson; Immunohistochemical staining	Positive: GFs are involved in early phases of healing promotion
Rodeo et al. ¹⁰⁷	In-vivo	Sheep	Acute full	BMP and VEGF	MRI, plain radiographs, histologic analysis, and biomechanical testing	Positive: Greater formation of new bone, fibrocartilage, and soft tissue, with an increase in tendon attachment strength
Angeline and Rodeo ⁴³	In-vivo	Sheep	Acute full	BMP and VEGF	Histologic analysis	Positive: Induced angiogenesis and vasculogenesis. Faster and better recovery
Manning et al. ¹¹⁰	In-vivo	Rat	Acute full	TGF-β3	Histologic and biomechanical analyses	Negative: Disorganised scar and inferior mechanical properties
Kim et al. ¹⁰⁸	In-vivo	Rat	Acute full	TGF-β3	Histologic and biomechanical analyses	Negative: Disorganised scar and inferior mechanical properties
Davies et al. ¹¹²	In-vivo	Mouse	Acute full	Inhibiting TGF-β1	Histologic analysis	Positive: Reduced fibrosis, fatty infiltration, and muscle atrophy
Jensen et al. ³²	In-vivo	Mouse	Acute full and partial	TGF-β3 + cytokine + MMP inhibitors	Reviewing the literature	Positive: Enhanced healing
Zhou et al. ¹¹³	In-vivo	Rat	Acute full	rhFGF-18	Histologic analysis	Positive: Promoted chondrogenesis and promoted healing and regeneration
Sitcheran et al. ¹¹⁴	In-vivo	Mouse	Acute full	TGF-β3	Histologic analysis	Negative: No improvement in healing
Gulotta et al. ¹¹⁵	In-vivo	Rat	Acute and chronic full	TNF inhibitor	Histologic and biomechanical analyses	Positive: Elevated fibrocartilage, and enhanced load to failure and stiffness
Dorman et al. ¹¹⁷	In-vivo	Mouse	Acute full	BMP-2-7	Histologic analysis	Positive: Fully healed enthesis without toxicity
Kabuto et al. ¹¹⁸	In-vivo	Rat	Acute full	BMP-2-7	Histologic and biomechanical analyses	Positive: Improved biomechanical properties

Note: bFGF: basic fibroblast growth factor; BMP: bone morphogenetic protein; COMP: cartilage oligomeric matrix protein; CTGF: connective tissue growth factor; GF: growth factor; MMP: matrix metalloproteinase; MRI: magnetic resonance imaging; PDGF: platelet derived growth factor; rhFGF: recombinant human fibroblast growth factor; TGF: transforming growth factor; TNF: tumour necrosis factor; VEGF: vascular endothelial growth factor.

Scaffolds and augments

Augmentations via ECM or synthetic materials could potentially optimise RC enthesis healing. Allografts, synthetic polymers, autografts, and xenografts have been used for RC repairs. Despite improvements in xenografts and allografts decellularisation procedures, they still carry the risk of infection and inflammatory responses from residual donor DNA, high degradation rates, inferior mechanical qualities, and non-specific induction abilities. These concerns have sparked

an interest to create grafts and scaffolds for enhanced repairs. Engineered scaffolds with a mix of natural and synthetic biomaterials offer an alternative solution to combat adverse effects caused by synthetic materials. Natural biomaterials including fibrin, collagen, elastin, and hyaluronic acid offer extracellular signals that facilitate cell infiltration and tissue regeneration, where positive results with good host tissue integration, remodelling, and improvement in biomechanical qualities are reported in animal trials employing them. 122, 123

Known to selectively differentiate BMSCs to chondrocytes, and gelatin methacrylol, a novel biomaterial with promising results in drug carrying, enhanced healing. kartogenin-loaded gelatin methacrylol hydrogel scaffold with bone marrow stimulation promoted fibrocartilage formation and resulted in superior mechanical properties in acute full RC repairs. ¹²⁴ Compared to traditional double row suture repairs, engineered tissue grafts with 6 months follow-up showed 11% increased elastic modulus, the repaired enthesis was formed similarly to the native tissue, suggesting potential long-term retear prevention. ⁹³

Exosome-delivered BMP-2 and polyaspartic acid in an acute full RC tear rabbit model resulted in increased tissue mineral density and ultimate load strength, via Smad/RUNX2 signaling pathway. Additionally, tendon regeneration- and cartilage differentiation-related expressions were upregulated, suggesting the positive potential of using bioactive scaffolds in enthesis healing. ¹²⁵ Since stem cells contribute to healing by autocrine/paracrine signaling through GFs and cytokines, delivering GFs via scaffolds is a simpler and more direct approach to improve RC healing. While interleukin-1 beta inhibited chondrogenesis and maturation of MSCs, the mechanical functionality of the tissue was preserved with three-dimensional (3D) woven polycaprolactone scaffolds. ¹²⁶ Highlighting the importance of developing strategies to protect against the deleterious effects of cytokines.

A novel strategy combined a 3D printed polylactide-coglycolide acid scaffold with a cell-laden collagen hydrogel to fabricate layered structures. Mechanical properties improved supporting growth, proliferation, tenogenic differentiation of human ADSCs, and excellent biocompatibility.¹²⁷

As for clinical studies, long-term effect of a polypropylene patch was assessed on large chronic RC repairs and compared with collagen patches, in a patient cohort of 66-year-old. Results showed superior outcomes in muscle strength, pain score, and tendon integrity via ultrasound after a 3-year follow-up. Paper Repairs augmented with 3D biological collagen-I mesh in moderate and large RC tears of patients over 50 years, followed-up for 24-month resulted in better arranged collagen fibres, inflammatory cells less infiltrated, and lower RC retears at 17%. Page 129

Although revealed to lower re-ruptures and tissue oedema in animals, porcine small intestine submucosa has been shown ineffective in humans. ^{130, 131} Due to poor functional results and significant complications, trials cautioned against using small intestine submucosa for RC repairs. ^{131, 132}

In general, scaffolds provide a growth microenvironment for cells and can be used as carriers for seeded cells and GFs, creating optimal conditions for RC enthesis healing. However, those technologies are still tackling the formation of fibrous cartilage, inflammatory reactions, elevated degeneration rates of the grafted scaffolds, and long-term follow-up.

Demineralised bone matrix

Demineralised bone matrix (DBM) is cancellous bone with osteoinductive and osteoconductive characteristics, known

to elicit good adhesion, proliferation, and differentiation of MSCs. ¹³³ DBM-based repairs have not been consistent in improving collagen organisation and fibrocartilage formation, nor result in higher bone mineral density. ¹³⁴ However, supplementing them with BMSCs resulted in superior outcomes in chronic full tears. ¹³⁵ Additionally, the use of DBM-based sponge hydrated with PRP resulted improved strength and histological structure in chronic full tears. ¹³⁶ Despite this, when evaluating the clinical outcomes of patients undergoing biologically enhanced DBM-augmented RC repairs, 50% of patients' magnetic resonance imaging (MRI) confirmed supraspinatus failure. ¹³⁷

A more recent study fabricated a gradient multi-tissue construct that mimics structural, compositional, and cellular heterogeneity of the native enthesis designed by 3D cellprinting and tissue-specific decellularised ECM bioinks, where findings suggested improvement in restoring shoulder function. 138 The first human pilot study was performed to assess the safety and efficacy of autologous dermal fibroblast (ADF) injections on full chronic RC repairs of patients aged 20-80 years. Fibroblast samples were obtained from the patients' gluteal region and cultured for four weeks. Patients from both, the control group who did not have an ADF injection, and experimental group, underwent standardised post-operative rehabilitation program involving ice application for 5 days and tramadol for a week, alongside abduction brace immobilisation for five weeks. Exercises such as shoulder shrug and active elbow and forearm range of motion were encouraged immediately after surgery. 10-12 weeks after surgery activeassisted, active range of motion, and strengthening exercises were performed, with return to sport allowed after 6 months. Successful healing without adverse effects was reported in 1-year follow-up. 139 However, this study was limited by the small number of participants and the optimal dosage of ADF was not established. It is difficult to conclude the effectiveness of DBM-based augments on the overall healing process. Larger studies are necessary to examine its true effectiveness and long-term effect.

Platelet-rich plasma

PRP provides a rich source of GFs and cytokines such as PDGF, TGF-β, FGF, VEGF, and IGF.¹⁴⁰ In a study comparing the clinical and structural outcomes between arthroscopic repair of full thickness RC tears with and without PRP supplementation, retear rates were lower with PRP in tears greater than 3 cm, with improved Constant score.¹⁴¹ Conversely, platelet-rich fibrin clot matrix, a variant of PRP with a fibrin matrix showed no difference in tendon thickness and greater tuberosity coverage in chronic full tears, and positive outcomes in partial tears.¹⁴² However, this could be due to variable PRP preparation used, the nature of PRP used, and level of platelet activation not being accounted.¹⁴³ Thus, future research should focus on controlled PRP formulations to judge the benefits of this intervention more accurately.

Systematic reviews revealed PRP injections intraoperatively to produce better long-term pain and shoulder function outcomes following repairs in chronic full tears. While for

short-term follow-ups, PRP was superior in improving function only, in partial RC tears. 144-147 Post-operative subacromial PRP injections on patients with partial RC tears revealed improvement in functional outcomes up to 6 months, despite magnetic resonance imaging structural outcomes being insignificant, and long-term follow-up not being recorded. 148 Moreover, PRP combined with sodium hyaluronate yielded better outcomes in partial RC tears in younger population after 12 months of follow-up. 149 Recent meta-analyses showed PRP augments with double-row constructs have the best outcomes for chronic full tears, according to visual analogue scale (VAS), Constant, Simple Shoulder Test and retears. 150, 151

Ilhanli et al.¹⁵² debated that PRP may be as effective as physiotherapy interventions in patients with chronic partial tears followed up for 12 months. Despite a study showing

an increase in PRP concentration after exercise, it remains unknown whether physiotherapy could enhance the effect of PRP. ¹⁵³ Moreover, the literature varies in terms of exercise type, intensity, and duration for RC tears. ¹⁵⁴ Standard rehabilitation protocols and ideal platelet concentrations for PRP augmented repairs are essential to preserve outcomes and create tailored care for patients with different backgrounds and abilities.

PRP is considered a suitable alternative to corticosteroid injections, which are commonly used clinically to manage RC tears. Achieving pain relief and good clinical outcomes add to the benefit of avoiding the adverse effects of corticosteroids on the body, which makes it an alternative approach to managing symptomatic RC tears, in both; young and athletic, and older populations with comorbidities. **Table 4** summarises the findings.

Table 4. Summary of scaffold-based therapies literature findings

Author	Study	Model	Tear	Intervention	Outcome measure	Study results
Huang et al. ¹²⁴	In-vivo	Rabbit	Acute full	KGN-loaded GelMA hydrogel + BMSC scaffold	Macroscopy, microcomputed tomography, histology, and biomechanical tests	Positive: Promoted fibrocartilage formation and superior mechanical properties
Novakova et al. ⁹³	In-vivo	Sheep	Acute full	Engineered tendon construct with BMSCs	X-ray and biomechanical tests	Positive: Native-like enthesis with higher modulus
Han et al. ¹²⁵	In-vivo	Rabbit	Acute full	BMP-2 + polyaspartic acid + Smad/RUNX2 signaling	Transmission electron microscopy staining; Biomechanics and histological assessment	Positive: Increased bone and tissue mineral density and ultimate load strength
Ousema et al. ¹²⁶	In-vitro	RC tear		3D woven PCL scaffold + IL-1 inhibition on MSCs	Histological, biomechanical, and immunohistochemistry analyses	Positive: Mechanical functionality preserved with the use of a 3D woven PCL scaffold
Jiang et al. ¹²⁷	In-vitro	RC tear		3D PLGA scaffold + a cell-laden collagen hydrogel + ADSCs	Histological and biomechanics analyses	Positive: Improvement in mechanical properties and biocompatibility
Iannotti et al. ¹³¹	Clinical	Human	Chronic full	SIS	Penn shoulder-score questionnaire and MRI	Negative: No improvement in healing and clinical results
Malcarney et al. ¹³²	Clinical	Human	Chronic full	SIS	Study discontinued due to adverse effects	Negative: Inflammatory reaction
Sclamberg et al. ¹³⁰	Clinical	Human	Chronic full	SIS	Patient questionnaire, MRI, and ASES	Negative: No improvement and worse pot-operative outcomes
Ciampi et al. ¹²⁸	Clinical	Aging human	Chronic full	Polypropylene augmentation patch	Ultrasound, muscle strength, and VAS	Positive: Improved muscle strength, pain score, and tendon integrity
Cai et al. ¹²⁹	Clinical	Aging human	Chronic full	3D biological collagen-I mesh	MRI, VAS, UCLA SST, and Constant score	Positive: Less retear rates
Hoberman et al. ¹³³	In-vitro	RC tear		DBM + BMSCs + PRP	Adhesion, proliferation, and differentiation assays	Positive: Better adhesion, proliferation, and differentiation
Thangarajah et al. ¹³⁴	In-vivo	Rat	Chronic full	DBM	Histological analysis	Negative: No improvement in collagen organisation and fibrocartilage formation
Thangarajah et al. ¹³⁵	In-vivo	Rat	Chronic full	DBM + MSCs	Histological analysis	Positive: Enhanced healing

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Table 4. Continued

Author	Study	Model	Tear	Intervention	Outcome measure	Study results
Smith et al. ¹³⁶	In-vivo	Canine	Chronic full	PRP + DBM	Histological and biomechanical analysis	Positive: Improvement in strength and histological structure
Wellington et al. ¹³⁷	Clinical	Human	Chronic full	DBM + MSCs	MRI	Negative: Supraspinatus failure
Chae et al. ¹³⁸	In-vivo	Mouse	Chronic full	3D cell-printed tendon-bone interface construct	Gait analysis, histological and biomechanical analysis	Positive: Fully formed enthesis, improved shoulder outcome and biomechanical properties
Yoon et al. ¹³⁹	Clinical	Aging and young human	Chronic full	ADF	ROM, VAS, and MRI	Positive: No adverse effects, improved VAS, and functional scores
Warth et al. ¹⁴¹	Clinical	Human	Chronic full	PRP	MRI and Constant score	Positive: Lower retear and improved constant score
Castricini et al. ¹⁴²	Clinical	Human	Chronic partial	Autologous PRFM	MRI	Positive: Improved tendon integrity
			Chronic full	Autologous PRFM	MRI	Negative: No difference in constant score and tendon integrity
Giovannetti de Sanctis et al. ¹⁴⁴	Systematic review	Human	Chronic partial	PRP	Shoulder function and VAS	Positive: Improved pain and shoulder function
Von Wehren et al. ¹⁴⁵	Clinical	Human	Chronic partial	PRP	MRI, Constant score, ASES, shoulder ROM, and VAS	Positive: Improved pain and function
Xu and Xue ¹⁴⁶	Systematic review	Human	Chronic full	PRP	Retear rate, Constant, UCLA, ASES, VAS, and adverse effects	Positive: Improved shoulder outcome and reduced retear rate
Rha et al. ¹⁴⁷	Clinical	Human	Chronic partial	PRP	Shoulder Pain and Disability Index, ROM, and ultrasound	Positive: No adverse effects, and improved shoulder pain and function
Shams et al. ¹⁴⁸	Clinical	Human	Chronic partial	PRP	MRI, ASES, Constant Score, SST, and VAS	Positive: Improved shoulder function and minor MRI improvement
Cai et al. ¹⁴⁹	Clinical	Young human	Acute partial	SH + PRP	VAS, Constant score, and MRI	Positive: Better VAS, constant score, and MRI findings
Ryan et al. ¹⁵⁰	Systematic review	Human	Chronic full	PRP	Constant, ASES, UCLA, SST, VAS, and retear rate	Positive: Reduced retear rates and improved clinical outcomes
Lavoie-Gagne et al. ¹⁵¹	Systematic review and meta- analysis	Human	Chronic full	PRP	Clinical characteristics, retear rates, ROM, and patient reported outcomes	Positive: Reduced retear rates and improved clinical outcomes
Ilhanli et al. ¹⁵²	Clinical	Human	Chronic partial	PRP	ROM, VAS, Disabilities of Arm, Shoulder and Hand questionnaire, Neer's, Hawkins' and drop arm tests and Beck Depression Inventory questionnaire	Negative: Results were not superior to physiotherapy

Note: 3D: three-dimensional; ADF: autologous dermal fibroblast; ADSC: adipose-derived stem cell; ASES: American Shoulder and Elbow Surgeons; BMP-2: bone morphogenetic protein-2; BMSC: bone marrow mesenchymal stem cell; DBM: demineralised bone matrix; GelMA: gelatin methacrylol; IL-1: interleukin-1; KGN: kartogenin; MRI: magnetic resonance imaging; PCL: polycaprolactone; PLGA: polylactide-co-glycolide acid; PRFM: platelet-rich fibrin clot matrix; PRP: platelet-rich plasma; RC: rotator cuff; ROM: range of motion; SH: sodium hyaluronate; SIS: small intestine submucosa; SST: Simple Shoulder Test; UCLA: University of California Los Angeles; VAS: visual analogue scale.

Biomechanical Loading Response

Loading tendons is critical for the maintenance of developing entheses, and tissue deformity, delayed mineralisation, and porous non-mineralised tissues are observed when unloaded. Mechanical stimulation increases TGF- $\beta1$ and Prrx1 $^+$ cells to stimulate enthesis repair. 155 Unloading affects chondrocyte hypertrophy, leading to fibrocartilage absence at the insertion by eight weeks of immobilisation. 36 Reduced humeral volume with flatter surface and morphological changes comparable to those with cerebral palsy were noted, which is attributed to an increase in osteoclasts activity. Hence, blocking osteoclasts activity by bisphosphonate drugs partially recovers bone mineralisation and volume. 36 , 37 This imbalance, therefore, contributes to developing an overall mechanically inferior tissue.

While prolonged and immediate mechanical loading induced adverse effects, low-intensity loading was beneficial as opposed to complete removal. Since immediate excessive and prolonged loading produce adverse effects, and removal causes malformation, a requirement for balance between loading and healing is essential. When compared to various rehabilitative regimens, simple progressive loading is ideal.

Discussion

RC tears are one of the most common causes of pain and dysfunctions in the shoulders, occurring in 9% under the age of 20 years to 62% until 80 years old. 160 Overhead-sport athletes are at higher risk of chronic RC injuries, due to repetitive loads leading to microtraumas, while contact-sport athletes have a higher risk of traumatic acute tears. 161 As for the aging population, degenerative pathologies are the most prevalent mechanism of injury. 162 Older patients with chronic tears would benefit from non-operative approaches, and younger athletes presenting acute tears require surgery, due to expectations of returning to competition. However, competitive athletes record lower return rates compared to recreational ones. 163 The reason for this remains multifactorial and may not be directly related to the repair, but rather a result of psychological factors such as fear of injury reoccurrence. 164, 165

RC repairs present good short-term pain and function improvement, and high retears in the long-term. ¹⁶⁶ Failure of complete healing urges the shift to alternative strategies aiming at restoring the native tissue's structure and biomechanical properties. Since the RC enthesis is made of a variety of cellular components organised in a gradient complex ECM structure, its regeneration requires the co-regulation of multiple factors. ¹⁶⁷

Stem cell role

The effectiveness of using stem cells to promote RC enthesis healing lies in regulating the differentiation of stem cells into target cells. This regulation is affected by certain factors: (1) the interaction between cells, (2) GFs and signaling molecules in promoting their expression, and (3) local microenvironment and mechanical stimulation. Obtaining stem cells and their survival rate are issues faced when used for RC enthesis healing, hence, improving this can broaden their use in hypoxic conditions. Additionally, methods in which stem cells are taken from patients can reduce patient inconvenience.¹⁶⁷

Growth factor role

GFs regulate RC enthesis healing in several ways: (1) different GFs are expressed in stages of healing, with GFs promoting angiogenesis present in the inflammatory stage, and those endorsing cell differentiation for collagen synthesis at the repair and remodelling stages.¹⁶⁸ (2) As a signaling molecule, by promoting the expression of genes and proteins related to regulate the biological behavior of cells. (3) By interacting with other GFs. It is essential to consider that: (1) GFs have a short half-life, thus carriers to delay their local release rate and maintain their effect are essential. (2) GFs have different effects based on the tear type. (3) The role of some GFs and their signaling pathways remains unclear. Considerable variations persist between studies in the preparation of GFs (recombinant, synthetic), delivery methods (local injections, systemic, within scaffold, coated sutures), and timing (pre-, peri-, post-operative, and repeated dosages).

Scaffold's role

The choice of scaffold material should meet the growing needs of specific cells, simulate its ECM, mechanically support, and provide a suitable growth environment. Since the main component at the RC insertion is collagen, the application of biomaterials with collagen matrix as the main component is the most effective. Patches designs are essential to assure its proper interaction with the microenvironment for proliferation, adhesion, differentiation, and morphology. 169-172

Summarised outcomes

Findings suggested a lack of single optimal strategy for restoring RC enthesis. Mechanical stimulation is necessary; its absence cause detrimental effects, and long-term progressive loading rehabilitation aids in achieving a comprehensive RC repair. 16, 36, 37, 155, 157, 159

Augmenting MSCs with adenoviral MT1-MMP had positive biomechanical outcomes in acute full tears 75 (**Figure 2**). ADSCs present contradicting outcomes in acute and chronic full tears, and BMSCs were recorded as negative in acute. $^{73,\,77,\,80,\,81}$ However infusing BMSCs with PRP and imbedding ADSCs in fibrin sealant scaffold enhance repairs. $^{72,\,78}$ Conversely, TGF- β 3-supplemented ADSCs did not enhance acute full tear healing. 80 For chronic full ones, MSCs combined with hyaluronic acid, BMSCs with repeated channeling, and ADSC sheets interposed improved outcomes. $^{68,\,69,\,79}$ Clinical trials in chronic partial tears with BMSC-enhanced augmentation improved healing and prevented retears. $^{70,\,71}$

For signaling molecules, systemic PTH produced positive outcomes in full acute and chronic tears. ⁸⁷⁻⁸⁹ Ihh with MSCs had positive outcomes in acute partial and full tears. ^{29, 82, 83} BMP-12–14, basic fibroblast growth factor, COMP, connective tissue growth factor, PDGFB, BMP-2–7, recombinant human FGF-18, TGF- β 1, and the combination of BMP and VEGF resulted positively, while TGF- β 3 remained ineffective in acute full tears. ^{43, 105-107, 110, 113, 114, 117, 118} Inhibiting TGF- β 1 was effective in acute full tears, and TNF inhibition was better in full acute and chronic tears. ^{112, 115} Most promising outcomes were attained by combining TGF- β 3 with a cytokine and MMP inhibitor in acute partial and full tears. ³²

Acute – Full	Chronic – Full	Chronic – Partial	Chronic – full – age	In-vitro	Acute - Partial	Acute – Partial –Young	Chronic – full – Young
BMP-12-14 ^{104, 105}	ADSC + suture ⁷⁶	Autologous PRFM (Human) ¹⁴⁰	ADF (Human) ¹³⁷	DBM + BMSCs + PRP ¹³¹	TGF-β + cytokine + MMP inhibitors ³²	SH + PRP (Human) ¹⁴⁷	ADF (Human) ¹³⁷
bFGF ^{104, 105}	MSC + HA ¹⁷²	BMSC ^{69, 70}	3D biological collagen-I mesh ¹²⁷	3D PLGA scaffold + a cell- laden collagen hydrogel + ADSCs ¹²⁵	lhh ^{29, 81}		
COMP ^{104, 105}	PRP + DBM ¹³⁴	PRP ^{142, 143, 145, 146}	Polypropylene ¹²⁶	3D woven PCL scaffold + IL-1 inhibition on MSCs ¹²⁴			
CTGF ¹⁰¹ , 103		PRP (Human) ¹⁵⁰			•		
PDGFB ^{104, 105}	ADSC ^{78, 79}						
TGF-β1 ^{104, 105}	DBM + MSCs ¹³³						
	PRP (Human) ^{139, 144, 148, 149}						
Smad/RUNX2 signaling ¹²⁴	Controlled systemic PTH injections ⁸⁷						
Inhibiting TGF-β1 ¹¹¹	TNF inhibitor ¹¹³						
124	3D cell-printed tendon- bone interface construct 136						
MMP inhibitors ³²	BMSC with repeated channeling ⁶⁸						
	ADSC ⁷⁸						
Systemic PTH injections ^{86,87}	SIS ¹²⁸⁻¹³⁰						
BMSCs ⁹²	DBM ¹³²						
BMP2-7 ^{116, 117}	DBM + MSCs (Human) ¹³⁵						
01	Autologous PRFM (Human) ¹⁴⁰						
MSC + Ihh ⁸¹		•					
ADSC imbedded in fibrin sealant scaffold ⁷⁷							
rhFGF-18 ¹¹²							
MT1-MMP- transduced MSCs ⁷⁴							
KGN-loaded GelMA hydrogel + BMSCs							
scaffold ¹²³ BMSC ⁷² ADSC ^{79,80}							
ADSC + TGF-β3 ⁷⁹ TGF-β3 ^{107, 109, 113}							

Figure 2. Categorised literature findings: stem cells, GFs, and scaffolds vary in effect depending on the extent of the tear, chronicity level, population, and their combinations. Majority of studies investigated acute full tears, while less studies were investigating partial and full chronic tears. Very few studies considered the age of the studied population. 3D: three-dimensional; ADF: autologous dermal fibroblast; ADSC: adipose-derived stem cell; bFGF: basic fibroblast growth factor; BMP: bone morphogenetic protein; BMSC: bone marrow mesenchymal stem cell; COMP: cartilage oligomeric matrix protein; CTGF: connective tissue growth factor; DBM: demineralised bone matrix; GelMA: gelatin methacrylol; GF: growth factor; HA: hyaluronic acid; lhh: Indian hedgehog; KGN: kartogenin; MMP: matrix metalloproteinase; MSC: mesenchymal stem cell; PCL: polycaprolactone; PDGFB: platelet-derived growth factor-B; PLGA: polylactide-coglycolide acid; PRFM: platelet-rich fibrin clot matrix; PRP: platelet-rich plasma; PTH: parathyroid hormone; SH: sodium hyaluronate; SIS: small intestine submucosa; TGF: transforming growth factor; TNF: tumour necrosis factor.

Outcomes from scaffold-based studies are promising and reached clinical trials. Polypropylene augmentation patch resulted in good outcomes in aging models with chronic full tears. 128 3D cell-printed tendon-bone interface construct revealed positive outcomes in chronic full tears contrasting small intestine submucosa. 130-132,138 Kartogenin-loaded gelatin methacrylol with BMSCs, engineered constructs with BMSCs, and BMP-2 with polyaspartic acid and Smad/RUNX2 improved acute full tears. 93, 124, 125 DBM-based scaffold was not beneficial, however, augmenting it with MSCs and PRP in separate animal models was effective. 134-136 Clinical trials on DBM and MSCs combination did not replicate similar outcomes.¹³⁷ This could be attributed to the varied forms of scaffold materials, since animal models have shown promising results with strips of cancellous DBM, while clinical studies used sponges. Sponges can be displaced from the interface site, whereas strips and patches can be more securely sutured into the interface.

Autologous platelet-rich fibrin clot matrix clinical studies were more beneficial in chronic partial than full tears. 142

ADF was safe and effective in younger and older populations with chronic full tears. ¹³⁹ While PRP is proven beneficial in chronic partial and full tears, it wasn't superior to physiotherapy. ^{126, 141, 144-148, 150-152} Sodium hyaluronate combined PRP was effective in acute and partial tears of younger populations. ¹⁴⁹ 3D collagen I mesh augmentation in older populations with chronic full tears resulted in positive long-term outcomes. ¹²⁹ Novel multifactorial lab-based tissue engineering strategies such as 3D-woven polycaprolactone scaffold with IL-1 inhibition on MSCs, 3D polylactide-coglycolide acid scaffold in cell-laden collagen hydrogel and ADSCs, BMSCs and PRP enhanced DBM-based scaffold had promising outcomes. ^{126, 127, 133}

Potential reasons for variations

Outcomes vary since there is no standardised controlled environment prior and after repairs. They differ in methodologies, animal models, and mechanical followups; some allow free mobilisation after repairs, while others impose treadmill walking regimens or restrict movement.

It is difficult to determine if variations are due to the interventions, animal models, mechanical environment, or extent and mechanism of tears. Some studies created acute dissection or detachment, immediately injecting it with the examined substances prior to repairs. However, that does not represent the clinical acute and chronic mechanism of tear, as it does not allow time for the inflammatory process to take place; no blood supply restriction, enthesis degeneration, and fatty infiltration. Other studies subject animals to a treadmill regimen prior to the intervention to exhaust tendons and replicate overuse. Findings should be similar in both scenarios, to indicate that the intervention has a definitive outcome. 173 Treadmill regimen does not ideally replicate injury process in athletes performing sports since childhood. Professionally trained athletes have a lower risk of RC injury compared to amateurs that lack proper loading techniques, healthy balanced diet, sleep routine, and psychological well-being. This might fit amateur athletes scenario who are exposed to improper unconditioned loading. Additionally, small animal models such as rats and rodents have a far better healing capacity than larger animals and humans. Their measures of success should not be solely determined based on morphology, but also the timeframe and long-term effects. Surgical techniques used also affect the outcome of biological therapies. Introducing stem cells via tunnels produced superior outcomes to surface repairs. Additionally, biomaterial patch that covered the repair addressed MSCs long-term survival. 174-176

Clinical outcomes are affected by patients' occupation, demographics, age, gender, comorbidities, and tear mechanism and extent. Moreover, patient outcome measures have a degree of subjectivity, affected by expectations and psychosocial wellbeing.

Conclusion

To take research of tendon-bone enthesis for RC tears forward, there are critical factors involved in the healing of the enthesis, which is supported by what has been presented and rationale for, where without them there wouldn't be a successful healing. Tissue engineering has promising outcomes in achieving the complex RC enthesis structure and function. Different cells and GFs at specific times and environmental control can maximise the restoration of the native enthesis. Despite the positive outcomes of tissue engineering strategies, progressive loading environment is essential for optimum healing. Although the effects of the discussed interventions have mostly been confirmed in animal experiments, the intrinsic connections between cells at the interface and the molecular signaling pathways involved in tissue repair and regeneration still need to be further explored in humans. Nevertheless, the limitations mentioned are important to be considered for future studies to be applied in clinical practice.

This paper described the microscopic anatomy of the healthy RC enthesis, its role in facilitating force transmission, and the biomechanical adaptations. It then discussed the structural changes that accompany injured RC enthesis and the factors influencing its healing potential. Positive outcomes in the field of stem cells, GFs, and scaffolds along with their challenges,

limitations, and variety of implementation are explained. The results support the proposed hypothesis that interventions should be tailored according to the tear extent, chronicity, and population studied.

Study limitations

Inconsistencies in reported outcomes derive from variations in patients' occupation, demographics, age, gender, comorbidities, and tear mechanism and extent. Moreover, patient reported outcome measures are subjective, and influenced by expectations and psychosocial well-being. Perspective studies would benefit from addressing the following:

- 1. The difficulties in advancing *in vivo* studies to clinical trials and the regulatory constraints associated with utilising biological interventions which have proven successful in the former, to the latter.
- 2. Biological materials availability and manufacturing in forms that can be utilised clinically in RC repair.
- 3. The potential for personalised biological intervention materials.
- 4. Solutions for aging patients and professional athletes requiring 100% recovery to return to competition.
- 5. Influential medical, environmental, social, and psychological factors.

Author contributions

Both authors contributed to the design, literature research, analysis, and preparation of the manuscript. Both authors revised the manuscript and approved the final version of the manuscript.

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Conflicts of interest statement

Both authors declare that they have no competing interests.

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- Liu, Y. X.; Thomopoulos, S.; Birman, V.; Li, J. S.; Genin, G. M. Bi-material attachment through a compliant interfacial system at the tendon-to-bone insertion site. *Mech Mater.* 2012, 44, 10.1016/j. mechmat.2011.1008.1005.
- Thomopoulos, S.; Williams, G. R.; Gimbel, J. A.; Favata, M.; Soslowsky, L. J. Variation of biomechanical, structural, and compositional properties along the tendon to bone insertion site. *J Orthop Res.* 2003, 21, 413-419.
- Shaw, H. M.; Benjamin, M. Structure-function relationships of entheses in relation to mechanical load and exercise. *Scand J Med Sci Sports.* 2007, 17, 303-315.
- 4. Lu, H. H.; Thomopoulos, S. Functional attachment of soft tissues to bone: development, healing, and tissue engineering. *Annu Rev Biomed Eng.* **2013**, *15*, 201-226.

 Milgrom, C.; Schaffler, M.; Gilbert, S.; van Holsbeeck, M. Rotator-cuff changes in asymptomatic adults. The effect of age, hand dominance and gender. J Bone Joint Surg Br. 1995, 77, 296-298.

- Yamaguchi, K.; Ditsios, K.; Middleton, W. D.; Hildebolt, C. F.; Galatz, L. M.; Teefey, S. A. The demographic and morphological features of rotator cuff disease. A comparison of asymptomatic and symptomatic shoulders. J Bone Joint Surg Am. 2006, 88, 1699-1704.
- Galatz, L. M.; Ball, C. M.; Teefey, S. A.; Middleton, W. D.; Yamaguchi, K. The outcome and repair integrity of completely arthroscopically repaired large and massive rotator cuff tears. *J Bone Joint Surg Am.* 2004, 86, 219-224.
- Shindle, M. K.; Chen, C. C.; Robertson, C.; DiTullio, A. E.; Paulus, M. C.; Clinton, C. M.; Cordasco, F. A.; Rodeo, S. A.; Warren, R. F. Full-thickness supraspinatus tears are associated with more synovial inflammation and tissue degeneration than partial-thickness tears. J Shoulder Elbow Surg. 2011, 20, 917-927.
- Benjamin, M.; Qin, S.; Ralphs, J. R. Fibrocartilage associated with human tendons and their pulleys. J Anat. 1995, 187 (Pt 3), 625-633.
- Benjamin, M.; Kumai, T.; Milz, S.; Boszczyk, B. M.; Boszczyk, A. A.;
 Ralphs, J. R. The skeletal attachment of tendons--tendon "entheses".
 Comp Biochem Physiol A Mol Integr Physiol. 2002, 133, 931-945.
- 11. Frowen, P.; Benjamin, M. Variations in the quality of uncalcified fibrocartilage at the insertions of the extrinsic calf muscles in the foot. *J Anat.* **1995**, *186* (Pt 2), 417-421.
- 12. Matyas, J. R.; Anton, M. G.; Shrive, N. G.; Frank, C. B. Stress governs tissue phenotype at the femoral insertion of the rabbit MCL. *J Biomech.* **1995**, *28*, 147-157.
- Yang, P. J.; Temenoff, J. S. Engineering orthopedic tissue interfaces. Tissue Eng Part B Rev. 2009, 15, 127-141.
- 14. Benjamin, M.; Ralphs, J. R. Fibrocartilage in tendons and ligaments--an adaptation to compressive load. *J Anat.* **1998**, *193* (*Pt 4*), 481-494.
- Connizzo, B. K.; Adams, S. M.; Adams, T. H.; Jawad, A. F.; Birk, D. E.; Soslowsky, L. J. Multiscale regression modeling in mouse supraspinatus tendons reveals that dynamic processes act as mediators in structurefunction relationships. *J Biomech.* 2016, 49, 1649-1657.
- Schwartz, A. G.; Lipner, J. H.; Pasteris, J. D.; Genin, G. M.;
 Thomopoulos, S. Muscle loading is necessary for the formation of a functional tendon enthesis. *Bone.* 2013, 55, 44-51.
- 17. Knese, K. H.; Biermann, H. Osteogenesis in tendon and ligament insertions in the area of the original chondral apophyses. *Z Zellforsch Mikrosk Anat.* **1958**, *49*, 142-187.
- Thomopoulos, S.; Hattersley, G.; Rosen, V.; Mertens, M.; Galatz,
 L.; Williams, G. R.; Soslowsky, L. J. The localized expression of extracellular matrix components in healing tendon insertion sites: an in situ hybridization study. J Orthop Res. 2002, 20, 454-463.
- Galatz, L. M.; Sandell, L. J.; Rothermich, S. Y.; Das, R.; Mastny, A.; Havlioglu, N.; Silva, M. J.; Thomopoulos, S. Characteristics of the rat supraspinatus tendon during tendon-to-bone healing after acute injury. *J Orthop Res.* 2006, 24, 541-550.
- Ralphs, J. R.; Tyers, R. N.; Benjamin, M. Development of functionally distinct fibrocartilages at two sites in the quadriceps tendon of the rat: the suprapatella and the attachment to the patella. *Anat Embryol (Berl)*. 1992, 185, 181-187.
- 21. Rufai, A.; Benjamin, M.; Ralphs, J. R. Development and ageing of phenotypically distinct fibrocartilages associated with the rat Achilles tendon. *Anat Embryol (Berl)*. **1992**, *186*, 611-618.
- Blitz, E.; Viukov, S.; Sharir, A.; Shwartz, Y.; Galloway, J. L.; Pryce, B.
 A.; Johnson, R. L.; Tabin, C. J.; Schweitzer, R.; Zelzer, E. Bone ridge

- patterning during musculoskeletal assembly is mediated through SCX regulation of Bmp4 at the tendon-skeleton junction. *Dev Cell.* **2009**, *17*, 861-873.
- 23. Blitz, E.; Sharir, A.; Akiyama, H.; Zelzer, E. Tendon-bone attachment unit is formed modularly by a distinct pool of Scx- and Sox9-positive progenitors. *Development.* **2013**, *140*, 2680-2690.
- Galatz, L.; Rothermich, S.; VanderPloeg, K.; Petersen, B.; Sandell, L.; Thomopoulos, S. Development of the supraspinatus tendon-to-bone insertion: localized expression of extracellular matrix and growth factor genes. *J Orthop Res.* 2007, 25, 1621-1628.
- Ito, Y.; Toriuchi, N.; Yoshitaka, T.; Ueno-Kudoh, H.; Sato, T.;
 Yokoyama, S.; Nishida, K.; Akimoto, T.; Takahashi, M.; Miyaki, S.;
 Asahara, H. The Mohawk homeobox gene is a critical regulator of tendon differentiation. *Proc Natl Acad Sci U S A*. 2010, 107, 10538-10542.
- Schweitzer, R.; Chyung, J. H.; Murtaugh, L. C.; Brent, A. E.; Rosen, V.; Olson, E. N.; Lassar, A.; Tabin, C. J. Analysis of the tendon cell fate using Scleraxis, a specific marker for tendons and ligaments. *Development.* 2001, 128, 3855-3866.
- Sugimoto, Y.; Takimoto, A.; Akiyama, H.; Kist, R.; Scherer, G.; Nakamura, T.; Hiraki, Y.; Shukunami, C. Scx+/Sox9+ progenitors contribute to the establishment of the junction between cartilage and tendon/ligament. *Development.* 2013, 140, 2280-2288.
- Killian, M. L.; Thomopoulos, S. Scleraxis is required for the development of a functional tendon enthesis. FASEB J. 2016, 30, 301-311
- Schwartz, A. G.; Long, F.; Thomopoulos, S. Enthesis fibrocartilage cells originate from a population of Hedgehog-responsive cells modulated by the loading environment. *Development*. 2015, 142, 196-206.
- 30. Dyment, N. A.; Breidenbach, A. P.; Schwartz, A. G.; Russell, R. P.; Aschbacher-Smith, L.; Liu, H.; Hagiwara, Y.; Jiang, R.; Thomopoulos, S.; Butler, D. L.; Rowe, D. W. Gdf5 progenitors give rise to fibrocartilage cells that mineralize via hedgehog signaling to form the zonal enthesis. *Dev Biol.* 2015, 405, 96-107.
- Dyment, N. A.; Hagiwara, Y.; Matthews, B. G.; Li, Y.; Kalajzic, I.;
 Rowe, D. W. Lineage tracing of resident tendon progenitor cells during growth and natural healing. *PLoS One.* 2014, 9, e96113.
- Jensen, P. T.; Lambertsen, K. L.; Frich, L. H. Assembly, maturation, and degradation of the supraspinatus enthesis. J Shoulder Elbow Surg. 2018, 27, 739-750.
- 33. Santoni, B. G.; McGilvray, K. C.; Lyons, A. S.; Bansal, M.; Turner, A. S.; Macgillivray, J. D.; Coleman, S. H.; Puttlitz, C. M. Biomechanical analysis of an ovine rotator cuff repair via porous patch augmentation in a chronic rupture model. *Am J Sports Med.* 2010, *38*, 679-686.
- 34. Mori, D.; Funakoshi, N.; Yamashita, F.; Wakabayashi, T. Effect of fatty degeneration of the infraspinatus on the efficacy of arthroscopic patch autograft procedure for large to massive rotator cuff tears. *Am J Sports Med.* **2015**, *43*, 1108-1117.
- Hakimi, O.; Mouthuy, P. A.; Zargar, N.; Lostis, E.; Morrey, M.; Carr,
 A. A layered electrospun and woven surgical scaffold to enhance endogenous tendon repair. *Acta Biomater.* 2015, 26, 124-135.
- Thomopoulos, S.; Kim, H. M.; Rothermich, S. Y.; Biederstadt, C.; Das, R.; Galatz, L. M. Decreased muscle loading delays maturation of the tendon enthesis during postnatal development. *J Orthop Res.* 2007, 25, 1154-1163.
- 37. Tatara, A. M.; Lipner, J. H.; Das, R.; Kim, H. M.; Patel, N.; Ntouvali, E.; Silva, M. J.; Thomopoulos, S. The role of muscle loading on bone (Re) modeling at the developing enthesis. *PLoS One.* **2014**, *9*, e97375.
- 38. Rhee, Y. G.; Cho, N. S.; Yoo, J. H. Clinical outcome and repair integrity

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- after rotator cuff repair in patients older than 70 years versus patients younger than 70 years. *Arthroscopy.* **2014**, *30*, 546-554.
- Melis, B.; Nemoz, C.; Walch, G. Muscle fatty infiltration in rotator cuff tears: descriptive analysis of 1688 cases. Orthop Traumatol Surg Res. 2009, 95, 319-324.
- Meyer, D. C.; Farshad, M.; Amacker, N. A.; Gerber, C.; Wieser, K.
 Quantitative analysis of muscle and tendon retraction in chronic rotator cuff tears. Am J Sports Med. 2012, 40, 606-610.
- 41. Saadat, F.; Deymier, A. C.; Birman, V.; Thomopoulos, S.; Genin, G. M. The concentration of stress at the rotator cuff tendon-to-bone attachment site is conserved across species. *J Mech Behav Biomed Mater.* **2016**, *62*, 24-32.
- 42. Kanazawa, T.; Gotoh, M.; Ohta, K.; Honda, H.; Ohzono, H.; Shimokobe, H.; Shiba, N.; Nakamura, K. Histomorphometric and ultrastructural analysis of the tendon-bone interface after rotator cuff repair in a rat model. *Sci Rep.* **2016**, *6*, 33800.
- Angeline, M. E.; Rodeo, S. A. Biologics in the management of rotator cuff surgery. Clin Sports Med. 2012, 31, 645-663.
- Harryman, D. T., 2nd; Mack, L. A.; Wang, K. Y.; Jackins, S. E.; Richardson, M. L.; Matsen, F. A., 3rd. Repairs of the rotator cuff. Correlation of functional results with integrity of the cuff. *J Bone Joint Surg Am.* 1991, 73, 982-989.
- 45. Liem, D.; Lichtenberg, S.; Magosch, P.; Habermeyer, P. Magnetic resonance imaging of arthroscopic supraspinatus tendon repair. *J Bone Joint Surg Am.* **2007**, *89*, 1770-1776.
- Charousset, C.; Bellaïche, L.; Kalra, K.; Petrover, D. Arthroscopic repair of full-thickness rotator cuff tears: is there tendon healing in patients aged 65 years or older? *Arthroscopy.* 2010, 26, 302-309.
- Cho, N. S.; Rhee, Y. G. The factors affecting the clinical outcome and integrity of arthroscopically repaired rotator cuff tears of the shoulder. *Clin Orthop Surg.* 2009, 1, 96-104.
- Nho, S. J.; Shindle, M. K.; Adler, R. S.; Warren, R. F.; Altchek, D. W.; MacGillivray, J. D. Prospective analysis of arthroscopic rotator cuff repair: subgroup analysis. J Shoulder Elbow Surg. 2009, 18, 697-704.
- 49. Oh, J. H.; Kim, S. H.; Kang, J. Y.; Oh, C. H.; Gong, H. S. Effect of age on functional and structural outcome after rotator cuff repair. *Am J Sports Med.* **2010**, *38*, 672-678.
- Tashjian, R. Z.; Hollins, A. M.; Kim, H. M.; Teefey, S. A.; Middleton, W. D.; Steger-May, K.; Galatz, L. M.; Yamaguchi, K. Factors affecting healing rates after arthroscopic double-row rotator cuff repair. *Am J Sports Med.* 2010, *38*, 2435-2442.
- Boileau, P.; Brassart, N.; Watkinson, D. J.; Carles, M.; Hatzidakis, A. M.; Krishnan, S. G. Arthroscopic repair of full-thickness tears of the supraspinatus: does the tendon really heal? *J Bone Joint Surg Am.* 2005, 87, 1229-1240.
- Thomazeau, H.; Boukobza, E.; Morcet, N.; Chaperon, J.; Langlais, F. Prediction of rotator cuff repair results by magnetic resonance imaging. Clin Orthop Relat Res. 1997, 275-283.
- Goutallier, D.; Postel, J. M.; Gleyze, P.; Leguilloux, P.; Van Driessche, S. Influence of cuff muscle fatty degeneration on anatomic and functional outcomes after simple suture of full-thickness tears. *J Shoulder Elbow* Surg. 2003, 12, 550-554.
- Galatz, L. M.; Silva, M. J.; Rothermich, S. Y.; Zaegel, M. A.; Havlioglu, N.; Thomopoulos, S. Nicotine delays tendon-to-bone healing in a rat shoulder model. *J Bone Joint Surg Am.* 2006, 88, 2027-2034.
- Bedi, A.; Fox, A. J.; Harris, P. E.; Deng, X. H.; Ying, L.; Warren, R.
 F.; Rodeo, S. A. Diabetes mellitus impairs tendon-bone healing after rotator cuff repair. *J Shoulder Elbow Surg.* 2010, *19*, 978-988.

- Chen, A. L.; Shapiro, J. A.; Ahn, A. K.; Zuckerman, J. D.; Cuomo, F. Rotator cuff repair in patients with type I diabetes mellitus. *J Shoulder Elbow Surg.* 2003, 12, 416-421.
- Tilley, B. J.; Cook, J. L.; Docking, S. I.; Gaida, J. E. Is higher serum cholesterol associated with altered tendon structure or tendon pain? A systematic review. *Br J Sports Med.* 2015, 49, 1504-1509.
- Bolam, S. M.; Park, Y. E.; Konar, S.; Callon, K. E.; Workman, J.; Monk, A. P.; Coleman, B.; Cornish, J.; Vickers, M. H.; Munro, J. T.; Musson, D. S. Obesity impairs enthesis healing after rotator cuff repair in a rat model. *Am J Sports Med.* 2021, *49*, 3959-3969.
- Angeline, M. E.; Ma, R.; Pascual-Garrido, C.; Voigt, C.; Deng, X. H.;
 Warren, R. F.; Rodeo, S. A. Effect of diet-induced vitamin D deficiency on rotator cuff healing in a rat model. *Am J Sports Med.* 2014, 42, 27-34.
- 60. Ryu, K. J.; Kim, B. H.; Lee, Y.; Dan, J.; Kim, J. H. Low serum vitamin d is not correlated with the severity of a rotator cuff tear or retear after arthroscopic repair. *Am J Sports Med.* **2015**, *43*, 1743-1750.
- Chung, S. W.; Oh, J. H.; Gong, H. S.; Kim, J. Y.; Kim, S. H. Factors affecting rotator cuff healing after arthroscopic repair: osteoporosis as one of the independent risk factors. *Am J Sports Med.* 2011, *39*, 2099-2107.
- Cohen, D. B.; Kawamura, S.; Ehteshami, J. R.; Rodeo, S. A. Indomethacin and celecoxib impair rotator cuff tendon-to-bone healing. *Am J Sports Med.* 2006, 34, 362-369.
- Chan, K. M.; Fu, S. C. Anti-inflammatory management for tendon injuries - friends or foes? Sports Med Arthrosc Rehabil Ther Technol. 2009, 1, 23.
- 64. Rak Kwon, D.; Jung, S.; Jang, J.; Park, G. Y.; Suk Moon, Y.; Lee, S. C. A 3-Dimensional bioprinted scaffold with human umbilical cord blood-mesenchymal stem cells improves regeneration of chronic full-thickness rotator cuff tear in a rabbit model. *Am J Sports Med.* 2020, 48, 947-958.
- García-Gómez, I.; Elvira, G.; Zapata, A. G.; Lamana, M. L.; Ramírez, M.; Castro, J. G.; Arranz, M. G.; Vicente, A.; Bueren, J.; García-Olmo, D. Mesenchymal stem cells: biological properties and clinical applications. *Expert Opin Biol Ther.* 2010, 10, 1453-1468.
- Lee, E. H.; Hui, J. H. The potential of stem cells in orthopaedic surgery. *J Bone Joint Surg Br.* 2006, 88, 841-851.
- 67. Morton-Gonzaba, N.; Carlisle, D.; Emukah, C.; Chorath, K.; Moreira, A. Mesenchymal stem cells and their application to rotator cuff pathology: A meta-analysis of pre-clinical studies. *Osteoarthr Cartil Open.* 2020, 2, 100047.
- Honda, H.; Gotoh, M.; Kanazawa, T.; Ohzono, H.; Nakamura, H.; Ohta, K.; Nakamura, K. I.; Fukuda, K.; Teramura, T.; Hashimoto, T.; Shichijo, S.; Shiba, N. Hyaluronic acid accelerates tendon-to-bone healing after rotator cuff repair. Am J Sports Med. 2017, 45, 3322-3330.
- Jo, C. H.; Shin, J. S.; Park, I. W.; Kim, H.; Lee, S. Y. Multiple channeling improves the structural integrity of rotator cuff repair. *Am J Sports Med*. 2013, 41, 2650-2657.
- Hernigou, P.; Flouzat Lachaniette, C. H.; Delambre, J.; Zilber, S.;
 Duffiet, P.; Chevallier, N.; Rouard, H. Biologic augmentation of rotator cuff repair with mesenchymal stem cells during arthroscopy improves healing and prevents further tears: a case-controlled study. *Int Orthop.* 2014, 38, 1811-1818.
- Taniguchi, N.; Suenaga, N.; Oizumi, N.; Miyoshi, N.; Yamaguchi, H.; Inoue, K.; Chosa, E. Bone marrow stimulation at the footprint of arthroscopic surface-holding repair advances cuff repair integrity. *J* Shoulder Elbow Surg. 2015, 24, 860-866.
- 72. Han, L.; Fang, W. L.; Jin, B.; Xu, S. C.; Zheng, X.; Hu, Y. G.

- Enhancement of tendon-bone healing after rotator cuff injuries using combined therapy with mesenchymal stem cells and platelet rich plasma. *Eur Rev Med Pharmacol Sci.* **2019**, *23*, 9075-9084.
- 73. Gulotta, L. V.; Kovacevic, D.; Ehteshami, J. R.; Dagher, E.; Packer, J. D.; Rodeo, S. A. Application of bone marrow-derived mesenchymal stem cells in a rotator cuff repair model. *Am J Sports Med.* **2009**, *37*, 2126-2133.
- 74. Nourissat, G.; Diop, A.; Maurel, N.; Salvat, C.; Dumont, S.; Pigenet, A.; Gosset, M.; Houard, X.; Berenbaum, F. Mesenchymal stem cell therapy regenerates the native bone-tendon junction after surgical repair in a degenerative rat model. *PLoS One.* **2010**, *5*, e12248.
- Gulotta, L. V.; Kovacevic, D.; Montgomery, S.; Ehteshami, J. R.; Packer, J. D.; Rodeo, S. A. Stem cells genetically modified with the developmental gene MT1-MMP improve regeneration of the supraspinatus tendon-to-bone insertion site. *Am J Sports Med.* 2010, 38, 1429-1437.
- 76. Vishnubalaji, R.; Al-Nbaheen, M.; Kadalmani, B.; Aldahmash, A.; Ramesh, T. Comparative investigation of the differentiation capability of bone-marrow- and adipose-derived mesenchymal stem cells by qualitative and quantitative analysis. *Cell Tissue Res.* 2012, 347, 419-427.
- Oh, J. H.; Chung, S. W.; Kim, S. H.; Chung, J. Y.; Kim, J. Y. 2013 Neer Award: Effect of the adipose-derived stem cell for the improvement of fatty degeneration and rotator cuff healing in rabbit model. *J Shoulder Elbow Surg.* 2014, 23, 445-455.
- Chen, Y.; Xu, Y.; Dai, G.; Shi, Q.; Duan, C. Enhanced Repaired Enthesis Using Tenogenically Differentiated Adipose-Derived Stem Cells in a Murine Rotator Cuff Injury Model. Stem Cells Int. 2022, 2022, 1309684.
- Choi, J. H.; Shim, I. K.; Shin, M. J.; Lee, Y. N.; Koh, K. H. Stem cell sheet interpositioned between the tendon and bone would be better for healing than stem cell sheet overlaid above the tendon-to-bone junction in rotator cuff repair of rats. *PLoS One.* 2022, 17, e0266030.
- Valencia Mora, M.; Antuña Antuña, S.; García Arranz, M.; Carrascal, M. T.; Barco, R. Application of adipose tissue-derived stem cells in a rat rotator cuff repair model. *Injury.* 2014, 45 Suppl 4, S22-27.
- 81. Rothrauff, B. B.; Smith, C. A.; Ferrer, G. A.; Novaretti, J. V.; Pauyo, T.; Chao, T.; Hirsch, D.; Beaudry, M. F.; Herbst, E.; Tuan, R. S.; Debski, R. E.; Musahl, V. The effect of adipose-derived stem cells on enthesis healing after repair of acute and chronic massive rotator cuff tears in rats. J Shoulder Elbow Surg. 2019, 28, 654-664.
- 82. Zong, J. C.; Mosca, M. J.; Degen, R. M.; Lebaschi, A.; Carballo, C.; Carbone, A.; Cong, G. T.; Ying, L.; Deng, X. H.; Rodeo, S. A. Involvement of Indian hedgehog signaling in mesenchymal stem cell-augmented rotator cuff tendon repair in an athymic rat model. J Shoulder Elbow Surg. 2017, 26, 580-588.
- Schwartz, A. G.; Galatz, L. M.; Thomopoulos, S. Enthesis regeneration: a role for Gli1+ progenitor cells. *Development.* 2017, 144, 1159-1164.
- 84. Nakazawa, T.; Nakajima, A.; Shiomi, K.; Moriya, H.; Einhorn, T. A.; Yamazaki, M. Effects of low-dose, intermittent treatment with recombinant human parathyroid hormone (1-34) on chondrogenesis in a model of experimental fracture healing. *Bone.* **2005**, *37*, 711-719.
- Kakar, S.; Einhorn, T. A.; Vora, S.; Miara, L. J.; Hon, G.; Wigner, N. A.; Toben, D.; Jacobsen, K. A.; Al-Sebaei, M. O.; Song, M.; Trackman, P. C.; Morgan, E. F.; Gerstenfeld, L. C.; Barnes, G. L. Enhanced chondrogenesis and Wnt signaling in PTH-treated fractures. *J Bone Miner Res.* 2007, 22, 1903-1912.
- Chen, Y.; Bai, B.; Zhang, S.; Ye, J.; Chen, Y.; Zeng, Y. Effects of parathyroid hormone on calcium ions in rat bone marrow mesenchymal stem cells. *Biomed Res Int.* 2014, 2014, 258409.
- 87. Hettrich, C. M.; Beamer, B. S.; Bedi, A.; Deland, K.; Deng, X. H.; Ying,

- L.; Lane, J.; Rodeo, S. A. The effect of rhPTH on the healing of tendon to bone in a rat model. *J Orthop Res.* **2012**, *30*, 769-774.
- 88. Duchman, K. R.; Goetz, J. E.; Uribe, B. U.; Amendola, A. M.; Barber, J. A.; Malandra, A. E.; Fredericks, D. C.; Hettrich, C. M. Delayed administration of recombinant human parathyroid hormone improves early biomechanical strength in a rat rotator cuff repair model. *J Shoulder Elbow Surg.* 2016, 25, 1280-1287.
- 89. Oh, J. H.; Kim, D. H.; Jeong, H. J.; Park, J. H.; Rhee, S. M. Effect of recombinant human parathyroid hormone on rotator cuff healing after arthroscopic repair. *Arthroscopy.* **2019**, *35*, 1064-1071.
- Gross, G.; Hoffmann, A. Therapeutic strategies for tendon healing based on novel biomaterials, factors and cells. *Pathobiology.* 2013, 80, 203-210.
- Caliari, S. R.; Harley, B. A. Composite growth factor supplementation strategies to enhance tenocyte bioactivity in aligned collagen-GAG scaffolds. Tissue Eng Part A. 2013, 19, 1100-1112.
- 92. Hee, C. K.; Dines, J. S.; Dines, D. M.; Roden, C. M.; Wisner-Lynch, L. A.; Turner, A. S.; McGilvray, K. C.; Lyons, A. S.; Puttlitz, C. M.; Santoni, B. G. Augmentation of a rotator cuff suture repair using rhPDGF-BB and a type I bovine collagen matrix in an ovine model. *Am J Sports Med.* 2011, 39, 1630-1639.
- 93. Novakova, S. S.; Mahalingam, V. D.; Florida, S. E.; Mendias, C. L.; Allen, A.; Arruda, E. M.; Bedi, A.; Larkin, L. M. Tissue-engineered tendon constructs for rotator cuff repair in sheep. *J Orthop Res.* **2018**, *36*, 289-299.
- Tokunaga, T.; Shukunami, C.; Okamoto, N.; Taniwaki, T.; Oka, K.; Sakamoto, H.; Ide, J.; Mizuta, H.; Hiraki, Y. FGF-2 stimulates the growth of tenogenic progenitor cells to facilitate the generation of tenomodulin-positive tenocytes in a rat rotator cuff healing model. *Am J Sports Med.* 2015, 43, 2411-2422.
- 95. Uggen, C.; Dines, J.; McGarry, M.; Grande, D.; Lee, T.; Limpisvasti, O. The effect of recombinant human platelet-derived growth factor BB-coated sutures on rotator cuff healing in a sheep model. *Arthroscopy*. 2010, 26, 1456-1462.
- Zhao, S.; Zhao, J.; Dong, S.; Huangfu, X.; Li, B.; Yang, H.; Zhao, J.;
 Cui, W. Biological augmentation of rotator cuff repair using bFGF-loaded electrospun poly(lactide-co-glycolide) fibrous membranes. *Int J Nanomedicine*. 2014, 9, 2373-2385.
- Sahoo, S.; Toh, S. L.; Goh, J. C. A bFGF-releasing silk/PLGA-based biohybrid scaffold for ligament/tendon tissue engineering using mesenchymal progenitor cells. *Biomaterials*. 2010, 31, 2990-2998.
- Ker, E. D.; Nain, A. S.; Weiss, L. E.; Wang, J.; Suhan, J.; Amon, C. H.; Campbell, P. G. Bioprinting of growth factors onto aligned sub-micron fibrous scaffolds for simultaneous control of cell differentiation and alignment. *Biomaterials*. 2011, 32, 8097-8107.
- James, R.; Kumbar, S. G.; Laurencin, C. T.; Balian, G.; Chhabra, A.
 B. Tendon tissue engineering: adipose-derived stem cell and GDF-5 mediated regeneration using electrospun matrix systems. *Biomed Mater*. 2011, 6, 025011.
- 100. Raabe, O.; Shell, K.; Fietz, D.; Freitag, C.; Ohrndorf, A.; Christ, H. J.; Wenisch, S.; Arnhold, S. Tenogenic differentiation of equine adiposetissue-derived stem cells under the influence of tensile strain, growth differentiation factors and various oxygen tensions. *Cell Tissue Res.* 2013, 352, 509-521.
- Holladay, C.; Abbah, S. A.; O'Dowd, C.; Pandit, A.; Zeugolis,
 D. I. Preferential tendon stem cell response to growth factor supplementation. J Tissue Eng Regen Med. 2016, 10, 783-798.
- 102. Durant, T. J.; Dyment, N.; McCarthy, M. B.; Cote, M. P.; Arciero, R. A.;

Tailored biological therapies in different rotator cuff tear

- Mazzocca, A. D.; Rowe, D. Mesenchymal stem cell response to growth factor treatment and low oxygen tension in 3-dimensional construct environment. *Muscles Ligaments Tendons J.* **2014**, *4*, 46-51.
- 103. Cheng, X.; Tsao, C.; Sylvia, V. L.; Cornet, D.; Nicolella, D. P.; Bredbenner, T. L.; Christy, R. J. Platelet-derived growth-factorreleasing aligned collagen-nanoparticle fibers promote the proliferation and tenogenic differentiation of adipose-derived stem cells. *Acta Biomater.* 2014, 10, 1360-1369.
- 104. Manning, C. N.; Schwartz, A. G.; Liu, W.; Xie, J.; Havlioglu, N.; Sakiyama-Elbert, S. E.; Silva, M. J.; Xia, Y.; Gelberman, R. H.; Thomopoulos, S. Controlled delivery of mesenchymal stem cells and growth factors using a nanofiber scaffold for tendon repair. *Acta Biomater.* 2013, *9*, 6905-6914.
- 105. Würgler-Hauri, C. C.; Dourte, L. M.; Baradet, T. C.; Williams, G. R.; Soslowsky, L. J. Temporal expression of 8 growth factors in tendon-tobone healing in a rat supraspinatus model. *J Shoulder Elbow Surg.* 2007, 16, S198-203.
- 106. Kobayashi, M.; Itoi, E.; Minagawa, H.; Miyakoshi, N.; Takahashi, S.; Tuoheti, Y.; Okada, K.; Shimada, Y. Expression of growth factors in the early phase of supraspinatus tendon healing in rabbits. *J Shoulder Elbow Surg.* **2006**, *15*, 371-377.
- 107. Rodeo, S. A.; Potter, H. G.; Kawamura, S.; Turner, A. S.; Kim, H. J.; Atkinson, B. L. Biologic augmentation of rotator cuff tendon-healing with use of a mixture of osteoinductive growth factors. *J Bone Joint Surg Am.* 2007, 89, 2485-2497.
- 108. Kim, H. M.; Galatz, L. M.; Das, R.; Havlioglu, N.; Rothermich, S. Y.; Thomopoulos, S. The role of transforming growth factor beta isoforms in tendon-to-bone healing. *Connect Tissue Res.* **2011**, *52*, 87-98.
- 109. Kovacevic, D.; Fox, A. J.; Bedi, A.; Ying, L.; Deng, X. H.; Warren, R. F.; Rodeo, S. A. Calcium-phosphate matrix with or without TGF-β3 improves tendon-bone healing after rotator cuff repair. *Am J Sports Med.* 2011, 39, 811-819.
- 110. Manning, C. N.; Kim, H. M.; Sakiyama-Elbert, S.; Galatz, L. M.; Havlioglu, N.; Thomopoulos, S. Sustained delivery of transforming growth factor beta three enhances tendon-to-bone healing in a rat model. *J Orthop Res.* 2011, 29, 1099-1105.
- 111. Nagura, I.; Kokubu, T.; Mifune, Y.; Inui, A.; Takase, F.; Ueda, Y.; Kataoka, T.; Kurosaka, M. Characterization of progenitor cells derived from torn human rotator cuff tendons by gene expression patterns of chondrogenesis, osteogenesis, and adipogenesis. J Orthop Surg Res. 2016, 11, 40.
- 112. Davies, M. R.; Liu, X.; Lee, L.; Laron, D.; Ning, A. Y.; Kim, H. T.; Feeley, B. T. TGF-β small molecule inhibitor SB431542 reduces rotator cuff muscle fibrosis and fatty infiltration by promoting fibro/adipogenic progenitor apoptosis. *PLoS One.* 2016, 11, e0155486.
- 113. Zhou, Z.; Song, W.; Zhang, G.; Zhan, S.; Cai, Z.; Yu, W.; He, Y. The recombinant human fibroblast growth factor-18 (sprifermin) improves tendon-to-bone healing by promoting chondrogenesis in a rat rotator cuff repair model. *J Shoulder Elbow Surg.* **2022**, *31*, 1617-1627.
- 114. Sitcheran, R.; Cogswell, P. C.; Baldwin, A. S., Jr. NF-kappaB mediates inhibition of mesenchymal cell differentiation through a posttranscriptional gene silencing mechanism. *Genes Dev.* 2003, 17, 2368-2373.
- 115. Gulotta, L. V.; Kovacevic, D.; Cordasco, F.; Rodeo, S. A. Evaluation of tumor necrosis factor *α* blockade on early tendon-to-bone healing in a rat rotator cuff repair model. *Arthroscopy.* **2011**, *27*, 1351-1357.
- 116. Waldorff, E. I.; Lindner, J.; Kijek, T. G.; Downie, B. K.; Hughes, R. E.; Carpenter, J. E.; Miller, B. S. Bone density of the greater tuberosity is

- decreased in rotator cuff disease with and without full-thickness tears. *J Shoulder Elbow Surg.* **2011**, *20*, 904-908.
- 117. Dorman, L. J.; Tucci, M.; Benghuzzi, H. In vitro effects of bmp-2, bmp-7, and bmp-13 on proliferation and differentation of mouse mesenchymal stem cells. *Biomed Sci Instrum.* 2012, 48, 81-87.
- 118. Kabuto, Y.; Morihara, T.; Sukenari, T.; Kida, Y.; Oda, R.; Arai, Y.; Sawada, K.; Matsuda, K.; Kawata, M.; Tabata, Y.; Fujiwara, H.; Kubo, T. Stimulation of rotator cuff repair by sustained release of bone morphogenetic protein-7 using a gelatin hydrogel sheet. *Tissue Eng Part A.* 2015, 21, 2025-2033.
- 119. Dahlgren, L. A.; Mohammed, H. O.; Nixon, A. J. Temporal expression of growth factors and matrix molecules in healing tendon lesions. *J Orthop Res.* **2005**, *23*, 84-92.
- 120. Carr, J. B., 2nd; Rodeo, S. A. The role of biologic agents in the management of common shoulder pathologies: current state and future directions. *J Shoulder Elbow Surg.* **2019**, *28*, 2041-2052.
- 121. Neviaser, J. S.; Neviaser, R. J.; Neviaser, T. J. The repair of chronic massive ruptures of the rotator cuff of the shoulder by use of a freezedried rotator cuff. *J Bone Joint Surg Am.* **1978**, *60*, 681-684.
- 122. Dejardin, L. M.; Arnoczky, S. P.; Ewers, B. J.; Haut, R. C.; Clarke, R. B. Tissue-engineered rotator cuff tendon using porcine small intestine submucosa. Histologic and mechanical evaluation in dogs. *Am J Sports Med.* 2001, *29*, 175-184.
- 123. Zheng, M. H.; Chen, J.; Kirilak, Y.; Willers, C.; Xu, J.; Wood, D. Porcine small intestine submucosa (SIS) is not an acellular collagenous matrix and contains porcine DNA: possible implications in human implantation. *J Biomed Mater Res B Appl Biomater.* **2005**, *73*, 61-67.
- 124. Huang, C.; Zhang, X.; Luo, H.; Pan, J.; Cui, W.; Cheng, B.; Zhao, S.; Chen, G. Effect of kartogenin-loaded gelatin methacryloyl hydrogel scaffold with bone marrow stimulation for enthesis healing in rotator cuff repair. J Shoulder Elbow Surg. 2021, 30, 544-553.
- 125. Han, L.; Liu, H.; Fu, H.; Hu, Y.; Fang, W.; Liu, J. Exosome-delivered BMP-2 and polyaspartic acid promotes tendon bone healing in rotator cuff tear via Smad/RUNX2 signaling pathway. *Bioengineered.* 2022, 13, 1459-1475.
- 126. Ousema, P. H.; Moutos, F. T.; Estes, B. T.; Caplan, A. I.; Lennon, D. P.; Guilak, F.; Weinberg, J. B. The inhibition by interleukin 1 of MSC chondrogenesis and the development of biomechanical properties in biomimetic 3D woven PCL scaffolds. *Biomaterials*. 2012, 33, 8967-8974.
- 127. Jiang, X.; Wu, S.; Kuss, M.; Kong, Y.; Shi, W.; Streubel, P. N.; Li, T.; Duan, B. 3D printing of multilayered scaffolds for rotator cuff tendon regeneration. *Bioact Mater.* **2020**, *5*, 636-643.
- 128. Ciampi, P.; Scotti, C.; Nonis, A.; Vitali, M.; Di Serio, C.; Peretti, G. M.; Fraschini, G. The benefit of synthetic versus biological patch augmentation in the repair of posterosuperior massive rotator cuff tears: a 3-year follow-up study. Am J Sports Med. 2014, 42, 1169-1175.
- 129. Cai, Y. Z.; Zhang, C.; Jin, R. L.; Shen, T.; Gu, P. C.; Lin, X. J.; Chen, J. D. Arthroscopic rotator cuff repair with graft augmentation of 3-dimensional biological collagen for moderate to large tears: a randomized controlled study. Am J Sports Med. 2018, 46, 1424-1431.
- 130. Sclamberg, S. G.; Tibone, J. E.; Itamura, J. M.; Kasraeian, S. Six-month magnetic resonance imaging follow-up of large and massive rotator cuff repairs reinforced with porcine small intestinal submucosa. *J Shoulder Elbow Surg.* 2004, 13, 538-541.
- 131. Iannotti, J. P.; Codsi, M. J.; Kwon, Y. W.; Derwin, K.; Ciccone, J.; Brems, J. J. Porcine small intestine submucosa augmentation of surgical repair of chronic two-tendon rotator cuff tears. A randomized, controlled trial. *J Bone Joint Surg Am.* **2006**, *88*, 1238-1244.

132. Malcarney, H. L.; Bonar, F.; Murrell, G. A. Early inflammatory reaction after rotator cuff repair with a porcine small intestine submucosal implant: a report of 4 cases. *Am J Sports Med.* **2005**, *33*, 907-911.

- 133. Hoberman, A. R.; Cirino, C.; McCarthy, M. B.; Cote, M. P.; Pauzenberger, L.; Beitzel, K.; Mazzocca, A. D.; Dyrna, F. Bone marrow-derived mesenchymal stromal cells enhanced by platelet-rich plasma maintain adhesion to scaffolds in arthroscopic simulation. *Arthroscopy*. 2018, 34, 872-881.
- 134. Thangarajah, T.; Henshaw, F.; Sanghani-Kerai, A.; Lambert, S. M.; Blunn, G. W.; Pendegrass, C. J. The effectiveness of demineralized cortical bone matrix in a chronic rotator cuff tear model. J Shoulder Elbow Surg. 2017, 26, 619-626.
- 135. Thangarajah, T.; Sanghani-Kerai, A.; Henshaw, F.; Lambert, S. M.; Pendegrass, C. J.; Blunn, G. W. Application of a demineralized cortical bone matrix and bone marrow-derived mesenchymal stem cells in a model of chronic rotator cuff degeneration. *Am J Sports Med.* 2018, 46, 98-108
- 136. Smith, M. J.; Pfeiffer, F. M.; Cook, C. R.; Kuroki, K.; Cook, J. L. Rotator cuff healing using demineralized cancellous bone matrix sponge interposition compared to standard repair in a preclinical canine model. *J Orthop Res.* 2018, 36, 906-912.
- 137. Wellington, I. J.; Muench, L. N.; Hawthorne, B. C.; Uyeki, C. L.; Antonacci, C. L.; McCarthy, M. B.; Connors, J. P.; Kia, C.; Mazzocca, A. D.; Berthold, D. P. Clinical outcomes following biologically enhanced demineralized bone matrix augmentation of complex rotator cuff repair. J Clin Med. 2022, 11, 2956.
- 138. Chae, S.; Yong, U.; Park, W.; Choi, Y. M.; Jeon, I. H.; Kang, H.; Jang, J.; Choi, H. S.; Cho, D. W. 3D cell-printing of gradient multi-tissue interfaces for rotator cuff regeneration. *Bioact Mater.* **2023**, *19*, 611-625.
- 139. Yoon, J. Y.; Park, J. H.; Rhee, S. M.; Jeong, H. J.; Han, J.; Lee, J. H.; Jeon, S.; Oh, J. H. Safety and efficacy of autologous dermal fibroblast injection to enhance healing after full-thickness rotator cuff repair: first-in-human pilot study. *Orthop J Sports Med.* 2021, *9*, 23259671211052996.
- 140. Randelli, P.; Randelli, F.; Ragone, V.; Menon, A.; D'Ambrosi, R.; Cucchi, D.; Cabitza, P.; Banfi, G. Regenerative medicine in rotator cuff injuries. *Biomed Res Int.* 2014, 2014, 129515.
- 141. Warth, R. J.; Dornan, G. J.; James, E. W.; Horan, M. P.; Millett, P. J. Clinical and structural outcomes after arthroscopic repair of full-thickness rotator cuff tears with and without platelet-rich product supplementation: a meta-analysis and meta-regression. *Arthroscopy.* **2015**, *31*, 306-320.
- 142. Castricini, R.; Longo, U. G.; De Benedetto, M.; Panfoli, N.; Pirani, P.; Zini, R.; Maffulli, N.; Denaro, V. Platelet-rich plasma augmentation for arthroscopic rotator cuff repair: a randomized controlled trial. Am J Sports Med. 2011, 39, 258-265.
- 143. Mazzocca, A. D.; McCarthy, M. B.; Chowaniec, D. M.; Cote, M. P.; Romeo, A. A.; Bradley, J. P.; Arciero, R. A.; Beitzel, K. Platelet-rich plasma differs according to preparation method and human variability. J Bone Joint Surg Am. 2012, 94, 308-316.
- 144. Giovannetti de Sanctis, E.; Franceschetti, E.; De Dona, F.; Palumbo, A.; Paciotti, M.; Franceschi, F. The efficacy of injections for partial rotator cuff tears: a systematic review. *J Clin Med.* **2020**, *10*, 51.
- 145. von Wehren, L.; Blanke, F.; Todorov, A.; Heisterbach, P.; Sailer, J.; Majewski, M. The effect of subacromial injections of autologous conditioned plasma versus cortisone for the treatment of symptomatic partial rotator cuff tears. Knee Surg Sports Traumatol Arthrosc. 2016, 24, 3787-3792.
- 146. Xu, W.; Xue, Q. Application of platelet-rich plasma in arthroscopic

- rotator cuff repair: a systematic review and meta-analysis. *Orthop J Sports Med.* **2021**, *9*, 23259671211016847.
- 147. Rha, D. W.; Park, G. Y.; Kim, Y. K.; Kim, M. T.; Lee, S. C. Comparison of the therapeutic effects of ultrasound-guided platelet-rich plasma injection and dry needling in rotator cuff disease: a randomized controlled trial. *Clin Rehabil.* **2013**, *27*, 113-122.
- 148. Shams, A.; El-Sayed, M.; Gamal, O.; Ewes, W. Subacromial injection of autologous platelet-rich plasma versus corticosteroid for the treatment of symptomatic partial rotator cuff tears. Eur J Orthop Surg Traumatol. 2016, 26, 837-842.
- 149. Cai, Y. U.; Sun, Z.; Liao, B.; Song, Z.; Xiao, T.; Zhu, P. Sodium hyaluronate and platelet-rich plasma for partial-thickness rotator cuff tears. *Med Sci Sports Exerc.* **2019**, *51*, 227-233.
- 150. Ryan, J.; Imbergamo, C.; Sudah, S.; Kirchner, G.; Greenberg, P.; Monica, J.; Gatt, C. Platelet-rich product supplementation in rotator cuff repair reduces retear rates and improves clinical outcomes: a meta-analysis of randomized controlled trials. *Arthroscopy.* 2021, 37, 2608-2624.
- 151. Lavoie-Gagne, O.; Fury, M. S.; Mehta, N.; Harkin, W. E.; Bernstein, D. N.; Berlinberg, E. J.; Parvaresh, K.; O'Donnell, E.; Forsythe, B. Doublerow repair with platelet-rich plasma optimizes retear rates after small to medium full-thickness rotator cuff repair: a systematic review and network meta-analysis of randomized controlled trials. *Arthroscopy*. 2022, 38, 2714-2729.
- Ilhanli, I.; Guder, N.; Gul, M. Platelet-rich plasma treatment with physical therapy in chronic partial supraspinatus tears. *Iran Red Crescent Med J.* 2015, 17, e23732.
- 153. Anz, A. W.; Parsa, R. S.; Romero-Creel, M. F.; Nabors, A.; Tucker, M. S.; Harrison, R. M.; Matuska, A. M. Exercise-mobilized platelet-rich plasma: short-term exercise increases stem cell and platelet concentrations in platelet-rich plasma. *Arthroscopy.* 2019, 35, 192-200.
- Osborne, J. D.; Gowda, A. L.; Wiater, B.; Wiater, J. M. Rotator cuff rehabilitation: current theories and practice. *Phys Sportsmed.* 2016, 44, 85-92.
- 155. Xiao, H.; Zhang, T.; Li, C.; Cao, Y.; Wang, L.; Chen, H.; Li, S.; Guan, C.; Hu, J.; Chen, D.; Chen, C.; Lu, H. Mechanical stimulation promotes enthesis injury repair by mobilizing Prrx1(+) cells via ciliary TGF-β signaling. Elife. 2022, 11, e73614.
- 156. Galatz, L. M.; Charlton, N.; Das, R.; Kim, H. M.; Havlioglu, N.; Thomopoulos, S. Complete removal of load is detrimental to rotator cuff healing. J Shoulder Elbow Surg. 2009, 18, 669-675.
- 157. Gimbel, J. A.; Van Kleunen, J. P.; Williams, G. R.; Thomopoulos, S.; Soslowsky, L. J. Long durations of immobilization in the rat result in enhanced mechanical properties of the healing supraspinatus tendon insertion site. *J Biomech Eng.* 2007, 129, 400-404.
- 158. Hettrich, C. M.; Gasinu, S.; Beamer, B. S.; Stasiak, M.; Fox, A.; Birmingham, P.; Ying, O.; Deng, X. H.; Rodeo, S. A. The effect of mechanical load on tendon-to-bone healing in a rat model. *Am J Sports Med.* 2014, 42, 1233-1241.
- 159. Chen, H.; Li, S.; Xiao, H.; Wu, B.; Zhou, L.; Hu, J.; Lu, H. Effect of exercise intensity on the healing of the bone-tendon interface: a mouse rotator cuff injury model study. *Am J Sports Med.* **2021**, *49*, 2064-2073.
- 160. Teunis, T.; Lubberts, B.; Reilly, B. T.; Ring, D. A systematic review and pooled analysis of the prevalence of rotator cuff disease with increasing age. J Shoulder Elbow Surg. 2014, 23, 1913-1921.
- 161. Kaplan, L. D.; Flanigan, D. C.; Norwig, J.; Jost, P.; Bradley, J. Prevalence and variance of shoulder injuries in elite collegiate football players. *Am J Sports Med.* **2005**, *33*, 1142-1146.

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- 162. Mall, N. A.; Lee, A. S.; Chahal, J.; Sherman, S. L.; Romeo, A. A.; Verma, N. N.; Cole, B. J. An evidenced-based examination of the epidemiology and outcomes of traumatic rotator cuff tears. *Arthroscopy.* 2013, 29, 366-376.
- 163. Altintas, B.; Anderson, N.; Dornan, G. J.; Boykin, R. E.; Logan, C.; Millett, P. J. Return to sport after arthroscopic rotator cuff repair: is there a difference between the recreational and the competitive athlete? Am J Sports Med. 2020, 48, 252-261.
- 164. Burns, J. P.; Snyder, S. J. Arthroscopic rotator cuff repair in patients younger than fifty years of age. J Shoulder Elbow Surg. 2008, 17, 90-96.
- 165. Antoni, M.; Klouche, S.; Mas, V.; Ferrand, M.; Bauer, T.; Hardy, P. Return to recreational sport and clinical outcomes with at least 2 years follow-up after arthroscopic repair of rotator cuff tears. *Orthop Traumatol Surg Res.* 2016, 102, 563-567.
- Cao, Y. S.; Wan, Y. F. Progress on tendon-to-bone healing after rotator cuff repair. Zhongguo Gu Shang. 2018, 31, 1172-1179.
- 167. Font Tellado, S.; Balmayor, E. R.; Van Griensven, M. Strategies to engineer tendon/ligament-to-bone interface: Biomaterials, cells and growth factors. *Adv Drug Deliv Rev.* **2015**, *94*, 126-140.
- 168. Prabhath, A.; Vernekar, V. N.; Sanchez, E.; Laurencin, C. T. Growth factor delivery strategies for rotator cuff repair and regeneration. *Int J Pharm.* **2018**, *544*, 358-371.
- 169. Chowdhury, F.; Na, S.; Li, D.; Poh, Y. C.; Tanaka, T. S.; Wang, F.; Wang, N. Material properties of the cell dictate stress-induced spreading and differentiation in embryonic stem cells. *Nat Mater.* 2010, 9, 82-88.
- 170. McMurray, R. J.; Gadegaard, N.; Tsimbouri, P. M.; Burgess, K. V.; McNamara, L. E.; Tare, R.; Murawski, K.; Kingham, E.; Oreffo, R. O.; Dalby, M. J. Nanoscale surfaces for the long-term maintenance of mesenchymal stem cell phenotype and multipotency. *Nat Mater.* 2011, 10, 637-644.

- 171. Dalby, M. J.; Gadegaard, N.; Tare, R.; Andar, A.; Riehle, M. O.; Herzyk, P.; Wilkinson, C. D.; Oreffo, R. O. The control of human mesenchymal cell differentiation using nanoscale symmetry and disorder. *Nat Mater.* **2007**, *6*, 997-1003.
- 172. Gentleman, E.; Swain, R. J.; Evans, N. D.; Boonrungsiman, S.; Jell, G.; Ball, M. D.; Shean, T. A.; Oyen, M. L.; Porter, A.; Stevens, M. M. Comparative materials differences revealed in engineered bone as a function of cell-specific differentiation. *Nat Mater.* 2009, 8, 763-770.
- 173. Derwin, K. A.; Baker, A. R.; Iannotti, J. P.; McCarron, J. A. Preclinical models for translating regenerative medicine therapies for rotator cuff repair. *Tissue Eng Part B Rev.* **2010**, *16*, 21-30.
- 174. Ficklscherer, A.; Serr, M.; Loitsch, T.; Niethammer, T. R.; Lahner, M.; Pietschmann, M. F.; Müller, P. E. The influence of different footprint preparation techniques on tissue regeneration in rotator cuff repair in an animal model. *Arch Med Sci.* **2017**, *13*, 481-488.
- 175. Li, X.; Shen, P.; Su, W.; Zhao, S.; Zhao, J. Into-tunnel repair versus onto-surface repair for rotator cuff tears in a rabbit model. *Am J Sports Med.* **2018**, *46*, 1711-1719.
- 176. Bilsel, K.; Yildiz, F.; Kapicioglu, M.; Uzer, G.; Elmadag, M.; Pulatkan, A.; Esrefoglu, M.; Bozdag, E.; Milano, G. Efficacy of bone marrow-stimulating technique in rotator cuff repair. *J Shoulder Elbow Surg.* 2017, 26, 1360-1366.

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