

REVIEW

Management of neonatal central nervous system viral infections: Knowledge gaps and research priorities

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Abstract

Congenital CMV, enteroviruses, human parechovirus and herpes simplex virus are all common causes of severe central nervous system (CNS) infection in neonates. The introduction of screening (i.e. newborn hearing screening programme), integration of molecular syndromic testing (i.e. multiplex polymerase chain reaction assays) and increase in sexually transmitted infections (i.e. anogenital herpes) have contributed to increases in each of these infections over the last decade. However, therapeutic options are highly limited in part due to the lack of epidemiological data informing trials. This review will describe our current understanding of the clinical burden and epidemiology of these severe neonatal CNS infections, outline the novel antiviral and vaccines in the pipeline and suggest future research studies which could help develop new therapeutics.

KEYWORDS

antiviral, CMV, enterovirus, herpes simplex virus, parechovirus, vaccine

1 | INTRODUCTION

Viral infections affecting the central nervous system (CNS), namely congenital cytomegalovirus (cCMV), enteroviruses (EV), human parechovirus (HPeV) and herpes simplex viruses (HSV) cause significant morbidity and occasional mortality in neonates. In England, the introduction of the NHSP (newborn hearing screening programme) in 2007 led to a 55% increase in detection of neonates with cCMV.¹ Increased use of sensitive molecular tools, such as multiplex PCR (polymerase chain reaction) in neonates with suspected meningitis has led to a sevenfold increase in the detected incidence of all causes of viral meningitis.² Furthermore, there has been an increasing burden of disease; over the last decade, surveillance studies in the USA, U.K and Australia have all shown a year-on-year increase in maternal anogenital HSV and subsequent increase in neonatal HSV

disease.^{1,3,4} However, therapeutic interventions to treat and prevent neonatal viral infections are highly limited due to a paucity of epidemiological data to inform therapeutic trials.

This review will provide an overview of the current epidemiology, available treatment modalities and novel agents being trialled in neonates with cCMV, EV, HPeV and HSV. We highlight specific studies that could be conducted to inform the therapeutic pipeline for these poorly understood infections.

2 | METHODS

Literature databases PubMed and [ClinicalTrials.gov](https://clinicaltrials.gov) were searched using terms ['neonatal', 'neonate' or 'paediatric'] alongside the following interchangeable terms: ['cytomegalovirus' or 'cCMV'] or

Abbreviations: cCMV, congenital cytomegalovirus; CNS, central nervous system; CSF, cerebrospinal fluid; ECMO, extracorporeal membrane oxygenation; EV, enterovirus; HLH, hemophagocytic lymphohistiocytosis; HPeV, human parechovirus; HSCT, haematopoietic stem cell transplantation; HSV, herpes simplex virus; IVIG, intravenous immune globulin; LMIC, low middle income countries; NHSP, newborn hearing screening programme; PCR, polymerase chain reaction; RCT, randomised controlled trial.

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['enterovirus' or 'enteroviral' or 'coxsackie'] or ['parechovirus' or 'hPeV'] or ['HSV' or 'herpes']. Relevant clinical trials, case reports/series, trials and systematic/narrative reviews were identified, with references in each also analysed for relevance.

2.1 | Congenital cytomegalovirus

2.1.1 | Current epidemiology

In the adult population, cytomegalovirus (CMV) is a highly prevalent human herpes viral infection, with worldwide seroprevalence varying from 45% to 100%.⁵ Although usually presenting with non-specific mild symptoms of malaise, headache, fever and potential transaminitis in immunocompetent individuals, it can present with significant morbidity in those who are immunosuppressed or in CMV-naïve transplant recipients.

Maternal CMV infection is acquired due to exposure to individuals shedding CMV in bodily fluids. A majority of mothers have been exposed to CMV in early childhood or adolescence. However, sources of primary exposure are via toddlers through saliva (i.e. sharing food/utensils) or urine (i.e. diaper changes). Primary maternal infection affects 1%–4% of pregnancies and rates of transplacental/intrapartum transmission from seronegative and seropositive women to foetus are 32% and 1.4% respectively.⁶ Congenital infection can be a result of both primary infection or infection in mothers with pre-conceptual immunity.⁷ Infection most commonly occurs via vertical transmission, through transplacental translocation.

cCMV is symptomatic (i.e. causes overt disease) in 12% of infected neonates worldwide and is the most common cause of non-genetic sensorineural hearing loss (SNHL).⁸ Of neonates with symptomatic disease, 35% present with SNHL.⁹ cCMV may cause neurodevelopmental impairment (including cerebral palsy, epilepsy, autistic spectrum disorder and feed intolerance) and affected infants require a range of educational, medical and psychological support. In the UK, the annual direct (incurred by the public sector) and indirect (societal) costs of managing affected infants is estimated to be £732 million.¹⁰

Maternal CMV seropositivity is significantly higher in lower/middle income countries (LMIC) (Iran, Benin and India: 95%–97%) than high income countries (Ireland, Canada and Germany: 44%–59%).^{11,12} cCMV affects 0.3%–0.7% live births in developed countries.¹³ In LMIC settings, women are likely to be seropositive due to increased rates of transmission in denser populations, higher number of children per household and reduced access to sanitation. The incidence of cCMV in LMICs is poorly understood due to lack of routine data, population-based screening studies and inconsistent clinical definitions/testing. However, small single centre/regional studies estimate the incidence at 1.2% in rural North India,¹⁴ 5.8% in Jakarta¹⁵ and 2.6% in Mozambique,¹⁶ correlating with the higher transmission rates in LMIC settings.

The global burden of cCMV occurs most commonly in seropositive pregnant women. However, research trials, public health

messaging and clinical concern often focus on hygiene strategies to prevent infection in seronegative women (i.e. careful hand hygiene after diaper changes, avoiding kissing toddlers on the lips, not sharing food/utensils) which are the mainstay in preventing acquisition. These measures reduced primary infection from 7.6% to 1.2% in a study of 646 CMV seronegative pregnant women.¹⁷ This focus should broaden through ensuring enrolment of seropositive women into research studies in order to optimise understanding of how to minimise the risk of re-activation/re-infection and prevent transmission to the foetus.

2.1.2 | Current treatments

Educational interventions in pregnant women to prevent acquisition during the antenatal period are limited and inconsistently delivered. A double-blind placebo-controlled randomised controlled trial (RCT) of 45 pregnant women receiving valaciclovir reduced the rate of vertical transmission in early pregnancy (11% vs. 30%). The primary endpoint in this study was evidence of infection in amniotic fluid and data additionally analysed any evidence of symptomatic neonatal infection (pregnancies not carried to term due to non-CMV reasons were excluded) and demonstrated 7% (3/44) incidence in neonates/infants in the valaciclovir arm versus 16% (7/43) in the placebo group.¹⁸ CMV-specific Human Immunoglobulin has not been shown to prevent cCMV in an RCT of 123 pregnant women with primary infection.¹⁹

Only two RCTs have evaluated the safety and efficacy of antiviral treatment in neonates (Table 1). Each trial took a decade to complete and in combination recruited fewer than 200 neonates, with limited follow-up. Kimberlin et al. demonstrated that 6 weeks of IV ganciclovir (6 mg/kg administered twice daily) in neonates younger than 30 days with CNS disease resulted in improved audiological outcomes at 12-month follow up. Almost two thirds of those receiving ganciclovir had neutrophil counts <0.5.²⁰ A second RCT (2015), commencing 6 months versus 6 weeks of oral valganciclovir in symptomatic infants in the first month of life, resulted in modest audiological improvement and better expressive and receptive language at 2 years follow-up.⁹ However, a fifth of babies receiving valganciclovir developed significant neutropenia.

A phase II, open-label trial to evaluate valganciclovir (16 mg/kg/dose BD for 4 months), as prevention for SNHL in 229 asymptomatic neonates (non-randomised, single group assignment), is expected to complete in 2024.³⁶ No other phase II/III studies have evaluated the efficacy of treating asymptomatic neonates. Pharmacokinetic data of 24 neonates a dose of 16 mg/kg achieved area under the curve over 12 h target of 27 mg × h/L,²³ but there are a lack of pharmacokinetic/pharmacodynamic data to inform dosing in preterm neonates. This is particularly important as cCMV is associated with preterm delivery. The national screening committee in the UK cited that the absence of an approved highly efficacious and well-tolerated antiviral treatment is a significant obstacle to population-based screening.³⁷ No countries have implemented an antenatal or postnatal universal screening programme.

TABLE 1 Current widely used treatments for severe neonatal CNS infections

Current treatments	Toxicity	Efficacy	PK/PD data	Duration of treatment
CMV IV ganciclovir	29 of 46 ganciclovir treated neonates (63%) developed neutropenia <0.5 ²⁰	Reduction in sensorineural hearing loss (16% [4/25] worsening vs. 41% [7/17] in controls at 6 months; 21% [5/24] vs. 68% [13/19] at 12 months). Improved neurodevelopmental outcomes in study of 100 neonates (Denver developmental test scores 4.46 vs. 7.51 in controls at 6 months; 10.06 vs. 17.14 at 12 months) ^{20,21}	Analysis of neonatal samples post infusion demonstrated clearance (volume of fluid completely cleared of drug of) 0.428 L/h ²²	6 weeks ^{20,21}
PO valganciclovir	21% (10/47)—neutropenia <0.5. Slight non-significant increases in ALT and AST in treatment group ⁹	Improved total ear hearing outcomes at 12 months (73% [58/79] vs. 57% [44/77] improved or normal in the 6 months vs. 6 weeks treatment) and 24 months (77% [54/70] vs. 64% [37/58]), and improved Bayley Scale for neurodevelopmental scoring at 24 months ⁹	16 mg/kg PO valganciclovir equivalent plasma concentration to IV ganciclovir ²³	6 months
EV Limited evidence: use of IVIG	IVIG administration in neonates is largely safe. Rare side effects include thrombosis (1%–18%), anaphylaxis (theoretical risk), apnoea (case reports), arrhythmias (case reports) and necrotising enterocolitis (odds ratio 4.53 in haemolytic patients compared to controls) ²⁴	IVIG with neutralisation titre >1:800 was associated with higher neutralisation titres and more rapid reduction viraemia/viruria ²⁵	-	IVIG within first 72 h associated with favourable prognosis (odds ratio 14.7) ²⁶
HPeV Limited evidence: use of IVIG	As above	In vitro & case reports IVIG has been shown to provide potent in vitro antiviral activity, ²⁷ and indeed was used in the treatment of a patient with dilated cardiomyopathy. ²⁸ A cohort study of 108 febrile neonates <60 days of which 4 were RT-PCR positive for HPeV showed for all 4 who received IVIG starting at day 1–4 demonstrated improvements in clinical presentation of rash, diarrhoea, apnoea and seizures (data on durations for individual HPeV-infected cohort was not included) ²⁹	-	Limited data

(Continues)

TABLE 1 (Continued)

	Current treatments	Toxicity	Efficacy	PK/PD data	Duration of treatment
HSV	IV high-dose aciclovir (60 mg/kg/day split into TDS dosing)	6 (21%) of 29 neonates receiving high-dose aciclovir experienced transient neutropenia, which all spontaneously improved. ³⁰ Renal toxicity (5.6% of 1017 neonates) ³¹	Use of standard-dose aciclovir reduced 1-year mortality from disseminated disease to 61% from 85% with no therapy; and from CNS disease to 14% from 35%. ³² Higher dosing versus standard dosing for CNS and disseminated disease: odd ratio 3.3 for survival and 6.6 for normal neurodevelopmental outcome at 12 months ³⁰	Elimination half-life is 3 h for neonates 36–41 weeks, 6.55 h for neonates 30–35 weeks and 10.2 h for <30 weeks. ³³ Limited data for CNS pharmacokinetics in neonates—in adults, valaciclovir 1000 mg administration peaked at 2 days and was minimal at 20 days	14 days for skin/eyes/mouth, 21 days for disseminated disease and CNS depending on nature of disease dissemination; also require 6 months of PO aciclovir as suppressive treatment
	Followed by PO aciclovir	Transient, self-resolving neutropenia in infants receiving aciclovir suppressive therapy ³⁴	Neonates with CNS involvement had significantly higher neurodevelopmental outcome scores at 12 months compared to placebo ³⁴	Half-life decreased during first 30 days of life from 10 to 15 h to 2.5 h ³⁵	6 months

Abbreviations: CMV, cytomegalovirus; CNS, central nervous system; EV, enterovirus; HPeV, human parechovirus; HSV, herpes simplex virus; IVIG, intravenous immune globulin.

2.1.3 | Novel treatments

Other viral agents in trials to treat CMV infection include nucleoside analogues and DNA polymerase inhibitors, with paediatric studies largely limited to cohort studies.

Cidofovir, a nucleoside analogue, has been used to treat CMV retinitis.³⁸ However a cohort study of 5 neonates with adenoviral infection demonstrated renal toxicity in 80%, requiring continuous renal replacement therapy or ECMO.³⁹ Brincidofovir (lipid conjugate form of cidofovir) has been evaluated in three Phase II/III studies of haematopoietic stem cell or solid organ recipients in adults and children (230, 48 and 232 participants respectively) with CMV, HSV, adenovirus, vaccinia and varicella infections. This demonstrated 80% of participants had improved renal function when switched from either cidofovir or foscarnet to brincidofovir.⁴⁰

Foscarnet (60 mg/kg/day), a DNA polymerase inhibitor, has been shown in case reports of immunocompetent neonates to show resolution of liver CMV infection. Further reports of foscarnet, using 100 mg/kg/day dosing, have been associated with clinical improvement in infants with CMV-driven haemophagocytic lymphohistiocytosis and no significant side effects.^{41,42}

Letermovir inhibits the terminase complex and can be administered intravenously or orally. It is also less myelosuppressive and nephrotoxic than valganciclovir. Letermovir is being more commonly used as prophylaxis against CMV in HSCT recipients in adults and currently being investigated in a phase 2b trial in children post HSCT (NCT03940586). However, no trials have yet been conducted in neonates.

2.1.4 | Vaccines in development

Vaccination offers the main public health strategy to controlling cCMV. A variety of approaches are being developed including attenuated whole virus vaccines, protein subunit vaccines and mRNA vaccines. Glycoprotein B (gB) and pp65 provide immunodominant targets in association with CMV-directed T cell responses.

A phase II placebo-controlled, RCT of a CMV gB/MF59 adjuvanted vaccine showed a significant reduction of maternal primary infections in women in the adjuvanted vaccine arm (18/225 [8%]) versus placebo (31/216 [14%]).⁷ In a further RCT of 120 CMV seropositive women versus 30 women receiving placebo, there were boosted antibody and CD4+ responses in the treatment group (mean titres of 146 vs. 39 [day 28], 111 versus 40 [day 180], 90 vs. 33 [day 360]).⁴³ A vaccine containing both CMV gB and pp65 protein to elicit a broader array of immune responses is currently being trialled in a phase II study of renal transplant recipients.⁴⁴

Messenger RNA (mRNA) vaccines act through delivering mRNA sequences coding for pathogen-specific proteins into host cells that then express the proteins encoded by these sequences. Candidate mRNA vaccines (mRNA-1647 encoding antigens gB and the pentameric complex, and mRNA-1443 encoding antigens for pp65) have been shown to elicit neutralising antibodies in mice and primate models⁴⁵ and have completed phase I trials.⁴⁶ Now in phase II trials of healthy adult cohorts, the mRNA-1647⁴⁷ candidate encodes gB protein and subunits of the CMV pentamer complex required for viral cell entry. Phase III trials in women of childbearing age are currently recruiting.⁴⁸

The induction of anti-CMV neutralising antibodies as well as cytotoxic T cell responses by a live-attenuated 'Towne' CMV strain led to a trial of chimaeric Towne/non-attenuated Toledo vaccine candidate in a phase I study in CMV-seronegative men. These isolates have been formulated by passaging, resulting in mutations in the 'ULb' genome region containing sequences spanning open reading frames, resulting in attenuation. This candidate was well-tolerated and yielded detectable, yet low, levels of neutralising antibodies, and CD8+ T cell responses.⁴⁹

The V160 vaccine is a highly passaged replication-defective CMV virus gH/gL/pUL128-131, a natural surface pentameric complex promoting infectivity in epithelial and endothelial cells. Phase I trials in seronegative recipients, demonstrated that it was well-tolerated and resulted in a significant increase in antibody titres that persisted to 18 months, as well as CD8+ effector, effector memory and memory B cell responses.⁵⁰

Other multivalent vaccines are being trialled in HSCT recipients including the triplex vaccine combination of pp65/IE1-exon4/IE2-exon5 peptide vaccines which underwent phase II trials in CMV positive HSCT recipients—pp65 and IE1 were chosen as are immunodominant CMV antigenic targets associated with recognition by human CD4+ and CD8+ T cells and associated reduction in viraemia.⁵¹ This was well-tolerated and reduced reactivation-related viraemia.⁵² Phase I/II studies are also underway in paediatric populations undergoing HSCT.⁵³ A further City of Hope peptide vaccine (HLA A*0201/pp65) is also being trialled in phase II studies.⁵⁴

2.2 | Enteroviruses

2.2.1 | Current epidemiology

There are over 110 genetically distinct EV, classified into four major species: A (including EV71, Coxsackie A6/16—which cause hand, foot and mouth disease), B (including echoviruses, Coxsackie B3 causing a variety of presentations including gastrointestinal or myocarditis/pericarditis), C (including poliovirus 1–3, presenting with myelitis) and D (including EV68, associated with acute flaccid myelitis). Our understanding of circulating EV types is limited and based predominantly on passive surveillance from regional or national repositories. Transmission occurs from exposure to infected stool or eye/nasal/oral fluid, or vertical intrapartum acquisition. Most neonatal EV infections are asymptomatic. However, EV is the commonest cause of neonatal meningitis and can cause coagulopathy, hepatitis, myocarditis, with some fatalities. Some serotypes, EV-71 and EV-68, have been identified as causes of acute flaccid paralysis in a small proportion of infection.

A case series of 668 EV-infected infants <90 days old showed that fever (85%), irritability (66%) and reduced feeding (54%) were the commonest presenting features and 11% of cases required intensive care admission.² In these infants, sequelae at 12 months (0.6%) were rare.² However, our understanding of long-term enteroviral sequelae is limited, with only four studies having attempted to investigate the neurodevelopmental outcomes of infants and children with non-EV71 meningitis.^{55–58}

A Korean study of 802 children with EV meningitis showed only 63% of neonates had cerebrospinal fluid (CSF) pleocytosis.⁵⁹ This emphasises the importance of syndromic testing of EV in CSF of neonates with suspected meningitis. A retrospective cohort study of 345 EV positive infants <6 months old showed that a positive test was associated with reduced lengths of stay (51.6 vs. 74.1 h) and discontinuation of empirical antibiotics (65.4% vs. 48.5%).⁶⁰

It could be anticipated that the reported rates of EV meningitis will continue to rise as multiplex PCR assays, such as the BioFire FilmArray, become increasingly integrated across paediatric clinical services. However, the clinical impact of these newer diagnostic assays is still being understood.

2.2.2 | Current treatments

Developing antivirals against EV has not been a priority for the pharmaceutical industry and there are no licenced treatments for EV infections. Yearly hand, foot and mouth disease outbreaks in South East Asia reflect a significant EV71 burden.⁶¹ However, potential revenue from marketing EV71 antivirals may be limited in these resource-deprived settings. The manufacturing of antivirals against EV needs to account for their high mutation rate. The need for multiple drug agents to act on different viral targets to minimise resistance and limited cross genotypic activity further increases the challenge to successfully trial new antivirals.

Hypogammaglobulinaemia is associated with risk of enteroviral disease.⁶² The use of intravenous immunoglobulin (IVIG) therapeutically in enteroviral disease was initially supported by a randomised trial of 16 neonates.²⁵ In this trial, 750 mg/kg IVIG administered to nine infected infants increased serum neutralisation titres, and doses at neutralisation titres > or equal to 1:800 to the patients' viral isolates were associated with significantly higher increases in serum neutralising titres and reduced blood and urine viral load. Clinical outcomes of fever, duration of symptoms and residual symptoms at discharge were the same. IVIG was well-tolerated with possible adverse effects of fever, tachycardia and tachypnoea in three infants.²⁵ Another trial in 41 neonates <1 month with severe EV infection (hepatitis, coagulopathy, thrombocytopenia, myocarditis or meningitis) showed odds of survival 14.7 higher in those receiving IVIG within the first 72 h of presentation.²⁶ In adult enteroviral cases, a systematic review of IVIG in prospective trial/case series/reports demonstrates association with possible survival advantage in immunosuppressed patients with rare incidence of serious adverse events.⁶²

2.2.3 | Novel treatments

The EV capsid shell is flanked by four structural proteins and contains a hydrophobic pocket that can be targeted with capsid inhibitors, preventing receptor binding and uncoating of the virion particles following cell entry. A number of drug candidates target the capsid, including pleconaril, pocapavir and suramin (Table 2).

TABLE 2 Novel antivirals in therapeutic pipeline for severe neonatal CNS infections

	Novel antivirals	Mode of action	Stage of development	Results thus far
CMV	Cidofovir Foscarnet Filiciclovir (cyclopropovir) CRISPR/Cas9—genome-editing TAL effector nuclease (TALEN) plasmids—Plasmids containing restriction endonucleases with a propensity to cleave specific DNA sequences	Nucleoside analogue DNA polymerase inhibitor Nucleoside analogue catalysed by viral enzyme pUL97 which inhibits viral DNA polymerase Genome-editing technology that originated as an archaeal bacterial defence mechanism consisting a Cas9 protein, which has helicase and nuclease activity, allowing the system to cleave specific foreign RNA sequences following guiding by a small guide RNA (sgRNA) sequence Plasmids containing restriction endonucleases with a propensity to cleave specific DNA sequences	It is being trialled in the lipid conjugate form CMX001 in a current phase II/III study for adults and children with CMV, HSV, adenovirus, vaccinia and variolavirus infections ⁶³ Trialled in a cohort of 107 AIDS neonatal patients ⁶⁴ at IV 60 mg/kg for 2 weeks and compared to a group of 127 patients given IV ganciclovir for 5 mg/kg for 2 weeks ⁶⁴ Phase I, double-blind, randomised placebo-controlled trial of 24 CMV-naïve patients ⁶⁵ In vitro studies ⁶⁶ Mouse models ⁶⁷	A cohort study in neonatal adenoviral infection demonstrated renal toxicity and little efficacy. ³⁹ Awaiting phase II/III data Median survival of 12.6 months compared to 8.6 months, with equivalent rates of progression of retinitis ⁶⁴ Generally well tolerated but one participant developed renal dysfunction ⁶⁵ Cleaving HCMV UL122/123 gene for lytic replication/reactivation prevented 90% gene expression in fibroblasts. ⁶⁸ In other studies cleaving non-essential genes were targeted causes frameshifts in essential genes Transfection into mouse models resulted in inhibition of murine CMV replication reduced titres by 60%–75% and 25% at days 3 and 5 compared to controls and reduced transcription of CMV genes in murine organs by 10–100-fold ⁶⁷
EV	Capsid inhibitors Pleconaril NLD-22 Pocapavir. (V-073) Suramin Peptidomimetics for example, 3Cpro inhibitors for example, rupintrivir, ⁶⁹ AG7404, DC07090 ⁷⁰ 3Dpol inhibitors ⁷⁰ gemcitabine, NITD008, ribavirin, amiloride 2C inhibitors for example, fluoxetine analogues Monoclonal antibodies NA11F12 4 Engineered bispecific antibody Bs(scFv)4-IgG-1 OSBP inhibitors: itraconazole	Bind to the hydrophobic pocket of the viral capsid in viral protein 1 As above As above As above, narrower spectrum Binding of the viral capsid of EV-A71 3Cpro inhibitors: 3Dpol in conjugation with proteins 3Cpro and 3AB, which are involved in viral priming, and 2C with ATP-dependent helicase activity and acts as a ATP-independent RNA chaperone ⁷⁰ 3C inhibitor—inhibits viral priming 2C inhibitor—inhibits viral priming Viral neutralisation Viral neutralisation Binding to OSBP (factor in molecular translocation involved in EV replication)	Randomised placebo-controlled trial ⁷¹ In vitro ⁷² Case reports ^{73,74} Currently being trialled in phase 1 ^{75,76} In vitro ⁶⁹ In vitro ⁷⁷ Mouse models ⁷⁸ In vitro ⁷⁹ In vitro ⁸⁰	Improved survival and shorter times to negative culture in neonates presenting with enteroviral sepsis Promising antiviral activity in vitro at present Successful use even in cases with fulminant neonatal myocarditis Dose-dependent reduction in virions in rhabdomyosarcoma cell lines, blocking viral attachment to cells EV inhibition in cell lines EV inhibition in cell lines EV inhibition in cell lines Dose- and time-dependent cross-neutralisation activity against lethal-dose infection in murine models against Coxsackie A 16 ⁷⁸ Cross-reactivity against EV71 and CA16 in mice with complete therapeutic efficacy against these infections in mice ⁷⁹ Inhibits EV replication in cell lines by binding to OSBP ⁸⁰

TABLE 2 (Continued)

	Novel antivirals	Mode of action	Stage of development	Results thus far
Parechovirus	IVIG Monoclonal antibodies ⁸¹	Viral neutralisation Viral neutralisation	In vitro & case reports ^{27,28} Molecular studies ⁸¹	IVIG has been shown to provide potent in vitro antiviral activity, ²⁷ and indeed was used in the treatment of a patient with dilated cardiomyopathy ²⁸ Antibodies AM18 and AM28 bind to epitopes on VP1, preventing integrin binding to the viral capsid, and VP0/VP3, blocking RNA uncoating, respectively ⁸¹
HSV	Brincidofovir/CMX001 (a lipophilic derivative of cidofovir without nephrotoxicity) Pritelivir Amenamivir CRISPR/Cas9 Small interfering RNA (siRNA)	Nucleoside analogue Viral helicase/primase inhibitor Viral helicase/primase inhibitor Cleave latent HSV neuronal infection Cleave latent HSV neuronal infection	Currently being evaluated from clinical trial ⁶³ Phase II studies ^{82,83} Phase II/III studies ^{84,85} In vitro ⁸⁶ In vitro ⁸⁷	Awaited Reduce viral shedding in HSV2-seropositive individuals with annually recurrent genital herpes, over 28 days compared to oral valaciclovir. ⁸² A further open-label, multi-centre randomised phase II trial ongoing ⁸³ 3 days course reduced median lesion duration by 20.2 h compared to placebo ⁸⁴ equivalent days to cessation of new lesion formation, complete crusting and virus disappearance as valaciclovir ⁸⁵ Successfully carried out in human cell culture models, ⁸⁶ although delivery of the technology remains a major hurdle Sequences targeting the UL-29 HSV sequences reduced HSV replication and stimulated increased innate immune cytokine responses ⁸⁷

Abbreviations: CMV, cytomegalovirus; CNS, central nervous system; EV, enterovirus; HSV, herpes simplex virus; IVIG, intravenous immune globulin.

Pleconaril inhibits the uncoating of viral RNA and progeny virions during EV replication. Pleconaril has also been shown to have good oral bioavailability. In a double-blind placebo controlled RCT, infants up to 12 months with suspected EV meningitis were randomised to receive pleconaril (5 mg/kg/dose) orally three times daily for 7 days or placebo. Twelve subjects received pleconaril and 8 received placebo. There were no differences in symptoms or length of hospital stay in either group. Overall, 10/12 (83%) in the pleconaril group and 6/9 (75%) in the placebo group had adverse events, most commonly diarrhoea, rash and fever. The lack of clear evidence of efficacy led to early termination of the trial by the pharmaceutical sponsor.⁸⁸ Pleconaril is currently not licenced by the Food and Drug Administration, and has substantial drug interactions as a CYP3A4 inducer.

Pocapavir, a viral capsid inhibitor, has also shown efficacy against EV infection. It has been used with favourable outcomes in a case of enteroviral sepsis, with significantly improved myocardial output and coagulopathy, although these were improving prior to commencement of drug.⁸⁹ Two isolated case reports of neonates with fulminant neonatal myocarditis have demonstrated viral clearance and recovery within 2 weeks without adverse effects—currently this investigational drug has been only available in these limited number of cases as emergency treatment.^{73,74} Emergency investigational new drug approval from the Food and Drug Administration, parental consent, and placental enteroviral typing and pocapavir susceptibility testing were required prior to the use of the first case.⁸⁹ A further capsid binder, suramin, which inhibits EV-A71 is being trialled in a Phase I study in 36 healthy adults.⁷⁵

RNA replication in EV is carried out by the viral RNA-dependent RNA polymerase 3D^{pol} in conjunction with proteins 3C^{pro} and 3AB involved in viral priming, and 2C which has helicase/chaperone activity. 3C^{pro} inhibitors (e.g. rupintrivir),⁶⁹ 3D^{pol} inhibitors (including nucleoside and non-nucleoside analogues) and 2C inhibitors (e.g. fluoxetine) have been trialled in murine models, demonstrating reduction in Cocksackie viral titres after intraperitoneal administration,⁹⁰ but in human cohort studies of enteroviral D68 acute flaccid paralysis was not associated with poorer neurological outcomes.⁹¹

2.2.4 | Vaccines in development

Vaccine development is an active area of research, with greatest global efforts focussed on EV71 as the most problematic and virulent serotype. EV71 epidemics in 2009 and 2010 led to over a million cases, with thousands of hospitalisations and mortality in the hundreds.⁹² Development of a vaccine candidate with durable and efficacious immunogenicity may provide a reduction in incidence of these endemic viruses and thus reduce cases of primary maternal infection. This research field is in its early stages with limited vaccine candidate options at present, consisting of inactivated, multivalent and experimental Virus-Like Particle (VLP) vaccines.

The inactivated one-dose Human Diploid cell, KMB-17 Cell EV vaccine has been licenced in China and has recently undergone phase IV trials in children aged 6–71 months against hand, foot and mouth disease, with previous reported efficacy of 97%.⁹³ Furthermore, the inactivated, vero cell-based EV71 Sinovac vaccine showed non-inferior immunogenicity in infants 3–5 years to those aged 6–35 months.⁹⁴ A phase I bivalent vaccine IN-B001 for EV71/CV16 is currently being trialled in healthy adults.⁹⁵ The use of such polyvalent approaches offers promise in overcoming the heterogeneity of enteroviral types.

An inactivated EV 71 vaccine (INV21) was safe in healthy adults in a phase I double-blind study.⁹⁶ Meanwhile another human diploid-cell-based inactivated EV71 C4 genotype vaccine showed 97.1% efficacy in healthy children for EV71-related hand, foot and mouth disease.⁹⁷

A recent multivalent candidate has been the combination of recombinant Cocksackie B1 nanoparticles/Norovirus VLPs/Rotavirus oligomeric rVP6, which generated IgG1 and IgG2a antibodies and broad T cell responses in mice.⁹⁸ Experimental VLP vaccines are currently being trialled in and have been shown to protect mice from lethal infection against different EV's including EV71.⁹⁹

2.3 | Human parechoviruses

2.3.1 | Current epidemiology

HPeVs are a genus of viruses within the family Picornaviridae, alongside the Enterovirus genus. These are non-enveloped RNA viruses, of which there are 18 circulating HPeVs. Seasonal outbreak monitoring for parechoviral disease is an important public health

measure, as has been demonstrated in Australia, where there have been recent notable outbreaks in 2013–2014, 2015–2016 and 2017, with >200 cases of hospitalised HPeV infection in young infants.¹⁰⁰ All three epidemics in Australia have now shown to have been primarily attributed to the novel recombinant HPeV3,¹⁰⁰ with viral phylogenetic analysis of a cohort of 33 hospitalised infants in the third epidemic demonstrating significant stability of the HPeV3 indicating need for further therapeutics and vaccines.¹⁰¹

HPeVs are associated with frequent and significant illness in neonates and young infants. HPeV3 is the second most common cause of viral meningitis in neonates. In the UK, surveillance data showed that 23% (8/35) of cases required intensive care.¹⁰² Neonatal HPeV3 encephalitis has been associated with gross motor delay, cerebral palsy and visual impairment and has characteristic deep periventricular white matter changes. A study in 426 post mortem specimens in infants showed that 4% of deaths were associated with HPeV.¹⁰³ An Australian study revealed impaired neurodevelopmental outcomes in 11 of 77 infants with HPeV meningitis and a further study demonstrated 5/8 young infants at 12 months with parechovirus encephalitis had neurodevelopmental sequelae¹⁰⁴: cerebral palsy, gross motor impairment¹⁰⁵; however these findings have not been consistently replicated. A British Paediatric Surveillance Unit Study demonstrated no sequelae in 35 neonates with HPeV meningitis² at 12 months follow up and a Dutch study of 58 children with CNS infection showed no neurodevelopmental outcomes at 24 months' follow-up.¹⁰⁶

CSF pleocytosis in infants with HPeV meningitis is uncommon, with over 90% of CSF samples in HPeV meningitis containing 0–10 WCC/mm.² Routine testing of all CSF samples, irrespective of degree of inflammation, in infants with suspected meningitis should therefore be conducted. As with EV, the lack of active clinical and virological surveillance mechanisms has meant there is a paucity of epidemiological data to inform therapeutic trials.

2.3.2 | Current treatments

There are no licenced antiviral drugs for HPeV. IVIG has been shown to provide dose-dependent antiviral activity, with HPeV3 RNA intracellular levels decreasing from 4.71×10^7 to 2.83×10^3 copies/ μ l in vitro.²⁷ An uncontrolled case series of four febrile RT-PCR HPeV3 positive neonates <60 days, who received IVIG starting at day 1–4 of illness, showed some improvements in clinical presentation of rash, diarrhoea, apnoea and seizures.²⁹

2.3.3 | Novel treatments

No current antivirals are in phase II/III trials under [ClinicalTrials.gov](https://clinicaltrials.gov) as searched/accessed on 3 July 2022. Unlike EV, HPeVs are intrinsically resistant to the capsid binders due to a shallower capsid pocket, impeding the fit of molecular binding. A Finnish group demonstrated use of monoclonal antibodies as therapeutic candidates—molecular mapping showed antibodies AM18 and AM28

targeted to viral capsid (epitopes of RGD motif and amino acids from loops of VP0/VP3 respectively) preventing integrin binding to the viral capsid and blocking RNA uncoating respectively.⁸¹

2.3.4 | Vaccines in development

No vaccine candidates are being investigated in Phase I/II/III studies at present under ClinicalTrials.gov as of search/access date 3 July 2022.

2.4 | Herpes simplex virus

2.4.1 | Current epidemiology

HSV-1 and HSV-2 are double-stranded DNA viruses within the *Herpesviridae* family and transmitted by direct (orolabial mucosa) and sexual contact. HSV-2 causes the majority of cases of genital herpes, although HSV-1 has also emerged as a major cause of genital herpes infections. Following initial epithelial infection, the virus migrates towards local neuronal ganglia, establishing latency with the potential for reactivation.

We use the term 'neonatal' to include all cases of possible transmission of HSV (i.e. encompasses intrauterine, intrapartum and perinatal transmission). Between 2010 and 2015, there were 14,257 reported worldwide cases of neonatal herpes of which 2/3 were thought to be due to HSV-2 and 1/3 due to HSV-1.¹⁰⁷ The reported incidence of neonatal HSV is rare in HIC: 17.6 per 100,000 infants (UK),¹ 1.6 per 100,000 (Switzerland), 3.2 per 100,000 (Netherlands) and 8.5 per 100,000 (Israel) and 5–33 per 100,000 (US)¹⁰⁸ although higher in LMICs such as Kenya (33 per 100,000 births per year). The estimates based on HSV-1 and HSV-2 incidences amongst women have suggested that neonatal infection occurs at 0.0031%–0.0072% live births respectively; greatest incidence is in the Americas for HSV-1 (0.0126%) and Africa for HSV-2 (0.0153%).¹⁰⁷

Primary maternal infection in particular, as well as outbreaks of recurrent herpes ulceration, are associated with neonatal infection via vaginal delivery. The risk of transmission from mother to neonate is highest in primary genital herpes (25%–60%) and much lower (approximately 5%) to mothers with recurrent genital infection.¹⁰⁹ HSV maternal seropositivity is highly variable with documented prevalence of 63% for HSV-1 and 22% for HSV-2 (US),¹¹⁰ 62.5%/8.3% (Germany),¹¹¹ 95.7%/40.1% (Brazil)¹¹¹ and 32% for HSV-2 (Kenya).¹¹²

In the UK, the last published prospective study of HSV in neonates was performed between 1986 and 1991 and showed an incidence of 0.5 per 100,000 live births.¹¹³ In 2013, a UK study of laboratory confirmed HSV meningo-encephalitis cases showed an apparent four-fold increased incidence of 2.2 per 100,000 live births, which was attributed by the authors to be due to the introduction of PCR leading to higher diagnostic rates.¹¹⁴ A longitudinal study of routine collected patient records showed that the rate of neonates diagnosed in hospital with HSV increased nearly four-fold between

1999–2016.¹ This corresponds with a 22% increase in diagnosis of new episodes of genital HSV in England between 2008–2017.¹¹⁵ The increase in reported cases of neonatal HSV therefore highlights the importance of safe and effective treatments to manage this catastrophic disease.

Neonatal disease has 3 distinct phenotypes: skin/eye/mouth (SEM) (45%), CNS (30%) and disseminated disease (25%).¹⁰⁹ Typically, SEM and disseminated disease occurs at 2 weeks and CNS disease at 3 weeks of life.¹⁰⁹ Approximately half of neonates with SEM have recurrent cutaneous lesions without suppressive treatment.¹⁰⁹ A fifth of neonates with disseminated disease have neurologic sequelae at 1 year and with 70% of those with CNS disease have permanent neurologic impairment.³⁴

2.4.2 | Current treatments

The mainstay of treatment in HSV infection involves aciclovir, with other nucleoside analogue options, which all target the thymidine kinase enzyme (Table 1).

Aciclovir, famciclovir and valaciclovir are nucleoside analogues that target thymidine kinase. Parenteral aciclovir has been the first-line agent in management of neonatal HSV. This has been based on two RCTs that each took a decade to recruit. The first RCT in 210 neonates comparing vidarabine (neonatal cases of: 28 disseminated, 31 SEM, 36 CNS) and aciclovir (18 disseminated, 54 SEM, 35 CNS) and demonstrated equivalent tolerability and efficacy of aciclovir, although with notable disproportions of disease phenotypes between arms.³² A further RCT (41 disseminated, 28 CNS, 10 SEM) in neonates used 60 mg/kg/day dosing compared to 45 mg/kg/day dosing and showed 3.3× greater odds of survival in the disseminated groups; odds were similar in CNS groups.³⁰ Aciclovir is usually divided as thrice-daily dosing for 14 days, or 21 days if disease is disseminated or with CNS involvement. Renal toxicity (5.6% (57/1017) in neonates in one retrospective US cohort study from 2011 to 2015 occurred more often in those with intensive care stay or mechanical ventilation³¹ and neutropenia (21% of 29 neonates; all spontaneously resolved³⁰) but these significant side effects require careful monitoring.

A suppressive course of oral aciclovir for 6 months follows the initial intravenous treatment course for neonatal HSV. This is based on a single phase III RCT of 74 neonates with CNS (45) or SEM disease (29) of which 6 and 3 neonates respectively did not complete the 6 months course; follow-up demonstrated 6 months of oral aciclovir reduced disease recurrence at 12 months (approximately 35% less for CNS disease and 30% less for SEM disease).³⁴ Neonates with CNS infection that had Bayley neurodevelopmental assessments undertaken and randomised to the aciclovir arm also had improved developmental outcomes at 12 months—69% (11/16) versus 33% (4/12); no differences were seen for SEM disease.³⁴ A study of 5-year duration is currently underway to investigate the safety and pharmacokinetics of enteral valaciclovir in 10 neonates 2–12 weeks in age and 34+ gestation with confirmed HSV.¹¹⁶

In women with HSV, management of the neonate depends on whether maternal infection is primary or recurrent. In primary infection, skin/mucosal swabs and blood for PCR should be sent at 24 h post-delivery, as well as CSF for HSV PCR, and IV aciclovir should be commenced.¹¹⁷ In recurrent infection, US guidance from the American Academy of Paediatrics advises that given likelihood of transmission to neonate is low (2%), acyclovir should not be started unless symptomatic; viral skin and mucosal swabs should be taken at 24 h and if negative alongside an asymptomatic neonate, the neonate can be discharged; if positive, the neonate should be treated for 10 days in order to prevent symptomatic infection.¹¹⁷ However, the British Association for Sexual Health and HIV (BASHH) guidance advises that in recurrent HSV with or without lesions, there is no need to test or treat the neonate.¹¹⁸

Foscarnet has some activity in aciclovir-resistant infections with risks of significant nephrotoxicity and metabolic derangement. Two case reports of foscarnet administered at day 18 and day 11 of infection in neonates demonstrated clinical recovery after HSV infection of the larynx in the first case and pneumonitis, hepatitis and DIC following herpes infection, resulting in death in the second.¹¹⁹ Aciclovir remains the mainstay of treating neonatal HSV. However, resistance through thymidine kinase mutations, could theoretically develop in longer treatment durations. Aciclovir also causes nephrotoxicity, marrow suppression and requires prolonged inpatient hospital stay during the intravenous treatment phase, highlighting the need for novel therapeutics.

2.4.3 | Novel treatments

Given the susceptibility for nucleoside analogue therapy to viral immune evasion via thymidine kinase mutations, there is a pressing need for development of novel agents with alternative mechanisms of action. These include broader acting nucleotide analogues (brincidofovir), pyrophosphate analogues that prevent binding onto DNA polymerase (foscarnet) and viral helicase/primase inhibitors (pritelivir and amenamevir).

Brincidofovir (a lipophilic derivative of cidofovir without nephrotoxicity also known as CMX001) and Pritelivir (helicase primase inhibitor) are drugs being evaluated for acyclic nucleoside resistant viruses. Brincidofovir was recently licenced by the FDA for use in smallpox infections. For HSV, thus far its use has been limited to case reports/series of immunocompromised adult and paediatric patients, where it demonstrated virological response,^{120,121} and in HSV prophylaxis of stem cell adult and paediatric transplant recipients where breakthrough was 1.0 per 1000 patient days.¹²² It is currently being evaluated in phase II/III trials for adults and children with DNA virus infections including HSV,⁶³ although its use may be limited due to toxicity profile and concerns of emerging cross-resistance.¹²³

Pritelivir (AIC316) and amenamevir (ASP2151) are viral helicase/primase inhibitors and inhibit viral replication by preventing the separation of DNA strands. 100 mg once daily oral pritelivir has been demonstrated in phase II double-blind study to reduce viral shedding

in HSV2-seropositive individuals with recurrent genital herpes.⁸² A further open-label, multi-centre randomised phase II trial (PRIOH-1) in immunocompromised adults of oral pritelivir versus intravenous foscarnet is ongoing.⁸³ A 3 days course of amenamevir was demonstrated, in recurrent genital herpes, to reduce median lesion duration by 20.2 h compared to placebo in a randomised trial of 437 adults and was well-tolerated.⁸⁴ Further phase III testing of 751 patients treated 72 h after development of rash with 7 days courses of amenamevir 400 mg OD, 200 mg OD and valaciclovir 1000 mg TDS for 7 days showed equivalent times to cessation of new lesion formation, complete crusting and virus disappearance.⁸⁵

2.4.4 | Vaccines in development

The development of HSV vaccination is an active area, with several late phase trials in progress. Several vaccine approaches are in trial including subunit vaccines, multi-epitope and live attenuated vaccines.¹²⁴ Of the 80+ proteins encoded by each of the HSV-1 and HSV-2 genomes, 12 glycoproteins are present on the viral envelope and five of which are recognised to interact with cellular receptors and promote viral entry (gB, gC, gD, gH and gL). Other important antigenic targets including the tegument proteins in the viral capsid involved in anchoring viral structure including during proliferation.¹²⁴

Previous HSV vaccine approaches utilised subunit vaccines containing gD (a surface viral protein involved in viral entry) and gB, although these have had limited efficacy—the latter demonstrated only 58% and 20% efficacy for HSV1 and HSV2 respectively in a randomised double-blind study of 8328 seronegative women.¹²⁵

One alternative approach is utilising different epitopes to produce broader T cell responses, including intraviral proteins such as UL25, ICP0 and ICP4 A DNA vaccine containing IL-2, IL-21 and MIP-1alpha elicited specific T cell response and viral elimination in mice.¹²⁶ gD with UL25 epitopes inserted into a recombinant adenovirus 5 vector also demonstrated immunogenicity through IFN-dependent T cell responses and reduction in genital lesions in mice.¹²⁷

To overcome the issue of subunit vaccines providing inadequate breadth of epitopes a live attenuated vaccine was developed. VC2 with gK and UL20 deletions prevents axonal translocation of virus and reduces mortality in mice¹²⁸ and reduced HSV-2 related neuronal infection in macaques.¹²⁹ A trivalent vaccine of gD2/gC2/gE2 containing gC2 and gE2 that are two HSV factors involved in immune evasion through binding to complement 3 and the Fc region of IgG respectively demonstrated immunogenicity in macaques.¹³⁰

2.5 | Future research priorities

We highlight key areas relating to future research priorities for improving the treatment of neonatal viral infections (Table 3). Broadly, few robust epidemiological studies have been carried out for these viral infections in LMICs to ascertain incidence of clinical

TABLE 3 Research priorities to inform the therapeutic pipeline to treat severe neonatal CNS viral infections

Issues		Study suggestions
cCMV		
Understanding pathophysiology	<p>Further evaluation of the phenotypic differences and epidemiology of primary versus non-primary maternal CMV infection in neonates is required</p> <p>Maternal CMV infection may result in placental immune dysfunction and placental insufficiency although the exact mechanisms for this have not been elucidated</p> <p>The role for T cells in CMV immunity was implied from demonstrations that adoptive T cell immunotherapy improved CMV response in allogenic haematopoietic stem cell recipients but has not been fully characterised</p>	<p>Antenatal screening studies to understand pathophysiological cCMV effects on neonates from primary versus non-primary infections and ascertain if clinical, immunological or virological biomarkers predict outcome</p> <p>Immunological studies evaluating placental markers and histopathological effects of maternal CMV non-primary and primary infection at different stages in pregnancy</p> <p>Animal knockout model studies may influence de novo vaccine targets</p>
Identification of infection	<p>To date, epidemiological data have described neonates with symptomatic disease. Our understanding of asymptomatic disease is highly limited</p> <p>There are no robust longitudinal data that describe the natural history of CMV disease</p>	<p>Universal screening studies of cCMV across different populations which enable opportunity to obtain host and virological data</p> <p>Registry that allows prospective longitudinal follow up and opportunity to trial novel interventions</p>
Treatment	<p>Use of therapeutic agents to reduce vertical transmission and investigated in randomised controlled trials</p> <p>Vaccine trials in both seronegative and seropositive women of childbearing age</p> <p>PK/PD data for valganciclovir and ganciclovir is required in neonates aged <32 weeks</p> <p>Trials of valganciclovir only enrolled in cases with severe disease</p> <p>Further trials on treatment of asymptomatic neonates and mild symptomatic disease are needed</p>	<p>Further phase II/III trials of anti-CMV agents in pregnancy, especially for cases of non-primary infection</p> <p>Further phase II/III vaccine trials for women of reproductive age who are seropositive and seronegative</p> <p>PK/PD studies of current therapeutic agents especially in neonates <32 weeks</p> <p>Investigation of effects of valganciclovir in isolated SNHL/mild disease and asymptomatic neonates</p> <p>Phase I-III clinical trials of asymptomatic neonates/symptomatic receiving ganciclovir/valganciclovir</p>
EV		
Understanding pathophysiology	<p>Further characterisation of disease subtypes by variation in EV virus type may optimise management</p>	<p>Active enhanced surveillance mechanism. This would be conducted by regional virology laboratories submitting EV positive samples to a reference laboratory for real-time molecular characterisation</p>
Identification of infection	<p>There is a lack of robust neurodevelopmental studies—more data is needed on long-term outcomes of viral meningitis</p> <p>There are no reliable biomarkers which predict neonates who develop severe disease/long term sequelae</p>	<p>Long-term neurodevelopmental outcomes of EV infection are required to assess burden of disease</p> <p>Collect samples for host transcriptomic and immunological profiling during acute illness and convalescence</p>
Treatment	<p>The largest trial of IVIG treatment in neonatal enteroviral sepsis recruited 41 participants²⁶</p> <p>Capsid inhibitors have not been utilised in neonatal trials other than case studies at present</p> <p>Combination therapy will be needed to prevent development of drug resistance; for example, consecutively alternating administration of pleconaril/MDL-860/oxoglaucine improved effect of individual administration in coxsackie-B1 infected mice¹³¹</p>	<p>RCT of IVIG in EV meningitis</p> <p>Phase I neonatal studies are required to evaluate the safety and efficacy of capsid inhibitors</p> <p>Animal and phase I trials of combination therapies of capsid inhibitors with other novel agents could improve therapeutic effect</p>
HPeV		
Understanding pathophysiology	<p>Understanding of host factors involved in RNA replication, protein synthesis and attachment and uncoating (relative to EV little understanding)</p>	<p>Further in vitro molecular work</p>
Identification of infection	<p>No large-scale epidemiological studies have been carried out</p> <p>At present there is a paucity of understanding of neurodevelopmental outcomes and mortality</p> <p>Disease markers of severity for example, urinary beta-2-microglobulin²⁹</p>	<p>Surveillance of HPeV in LMIC using syndromic testing of HPeV in neonates with suspected CNS infection</p> <p>Long term neurodevelopmental studies using robust tools for example, Bayley assessment</p> <p>Further identification of disease markers</p>

(Continues)

TABLE 3 (Continued)

	Issues	Study suggestions
Treatment	Development of therapeutic targets to host factors involved in RNA replication, protein synthesis and attachment and uncoating Monoclonal antibodies have been promising therapeutic candidates	Further molecular work is required before in vitro studies Further molecular studies to identify appropriate pathogenic targets
HSV		
Understanding pathophysiology	Understanding burden of viral evasion of suppressive therapy for ongoing viral shedding	Further epidemiological studies are required
Identification of infection	Low proportions of testing—need to evaluate outcomes with improved recognition of possible HSV cases	Large scale cohort studies following increased PCR testing or testing algorithms
Treatment	Viral evasion of maternal suppressive therapy for ongoing viral shedding Development of aciclovir resistance; further studies required within neonatal population of polymerase/helicase/primase inhibitors—foscarnet and cidofovir have been poorly studied in this population Current subunit and live attenuated vaccinations are being trialled in non-pregnant adults alone	Phase I studies of combination therapies to reduce viral shedding Phase I studies in neonatal population An understanding of effect on vaccination on vertical transmission will require phase I trials in pregnant women

Abbreviations: cCMV, congenital cytomegalovirus; CNS, central nervous system; EV, enterovirus; HPeV, human parechovirus; HSV, herpes simplex virus; IVIG, intravenous immune globulin; LMIC, low middle income countries; PCR, polymerase chain reaction; RCT, randomised controlled trial; SNHL, sensorineural hearing loss.

burden, and ongoing epidemiological analysis is required to drive further research on therapies.

In particular, the advent of mRNA vaccine rollout in the COVID pandemic offers a new avenue of vaccine development for this range of viruses. mRNA vaccine technology could potentiate increased output of vaccine manufacture and the breadth of feasible antigenic targets limited by historical expression technologies.¹³²

2.5.1 | CMV

Preventing transmission of infection to the foetus is critical. To achieve this, developing a range of vaccine candidates and investing their potential in phase II/III trials is essential. In the meantime, qualitative studies are required to help implement behavioural strategies that minimise acquisition of infection during pregnancy and can be promptly implemented into routine antenatal care. International collaborative epidemiological studies, through universal screening and development of a registry of affected cases, are required to understand the burden of congenital disease and prioritise treatment trials accordingly. Treatment trials of asymptomatic/mildly symptomatic disease are required to determine their efficacy in preventing late onset SHNL.

2.5.2 | Enteroviruses

Improving our understanding of the molecular epidemiology is essential to understand the commonest and most severe forms of EV meningitis. A phase III RCT of IVIG is needed to better understand its

effect on clinical outcome. In addition, Phase I/II studies of novel capsid inhibitors should be encouraged. It is likely that combination therapies will be required to prevent development of drug resistance. Further data on long-term neurodevelopmental outcomes of EV meningitis is required to inform the evidence base to manage affected infants after hospital discharge.

2.5.3 | Human parechoviruses

Surveillance studies should be conducted in LMIC in order to gain an understanding for the impact of HPeV in neonates. Improved our knowledge of protein synthesis, molecular cascades and targeting attachment/uncoating will permit development of further drug targets.

2.5.4 | HSV

The issue of aciclovir-resistant infections gives rise to the need for further studies within neonatal populations of foscarnet and cidofovir, as well as novel helicase/primase inhibitors. Given the HSV evasion of suppressive therapy, an evaluation of current vaccine candidates on vertical transmission is needed and will require phase I trials in pregnant women.

3 | CONCLUSIONS

The burden of neonatal viral infections is a significant cause of morbidity and mortality worldwide. However, therapeutic options are

highly limited. Academic institutions, pharmaceutical industry and policymakers should work together to develop the evidence to better treat neonates from this poorly understood group of severe infections.

AUTHOR CONTRIBUTIONS

Seilesh Kadambari conceived the idea for the paper. Movin Abeywickrema wrote the first draft of the manuscript. Movin Abeywickrema, Dominic Kelly and Seilesh Kadambari contributed to the literature review and provided scientific content to each section of the manuscript.

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DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available in Pubmed at <https://pubmed.ncbi.nlm.nih.gov/>. These data were derived from the following resources available in the public domain: <https://pubmed.ncbi.nlm.nih.gov/>.

REFERENCES

- Kadambari S, Pollard AJ, Goldacre MJ, et al. Congenital viral infections in England over five decades: a population-based observational study. *Lancet Infect Dis*. 2020;20:220-229. [https://doi.org/10.1016/S1473-3099\(19\)30416-5](https://doi.org/10.1016/S1473-3099(19)30416-5)
- Kadambari S, Braccio S, Ribeiro S, et al. Enterovirus and parvovirus meningitis in infants younger than 90 days old in the UK and Republic of Ireland: a British Paediatric Surveillance Unit study. *Arch Dis Child*. 2019;104(6):552-557. <https://doi.org/10.1136/archdischild-2018-315643>
- Jones CA, Raynes-Greenow C, Isaacs D; on behalf of the Neonatal HSV Study Investigators and Contributors to the Australian Paediatric Surveillance Unit. Population-based surveillance of neonatal herpes simplex virus infection in Australia, 1997–2011. *Clin Infect Dis*. 2014;59(4):525-531. <https://doi.org/10.1093/cid/ciu381>
- Handel S, Klingler EJ, Washburn K, Blank S, Schillinger JA. Population-based surveillance for neonatal herpes in New York City, April 2006–September 2010. *Sex Transm Dis*. 2011;38(8):705-711. <https://doi.org/10.1097/OLQ.0b013e31821b178f>
- Chierighin A, Verucchi G, Lazzarotto T. CMV-specific cell-mediated immunity in immunocompetent adults with primary CMV infection: a case series and review of the literature. *Viruses*. 2021;13(5):816. <https://doi.org/10.3390/v13050816>
- Kenneson A, Cannon MJ. Review and meta-analysis of the epidemiology of congenital cytomegalovirus (CMV) infection. *Rev Med Virol*. 2007;17(4):253-276. <https://doi.org/10.1002/rmv.535>
- Pass RF, Zhang C, Evans A, et al. Vaccine prevention of maternal cytomegalovirus infection. *N Engl J Med*. 2009;360(12):1191-1199. <https://doi.org/10.1056/NEJMoa0804749>
- Barton M, Forrester AM, McDonald J. Update on congenital cytomegalovirus infection: prenatal prevention, newborn diagnosis, and management. *Paediatr Child Health*. 2020;25:395-396. <https://doi.org/10.1093/pch/pxaa083>
- Kimberlin DW, Jester PM, Sánchez PJ, et al. Valganciclovir for symptomatic congenital cytomegalovirus disease. *N Engl J Med*. 2015;372(10):933-943. <https://doi.org/10.1056/NEJMoa1404599>
- Retzler J, Hex N, Bartlett C, et al. Economic cost of congenital CMV in the UK. *Arch Dis Child*. 2019;104(6):559-563. <https://doi.org/10.1136/archdischild-2018-316010>
- Zuhair M, Smit GSA, Wallis G, et al. Estimation of the worldwide seroprevalence of cytomegalovirus: a systematic review and meta-analysis. *Rev Med Virol*. 2019;29(3):e2034. <https://doi.org/10.1002/rmv.2034>
- Prasoon KR, Srinadh B, Sunitha T, et al. Seroprevalence and influence of torch infections in high risk pregnant women: a large study from South India. *J Obstet Gynaecol India*. 2015;65(5):301-309. <https://doi.org/10.1007/s13224-014-0615-3>
- Marsico C, Kimberlin DW. Congenital cytomegalovirus infection: advances and challenges in diagnosis, prevention and treatment. *Ital J Pediatr*. 2017;43(1):38. <https://doi.org/10.1186/s13052-017-0358-8>
- Dar L, Namdeo D, Kumar P, et al. Congenital cytomegalovirus infection and permanent hearing loss in rural North Indian children. *Pediatr Infect Dis J*. 2017;36(7):670-673. <https://doi.org/10.1097/INF.0000000000001527>
- Putri ND, Wiyatno A, Dhenni R, et al. Birth prevalence and characteristics of congenital cytomegalovirus infection in an urban birth cohort, Jakarta, Indonesia. *Int J Infect Dis*. 2019;86:31-39. <https://doi.org/10.1016/j.ijid.2019.06.009>
- Madrid L, Varo R, Maculuvé S, et al. Congenital cytomegalovirus, parvovirus and enterovirus infection in Mozambican newborns at birth: a cross-sectional survey. *PLoS One*. 2018;13(3):e0194186. <https://doi.org/10.1371/journal.pone.0194186>
- Revello MG, Tibaldi C, Masuelli G, et al. Prevention of primary cytomegalovirus infection in pregnancy. *EBioMedicine*. 2015;2(9):1205-1210. <https://doi.org/10.1016/j.ebiom.2015.08.003>
- Shahar-Nissan K, Pardo J, Peled O, et al. Valaciclovir to prevent vertical transmission of cytomegalovirus after maternal primary infection during pregnancy: a randomised, double-blind, placebo-controlled trial. *Lancet*. 2020;396(10253):779-785. [https://doi.org/10.1016/S0140-6736\(20\)31868-7](https://doi.org/10.1016/S0140-6736(20)31868-7)
- Revello MG, Lazzarotto T, Guerra B, et al. A randomized trial of hyperimmune globulin to prevent congenital cytomegalovirus. *N Engl J Med*. 2014;370(14):1316-1326. <https://doi.org/10.1056/NEJMoa1310214>
- Kimberlin DW, Lin C-Y, Sánchez PJ, et al. Effect of ganciclovir therapy on hearing in symptomatic congenital cytomegalovirus disease involving the central nervous system: a randomized, controlled trial. *J Pediatr*. 2003;143(1):16-25. [https://doi.org/10.1016/s0022-3476\(03\)00192-6](https://doi.org/10.1016/s0022-3476(03)00192-6)
- Oliver SE, Cloud GA, Sánchez PJ, et al. Neurodevelopmental outcomes following ganciclovir therapy in symptomatic congenital cytomegalovirus infections involving the central nervous system. *J Clin Virol*. 2009;46(suppl 4):S22-S26. <https://doi.org/10.1016/j.jcv.2009.08.012>
- Zhou XJ, Gruber W, Demmler G, et al. Population pharmacokinetics of ganciclovir in newborns with congenital cytomegalovirus infections. NIAID Collaborative Antiviral Study Group. *Antimicrob Agents Chemother*. 1996;40(9):2202-2205. <https://doi.org/10.1128/aac.40.9.2202>
- Kimberlin DW, Acosta EP, Sánchez PJ, et al. Pharmacokinetic and pharmacodynamic assessment of oral valganciclovir in the treatment of symptomatic congenital cytomegalovirus disease. *J Infect Dis*. 2008;197(6):836-845. <https://doi.org/10.1086/528376>
- Alsalem M. Intravenous immune globulin uses in the fetus and neonate: a review. *Antibodies*. 2020;9(4):60. <https://doi.org/10.3390/antib9040060>
- Abzug MJ, Keyserling HL, Lee ML, Levin MJ, Rotbart HA. Neonatal enterovirus infection: virology, serology, and effects of intravenous immune globulin. *Clin Infect Dis*. 1995;20(5):1201-1206. <https://doi.org/10.1093/clinids/20.5.1201>
- Yen M-H, Huang Y-C, Chen M-C, et al. Effect of intravenous immunoglobulin for neonates with severe enteroviral infections

- with emphasis on the timing of administration. *J Clin Virol*. 2015;64:92-96. <https://doi.org/10.1016/j.jcv.2015.01.013>
27. Izumita R, Aizawa Y, Watanabe K, Saitoh A. A role of intravenous immunoglobulin in human parechovirus type 3 infection in an in vitro model. *Open Forum Infect Dis*. 2016;3(suppl_1). <https://doi.org/10.1093/ofid/ofw172.20>
 28. Wildenbeest JG, Wolthers KC, Straver B, Pajkrt D. Successful IVIG treatment of human parechovirus-associated dilated cardiomyopathy in an infant. *Pediatrics*. 2013;132(1):e243-e247. <https://doi.org/10.1542/peds.2012-1136>
 29. Azuma J, Yamamoto T, Sakurai M, et al. Urinary β 2-microglobulin as an early marker of infantile enterovirus and human parechovirus infections. *Medicine (Baltim)*. 2018;97(43):e12930. <https://doi.org/10.1097/MD.00000000000012930>
 30. Kimberlin DW, Lin C-Y, Jacobs RF, et al. Safety and efficacy of high-dose intravenous acyclovir in the management of neonatal herpes simplex virus infections. *Pediatrics*. 2001;108(2):230-238. <https://doi.org/10.1542/peds.108.2.230>
 31. Downes KJ, Boge CLK, Baro E, et al. Acute kidney injury during treatment with intravenous acyclovir for suspected or confirmed neonatal herpes simplex virus infection. *J Pediatr*. 2020;219:126-132.e2. <https://doi.org/10.1016/j.jpeds.2019.12.056>
 32. Whitley R, Arvin A, Prober C, et al. A controlled trial comparing vidarabine with acyclovir in neonatal herpes simplex virus infection. *N Engl J Med*. 1991;324(7):444-449. <https://doi.org/10.1056/NEJM199102143240703>
 33. Sampson MR, Bloom BT, Lenfestey RW, et al. Population pharmacokinetics of intravenous acyclovir in preterm and term infants. *Pediatr Infect Dis J*. 2014;33(1):42-49. <https://doi.org/10.1097/01.inf.0000435509.75114.3d>
 34. Kimberlin DW, Whitley RJ, Wan W, et al. Oral acyclovir suppression and neurodevelopment after neonatal herpes. *N Engl J Med*. 2011;365(14):1284-1292. <https://doi.org/10.1056/NEJMoa1003509>
 35. Tod M, Lokiec F, Bidault R, et al. Pharmacokinetics of oral acyclovir in neonates and in infants: a population analysis. *Antimicrob Agents Chemother*. 2001;45(1):150-157. <https://doi.org/10.1128/AAC.45.1.150-157.2001>
 36. National Institute of Allergy and Infectious Diseases (NIAID). A phase II, single stage, single-arm investigation of oral valganciclovir therapy in infants with asymptomatic congenital cytomegalovirus infection. [clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/NCT03301415). 2020. Accessed December 6, 2020. <https://clinicaltrials.gov/ct2/show/NCT03301415>
 37. Cytomegalovirus. Accessed January 26, 2021. <https://legacyscreening.phe.org.uk/cytomegalovirus>
 38. Marshall BC, Koch WC. Antivirals for cytomegalovirus infection in neonates and infants: focus on pharmacokinetics, formulations, dosing, and adverse events. *Paediatr Drugs*. 2009;11(5):309-321. <https://doi.org/10.2165/11316080-000000000-00000>
 39. Vora SB, Brothers AW, Englund JA. Renal toxicity in pediatric patients receiving cidofovir for the treatment of adenovirus infection. *J Pediatric Infect Dis Soc*. 2017;6(4):399-402. <https://doi.org/10.1093/jpids/pix011>
 40. Tippin TK, Morrison ME, Brundage TM, Mommeja-Marin H. Brincidofovir is not a substrate for the human organic anion transporter 1: a mechanistic explanation for the lack of nephrotoxicity observed in clinical studies. *Ther Drug Monit*. 2016;38(6):777-786. <https://doi.org/10.1097/FTD.0000000000000353>
 41. Nigro G, Sali E, Anceschi MM, et al. Foscarnet therapy for congenital cytomegalovirus liver fibrosis following prenatal ascites. *J Matern Fetal Neonatal Med*. 2004;15(5):325-329. <https://doi.org/10.1080/14767050410001701349>
 42. Knorr B, Kessler U, Pöschl J, Poschl J, Fickenscher H, Linderkamp O. A haemophagocytic lymphohistiocytosis (HLH)-like picture following breastmilk transmitted cytomegalovirus infection in a preterm infant. *Scand J Infect Dis*. 2007;39(2):173-176. <https://doi.org/10.1080/00365540600786598>
 43. Sabbaj S, Pass RF, Goepfert PA, et al. Glycoprotein B vaccine is capable of boosting both antibody and CD4 T-cell responses to cytomegalovirus in chronically infected women. *J Infect Dis*. 2011;203(11):1534-1541. <https://doi.org/10.1093/infdis/jir138>
 44. Hookipa Biotech GmbH. A randomized, placebo-control, phase 2 study of HB-101, a bivalent cytomegalovirus (CMV) vaccine, in CMV-seronegative recipient (R-) patients awaiting kidney transplantation from living CMV-seropositive donors (D+). [clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/NCT03629080). 2020. Accessed December 30, 2020. <https://clinicaltrials.gov/ct2/show/NCT03629080>
 45. John S, Yuzhakov O, Woods A, et al. Multi-antigenic human cytomegalovirus mRNA vaccines that elicit potent humoral and cell-mediated immunity. *Vaccine*. 2018;36(12):1689-1699. <https://doi.org/10.1016/j.vaccine.2018.01.029>
 46. ModernaTX, Inc. A phase 1, randomized, observer-blind, placebo-controlled, dose-ranging study to evaluate the safety, reactivity, and immunogenicity of cytomegalovirus vaccines mRNA-1647 and mRNA-1443 when administered to healthy adults. clinicaltrials.gov. 2019. Accessed December 7, 2020. <https://clinicaltrials.gov/ct2/show/NCT03382405>
 47. ModernaTX, Inc. A phase 2, randomized, observer-blind, placebo-controlled, dose-finding trial to evaluate the safety and immunogenicity of cytomegalovirus vaccine mRNA-1647 in healthy adults. clinicaltrials.gov. 2020. Accessed December 7, 2020. <https://clinicaltrials.gov/ct2/show/NCT04232280>
 48. Moderna completes enrollment of cytomegalovirus (CMV) vaccine (mRNA-1647) phase 2 study | Moderna, Inc. Accessed February 23, 2021. <https://investors.modernatx.com/news-releases/news-release-details/moderna-completes-enrollment-cytomegalovirus-cmv-vaccine-mrna/>
 49. Adler SP, Manganello A-M, Lee R, et al. A phase 1 study of 4 live, recombinant human cytomegalovirus townes/toledo chimera vaccines in cytomegalovirus-seronegative men. *J Infect Dis*. 2016;214(9):1341-1348. <https://doi.org/10.1093/infdis/jiw365>
 50. Adler SP, Lewis N, Conlon A, et al. Phase 1 clinical trial of a conditionally replication-defective human cytomegalovirus (CMV) vaccine in CMV-seronegative subjects. *J Infect Dis*. 2019;220(3):411-419. <https://doi.org/10.1093/infdis/jiz141>
 51. Wang Z, La Rosa C, Li Z, et al. Vaccine properties of a novel marker gene-free recombinant modified vaccinia ankara expressing immunodominant CMV antigens pp65 and IE1. *Vaccine*. 2007;25(6):1132-1141. <https://doi.org/10.1016/j.vaccine.2006.09.067>
 52. Plotkin SA, Wang D, Oualim A, et al. The status of vaccine development against the human cytomegalovirus. *J Infect Dis*. 2020;221(supplement_1):S113-S122. <https://doi.org/10.1093/infdis/jiz447>
 53. Multi-antigen CMV-modified vaccinia ankara vaccine in treating pediatric patients with positive cytomegalovirus undergoing donor stem cell transplant - full text view - ClinicalTrials.gov. Accessed December 8, 2020. <https://clinicaltrials.gov/ct2/show/NCT03354728>
 54. City of Hope Medical Center. A phase II randomized, placebo-controlled, multicenter trial to evaluate protective function of an optimized dose of CMVPepVax in recipients of an allogeneic hematopoietic stem cell transplant. clinicaltrials.gov. 2019. Accessed December 30, 2020. <https://clinicaltrials.gov/ct2/show/NCT02396134>
 55. Baker RC, Kummer AW, Schultz JR, Ho M, Gonzalez del Rey J. Neurodevelopmental outcome of infants with viral meningitis in the first three months of life. *Clin Pediatr (Phila)*. 1996;35(6):295-301. <https://doi.org/10.1177/000992289603500602>
 56. Balasubramanian H, Wagh D, Rao S, Keil AD, McMichael J. Developmental outcomes in cerebrospinal fluid proven enteroviral meningitis in neonates > 32 weeks of gestation. *J Paediatr Child Health*. 2016;52(3):327-332. <https://doi.org/10.1111/jpc.13083>
 57. Chang L-Y, Huang L-M, Gau SS-F, et al. Neurodevelopment and cognition in children after enterovirus 71 infection. *N Engl J Med*.

- 2007;356(12):1226-1234. <https://doi.org/10.1056/NEJMoa065954>
58. Gau SS-F, Chang L-Y, Huang L-M, Fan TY, Wu YY, Lin TY. Attention-deficit/hyperactivity-related symptoms among children with enterovirus 71 infection of the central nervous system. *Pediatrics*. 2008;122(2):e452-e458. <https://doi.org/10.1542/peds.2007-3799>
 59. Song JY, Nam SO, Kim YA, et al. Cerebrospinal fluid non-pleocytosis in pediatric enteroviral meningitis: large-scale review. *Pediatr Int*. 2018;60(9):855-861. <https://doi.org/10.1111/ped.13658>
 60. Lee BR, Sasidharan A, Harrison CJ, Selvarangan R. Positive impact of routine testing for enterovirus and parechovirus on length of hospitalization and antimicrobial use among inpatients ≤ 6 months of age. *J Clin Microbiol*. 2020;59(1). <https://doi.org/10.1128/JCM.02106-20>
 61. Puenpa J, Wanlapakorn N, Vongpunsawad S, Poovorawan Y. The history of enterovirus A71 outbreaks and molecular epidemiology in the Asia-Pacific region. *J Biomed Sci*. 2019;26(1):75. <https://doi.org/10.1186/s12929-019-0573-2>
 62. Wagner JN, Leibetseder A, Troescher A, Panholzer J, von Oertzen TJ. Characteristics and therapy of enteroviral encephalitis: case report and systematic literature review. *Int J Infect Dis*. 2021;113:93-102. <https://doi.org/10.1016/j.ijid.2021.10.002>
 63. Chimerix A. Multicenter, open-label study of CMX001 treatment of serious diseases or conditions caused by dsDNA viruses. clinicaltrials.gov. 2020. Accessed November 17, 2020. <https://clinicaltrials.gov/ct2/show/NCT01143181>
 64. Reusser P, Einsele H, Lee J, et al. Randomized multicenter trial of foscarnet versus ganciclovir for preemptive therapy of cytomegalovirus infection after allogeneic stem cell transplantation. *Blood*. 2002;99(4):1159-1164. <https://doi.org/10.1182/blood.v99.4.1159>
 65. Roupael NG, Hurwitz SJ, Hart M, et al. Phase Ib trial to evaluate the safety and pharmacokinetics of multiple ascending doses of filociclovir (MBX-400, cyclopropavir) in healthy volunteers. *Antimicrob Agents Chemother*. 2019;63(9). <https://doi.org/10.1128/AAC.00717-19>
 66. Gergen J, Coulon F, Creneguy A, et al. Multiplex CRISPR/Cas9 system impairs HCMV replication by excising an essential viral gene. *PLoS One*. 2018;13(2):e0192602. <https://doi.org/10.1371/journal.pone.0192602>
 67. Chen S-J, Chen Y-C. Potential application of TALENs against murine cytomegalovirus latent infections. *Viruses*. 2019;11(5):414. <https://doi.org/10.3390/v11050414>
 68. King MW, Munger J. Editing the human cytomegalovirus genome with the CRISPR/Cas9 system. *Virology*. 2019;529:186-194. <https://doi.org/10.1016/j.virol.2019.01.021>
 69. Zhang X, Song Z, Qin B, et al. Rupintrivir is a promising candidate for treating severe cases of enterovirus-71 infection: evaluation of antiviral efficacy in a murine infection model. *Antivir Res*. 2013;97(3):264-269. <https://doi.org/10.1016/j.antiviral.2012.12.029>
 70. Baggen J, Thibaut HJ, Strating JRPM, van Kuppeveld FJM. The life cycle of non-polio enteroviruses and how to target it. *Nat Rev Microbiol*. 2018;16(6):368-381. <https://doi.org/10.1038/s41579-018-0005-4>
 71. Abzug MJ, Michaels MG, Wald E, et al. A randomized, double-blind, placebo-controlled trial of pleconaril for the treatment of neonates with enterovirus sepsis. *J Pediatric Infect Dis Soc*. 2016;5(1):53-62. <https://doi.org/10.1093/jpids/piv015>
 72. Zhang M, Wang Y, He W, et al. Design, synthesis, and evaluation of novel enterovirus 71 inhibitors as therapeutic drug leads for the treatment of human hand, foot, and mouth disease. *J Med Chem*. 2020;63(3):1233-1244. <https://doi.org/10.1021/acs.jmedchem.9b01414>
 73. Wittekind SG, Allen CC, Jefferies JL, et al. Neonatal enterovirus myocarditis with severe dystrophic calcification: novel treatment with pocapavir. *J Investig Med High Impact Case Rep*. 2017;5(3):2324709617729393. <https://doi.org/10.1177/2324709617729393>
 74. Amdani SM, Kim HS, Orvedahl A, John AO, Said A, Simpson K. Successful treatment of fulminant neonatal enteroviral myocarditis in monochorionic diamniotic twins with cardiopulmonary support, intravenous immunoglobulin and pocapavir. *Case Rep*. 2018;2018:bcr-2017-224133. <https://doi.org/10.1136/bcr-2017-224133>
 75. Study to evaluate the tolerance and pharmacokinetics of suramin sodium - full text view - ClinicalTrials.gov. Accessed December 13, 2020. <https://clinicaltrials.gov/ct2/show/NCT03804749>
 76. Ren P, Zou G, Bailly B, et al. The approved pediatric drug suramin identified as a clinical candidate for the treatment of EV71 infection—suramin inhibits EV71 infection in vitro and in vivo. *Emerg Microb Infect*. 2014;3(1):e62-e69. <https://doi.org/10.1038/emi.2014.60>
 77. Manganaro R, Zonsics B, Bauer L, et al. Synthesis and antiviral effect of novel fluoxetine analogues as enterovirus 2C inhibitors. *Antivir Res*. 2020;178:104781. <https://doi.org/10.1016/j.antiviral.2020.104781>
 78. Du R, Mao Q, Hu Y, et al. A potential therapeutic neutralization monoclonal antibody specifically against multi-coxsackievirus A16 strains challenge. *Hum Vaccines Immunother*. 2019;15(10):2343-2350. <https://doi.org/10.1080/21645515.2019.1565266>
 79. Zhou B, Xu L, Zhu R, et al. A bispecific broadly neutralizing antibody against enterovirus 71 and coxsackievirus A16 with therapeutic potential. *Antivir Res*. 2019;161:28-35. <https://doi.org/10.1016/j.antiviral.2018.11.001>
 80. Strating JRPM, van der Linden L, Albulescu L, et al. Itraconazole inhibits enterovirus replication by targeting the oxysterol-binding protein. *Cell Rep*. 2015;10(4):600-615. <https://doi.org/10.1016/j.celrep.2014.12.054>
 81. Shakeel S, Westerhuis BM, Ora A, et al. Structural basis of human parechovirus neutralization by human monoclonal antibodies. *J Virol*. 2015;89(18):9571-9580. <https://doi.org/10.1128/JVI.01429-15>
 82. Wald A, Timmler B, Magaret A, et al. Effect of pritelivir compared with valacyclovir on genital HSV-2 shedding in patients with frequent recurrences: a randomized clinical trial. *JAMA*. 2016;316(23):2495-2503. <https://doi.org/10.1001/jama.2016.18189>
 83. AiCuris Anti-infective Cures GmbH. A randomized, open label, multi-center, comparative trial, to assess the efficacy and safety of pritelivir versus foscarnet for the treatment of acyclovir-resistant mucocutaneous HSV (herpes simplex virus) infections in immunocompromised adults (PRIOH-1). clinicaltrials.gov. 2020. Accessed December 30, 2020. <https://clinicaltrials.gov/ct2/show/NCT03073967>
 84. Tyring S, Wald A, Zadeikis N, Dhadda S, Takenouchi K, Rorig R. ASP2151 for the treatment of genital herpes: a randomized, double-blind, placebo- and valacyclovir-controlled, dose-finding study. *J Infect Dis*. 2012;205(7):1100-1110. <https://doi.org/10.1093/infdis/jis019>
 85. Kawashima M, Nemoto O, Honda M, et al. Amenamevir, a novel helicase-primase inhibitor, for treatment of herpes zoster: a randomized, double-blind, valacyclovir-controlled phase 3 study. *J Dermatol*. 2017;44(11):1219-1227. <https://doi.org/10.1111/1346-8138.13948>
 86. Roehm PC, Shekarabi M, Wollebo HS, et al. Inhibition of HSV-1 replication by gene editing strategy. *Sci Rep*. 2016;6(1):23146. <https://doi.org/10.1038/srep23146>
 87. Paavilainen H, Romanovskaya A, Nygårdas M, Bamford DH, Poranen MM, Hukkanen V. Innate responses to small interfering RNA pools inhibiting herpes simplex virus infection in astrocytoid and epithelial cells. *Innate Immun*. 2014;21(4):349-357. <https://doi.org/10.1177/1753425914537921>
 88. Abzug MJ, Cloud G, Bradley J, et al. Double blind placebo-controlled trial of pleconaril in infants with enterovirus

- meningitis. *Pediatr Infect Dis J*. 2003;22(4):335-341. <https://doi.org/10.1097/01.inf.0000059765.92623.70>
89. Torres-Torres S, Myers AL, Klatte JM, et al. First use of investigational antiviral drug pocapavir (V-073) for treating neonatal enteroviral sepsis. *Pediatric Infect Dis J*. 2015;34(1):52-54. <https://doi.org/10.1097/INF.0000000000000497>
 90. Benkahla MA, Alidjinou EK, Sane F, Desailoud R, Hober D. Fluoxetine can inhibit coxsackievirus-B4 E2 in vitro and in vivo. *Antivir Res*. 2018;159:130-133. <https://doi.org/10.1016/j.antiviral.2018.10.002>
 91. Messacar K, Sillau S, Hopkins SE, et al. Safety, tolerability, and efficacy of fluoxetine as an antiviral for acute flaccid myelitis. *Neurology*. 2019;92:e2118-e2126. <https://doi.org/10.1212/WNL.0000000000006670>
 92. Immunization in the Asia-Pacific region - ScienceDirect. Accessed June 18, 2022. <https://www.sciencedirect.com/science/article/pii/B978145570095000690>
 93. Li Q. The safety, immune persistence and consistency of inactivated enterovirus 71 vaccine (human diploid cell, KMB-17). clinicaltrials.gov. 2018. Accessed June 22, 2022. <https://clinicaltrials.gov/ct2/show/NCT02999828>
 94. Gu W, Zeng G, Hu Y-M, et al. A comparative analysis of immunogenicity and safety of an enterovirus 71 vaccine between children aged 3-5 years and infants aged 6-35 months. *Expert Rev Vaccines*. 2018;17(3):257-262. <https://doi.org/10.1080/14760584.2018.1430572>
 95. HK inno.N Corporation. A randomized, double-blind, placebo-controlled, phase 1 clinical trial to investigate the safety and immunogenicity of high-dose IN-B001 after administration in healthy subjects. clinicaltrials.gov. 2020. Accessed December 10, 2020. <https://clinicaltrials.gov/ct2/show/NCT04637919>
 96. Tambyah PA, Oon J, Asli R, et al. An inactivated enterovirus 71 vaccine is safe and immunogenic in healthy adults: a phase I, double blind, randomized, placebo-controlled, study of two dosages. *Vaccine*. 2019;37(31):4344-4353. <https://doi.org/10.1016/j.vaccine.2019.06.023>
 97. Li R, Liu L, Mo Z, et al. An inactivated enterovirus 71 vaccine in healthy children. *N Engl J Med*. 2014;370(9):829-837. <https://doi.org/10.1056/NEJMoa1303224>
 98. Heinimäki S, Hankaniemi MM, Siiofy-Khojine A-B, et al. Combination of three virus-derived nanoparticles as a vaccine against enteric pathogens; enterovirus, norovirus and rotavirus. *Vaccine*. 2019;37(51):7509-7518. <https://doi.org/10.1016/j.vaccine.2019.09.072>
 99. Wu Y, Zhu R, Xu L, et al. A novel combined vaccine based on monochimeric VLP co-displaying multiple conserved epitopes against enterovirus 71 and varicella-zoster virus. *Vaccine*. 2017;35(20):2728-2735. <https://doi.org/10.1016/j.vaccine.2017.03.065>
 100. Britton PN, Jones CA, Macartney K, Cheng AC. Parechovirus: an important emerging infection in young infants. *Med J Aust*. 2018;208(8):365-369. <https://doi.org/10.5694/mja18.00149>
 101. Chamings A, Druce J, Caly L, et al. Evolutionary analysis of human parechovirus type 3 and clinical outcomes of infection during the 2017-18 Australian epidemic. *Sci Rep*. 2019;9(1):8906. <https://doi.org/10.1038/s41598-019-45445-z>
 102. Kadambari S, Harvala H, Simmonds P, Pollard AJ, Sadarangani M. Strategies to improve detection and management of human parechovirus infection in young infants. *Lancet Infect Dis*. 2019;19(2):e51-e58. [https://doi.org/10.1016/S1473-3099\(18\)30288-3](https://doi.org/10.1016/S1473-3099(18)30288-3)
 103. Sedmak G, Nix WA, Jentzen J, et al. Infant deaths associated with human parechovirus infection in Wisconsin. *Clin Infect Dis*. 2010;50(3):357-361. <https://doi.org/10.1086/649863>
 104. Britton PN, Dale RC, Nissen MD, et al. Parechovirus encephalitis and neurodevelopmental outcomes. *Pediatrics*. 2016;137(2). <https://doi.org/10.1542/peds.2015-2848>
 105. Hudson JA, Broad J, Martin NG, et al. Outcomes beyond hospital discharge in infants and children with viral meningitis: a systematic review. *Rev Med Virol*. 2020;30(2):e2083. <https://doi.org/10.1002/rmv.2083>
 106. van Hinsbergh TMT, Elbers RG, van Furth MAM, Obihara CCC. Longitudinal association between human parechovirus central nervous system infection and gross-motor neurodevelopment in young children. *Pediatr Infect Dis J*. 2019;38(2):110-114. <https://doi.org/10.1097/INF.0000000000002052>
 107. Looker KJ, Magaret AS, May MT, et al. First estimates of the global and regional incidence of neonatal herpes infection. *Lancet Global Health*. 2017;5(3):e300-e309. [https://doi.org/10.1016/S2214-109X\(16\)30362-X](https://doi.org/10.1016/S2214-109X(16)30362-X)
 108. Pinninti SG, Kimberlin DW. Management of neonatal herpes simplex virus infection and exposure. *Arch Dis Child Fetal Neonatal Ed*. 2014;99(3):F240-F244. <https://doi.org/10.1136/archdischild-2013-303762>
 109. Fernandes ND, Arya K, Ward R. Congenital herpes simplex. In: *StatPearls*. StatPearls Publishing; 2020.
 110. Xu F, Markowitz LE, Gottlieb SL, Berman SM. Seroprevalence of herpes simplex virus types 1 and 2 in pregnant women in the United States. *Am J Obstet Gynecol*. 2007;196:43.e1-43.e436. <https://doi.org/10.1016/j.ajog.2006.07.051>
 111. Warnecke JM, Pollmann M, Borchardt-Lohölter V, et al. Seroprevalences of antibodies against ToRCH infectious pathogens in women of childbearing age residing in Brazil, Mexico, Germany, Poland, Turkey and China. *Epidemiol Infect*. 2020;148:e271. <https://doi.org/10.1017/S0950268820002629>
 112. Ju N, Sanders EJ, Ngetsa C, et al. Seroprevalence, predictors and estimated incidence of maternal and neonatal herpes simplex virus Type 2 infection in semi-urban women in Kilifi, Kenya. *BMC Infect Dis*. 2011;11(1):155. <https://doi.org/10.1186/1471-2334-11-155>
 113. Tooke P, Peckham CS. Neonatal herpes simplex virus infection in the British Isles. *Paediatr Perinat Epidemiol*. 1996;10(4):432-442. <https://doi.org/10.1111/j.1365-3016.1996.tb00070.x>
 114. Kadambari S, Okike I, Ribeiro S, et al. Seven-fold increase in viral meningo-encephalitis reports in England and Wales during 2004-2013. *J Infect*. 2014;69(4):326-332. <https://doi.org/10.1016/j.jinf.2014.05.012>
 115. *Sexually Transmitted Infections (STIs): Annual Data Tables*. GOV.UK. Accessed February 23, 2021. <https://www.gov.uk/government/statistics/sexually-transmitted-infections-stis-annual-data-tables>
 116. Samies N. Evaluation of the pharmacokinetics and pharmacodynamics of valacyclovir in neonates with neonatal herpes simplex virus disease who have completed standard of care treatment with acyclovir. clinicaltrials.gov. 2020. Accessed December 21, 2020. <https://clinicaltrials.gov/ct2/show/NCT04448392>
 117. Kimberlin DW, Baley J, Brady MT, et al. Guidance on management of asymptomatic neonates born to women with active genital herpes lesions. *Pediatrics*. 2013;131(2):e635-e646. <https://doi.org/10.1542/peds.2012-3216>
 118. BASHH Guidelines. Accessed January 16, 2021. <https://www.bashhguidelines.org/current-guidelines/genital-ulceration/anogenital-herpes-2014/>
 119. Wang Y, Smith KP. Safety of alternative antiviral agents for neonatal herpes simplex virus encephalitis and disseminated infection. *J Pediatr Pharmacol Therapeut*. 2014;19(2):72-82. <https://doi.org/10.5863/1551-6776-19-2-72>
 120. El-Haddad D, El Chaer F, Vanichanan J, et al. Brincidofovir (CMX-001) for refractory and resistant CMV and HSV infections in immunocompromised cancer patients: a single-center experience. *Antivir Res*. 2016;134:58-62. <https://doi.org/10.1016/j.antiviral.2016.08.024>
 121. Voigt S, Hofmann J, Edelmann A, Sauerbrei A, Kuhl JS. Brincidofovir clearance of acyclovir-resistant herpes simplex virus-1 and

- adenovirus infection after stem cell transplantation. *Transpl Infect Dis.* 2016;18(5):791-794. <https://doi.org/10.1111/tid.12582>
122. Lee YJ, Neofytos D, Kim SJ, et al. Efficacy of brincidofovir as prophylaxis against HSV and VZV in hematopoietic cell transplant recipients. *Transpl Infect Dis.* 2018;20(6):e12977. <https://doi.org/10.1111/tid.12977>
 123. Majewska A, Mlynarczyk-Bonikowska B. 40 Years after the registration of acyclovir: do we need new anti-herpetic drugs? *Int J Mol Sci.* 2022;23(7):3431. <https://doi.org/10.3390/ijms23073431>
 124. Xu X, Zhang Y, Li Q. Characteristics of herpes simplex virus infection and pathogenesis suggest a strategy for vaccine development. *Rev Med Virol.* 2019;29(4):e2054. <https://doi.org/10.1002/rmv.2054>
 125. Belshe RB, Leone PA, Bernstein DI, et al. Efficacy results of a trial of a herpes simplex vaccine. *N Engl J Med.* 2012;366(1):34-43. <https://doi.org/10.1056/NEJMoa1103151>
 126. Kim HC, Oh DS, Park JH, et al. Multivalent DNA vaccine protects against genital herpes by T-cell immune induction in vaginal mucosa. *Antivir Res.* 2020;177:104755. <https://doi.org/10.1016/j.antiviral.2020.104755>
 127. Liu W, Zhou Y, Wang Z, et al. Evaluation of recombinant adenovirus vaccines based on glycoprotein D and truncated UL25 against herpes simplex virus type 2 in mice. *Microbiol Immunol.* 2017; 61(5):176-184. <https://doi.org/10.1111/1348-0421.12482>
 128. Stanfield BA, Pahar B, Chouljenko VN, Veazey R, Kousoulas KG. Vaccination of rhesus macaques with the live-attenuated HSV-1 vaccine VC2 stimulates the proliferation of mucosal T cells and germinal center responses resulting in sustained production of highly neutralizing antibodies. *Vaccine.* 2017;35(4):536-543. <https://doi.org/10.1016/j.vaccine.2016.12.018>
 129. Bernstein DI, Pullum DA, Cardin RD, Bravo FJ, Dixon DA, Kousoulas KG. The HSV-1 live attenuated VC2 vaccine provides protection against HSV-2 genital infection in the guinea pig model of genital herpes. *Vaccine.* 2019;37(1):61-68. <https://doi.org/10.1016/j.vaccine.2018.11.042>
 130. An HSV-2 trivalent vaccine is immunogenic in rhesus macaques and highly efficacious in Guinea pigs. Accessed November 18, 2020. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5245903/>
 131. Stoyanova A, Nikolova I, Pürstinger G, et al. Anti-enteroviral triple combination of viral replication inhibitors: activity against coxsackievirus B1 neuroinfection in mice. *Antivir Chem Chemother.* 2015; 24(5-6):136-147. <https://doi.org/10.1177/2040206616671571>
 132. The promise of mRNA vaccines: a biotech and industrial perspective | npj Vaccines. Accessed June 24, 2022. <https://www.nature.com/articles/s41541-020-0159-8>

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