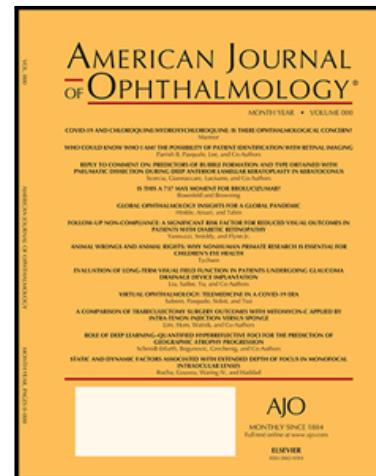


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**Longitudinal Changes in Scotopic and Mesopic Macular Function as Assessed with Microperimetry in Patients with Stargardt Disease: SMART Study Report No. 2**

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Short Title:

Scotopic and Mesopic Macular Functions in Stargardt Disease

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**Abstract**

**Purpose:** To estimate and compare cross-sectional scotopic versus mesopic macular sensitivity losses measured by microperimetry, and to report and compare the longitudinal rates of scotopic and mesopic macular sensitivity losses in *ABCA4* gene associated Stargardt Disease (STGD1).

**Design:** Multicenter prospective cohort study.

**Methods:** Participants: 127 molecular confirmed STGD1 patients enrolled from 6 centers in the USA and Europe and followed every 6 months for up to 2 years.

Observation Procedures: The Nidek MP-1S device was used to measure macular sensitivities of the central 20° under mesopic and scotopic conditions. The mean deviations (MD) from normal for mesopic macular sensitivity for the fovea (within 2° eccentricity) and extrafovea (4°-10° eccentricity), and the MD for scotopic sensitivity for the extrafovea were calculated. Linear mixed effects models were used to estimate mesopic and scotopic changes.

Main Outcome Measures: Baseline mesopic mean deviation (mMD) and scotopic MD (sMD) and rates of longitudinal changes in the mMDs and sMD.

**Results:** At baseline, all eyes had larger sMD, and the difference between extrafoveal sMD and mMD was 10.7 dB ( $p<.001$ ). Longitudinally, all eyes showed a statistically significant worsening trend: the rates of foveal mMD and extrafoveal mMD and sMD changes were 0.72 (95%CI: 0.37 to 1.07), 0.86 (95%CI: 0.58 to 1.14) and 1.12 (95%CI: 0.66 to 1.57) dB/year, respectively.

**Conclusions:** In STGD1, in extrafovea, loss of scotopic macular function preceded and was faster than the loss of mesopic macular function. Scotopic and mesopic

macular sensitivities using microperimetry provide alternative visual function outcomes for STGD1 treatment trials.

### ***Introduction***

Stargardt disease type 1 (STGD1) is the most prevalent juvenile macular dystrophy with an estimated population prevalence of 1/6,500 in the US, and can affect both children and adults<sup>1-4</sup>. It is caused by variants in the *ABCA4* gene, and primarily inherited as an autosomal recessive trait<sup>3</sup>. The disease exhibits great genotypic and phenotypic heterogeneity, but the typical pathology involves atrophy of the para-foveal and/or foveal regions of the macula, with expansion to the peripheral retina over time<sup>3</sup>. Clinically, patients experience a gradual loss of central vision, and may reach legal blindness over decades<sup>5,6</sup>.

The *ABCA4* gene is expressed in both rod and cone photoreceptors<sup>7-9</sup>, and both cone and rod functions are affected in STGD1 as a consequence of pathogenic variants in *ABCA4*. A study at the photoreceptor level using Adaptive Optics Scanning Light Ophthalmoscopy (AOSLO) technology suggested that at the early stage loss of cone photoreceptors focused in the fovea whereas peripheral loss focused on rod photoreceptors<sup>10</sup>. Understanding clinically the temporal and spatial loss of different photoreceptor types will facilitate a better understanding of disease pathophysiology and can inform choices of clinical measurements based on the specific pathways targeted by therapeutic interventions.

Microperimetry (MP), an automated fundus perimetry with live eye tracking, has become a useful clinical tool to monitor retinal disease progression <sup>11</sup>. The quantitative

macular sensitivity measurements generated from MP also provide potential visual function outcome measures for STGD1 treatment trials. A few studies have reported macular sensitivity loss in STGD1 under mesopic conditions using cross-sectional and longitudinal MP data<sup>[12-19](#)</sup>, but there is little available data describing scotopic macular function loss in STGD1<sup>[20 21](#)</sup>.

The prospective “Progression of Atrophy Secondary to Stargardt Disease” (ProgStar) study (ClinicalTrials.gov ID: NCT01977846) is a multicenter natural history study designed to assess visual function and retinal morphological changes in STGD1 and to help identify appropriate outcome measures for STGD1 treatment trials<sup>[22](#)</sup>. Its ancillary study, the “Scotopic Microperimetric Assessment of Rod Function in Stargardt Disease” (SMART) study, focused on assessing macular function loss under scotopic conditions in STGD1<sup>[23](#)</sup>.

To better understand macular function loss in STGD1 natural history and to assess the potential of macular sensitivity measurements as primary outcome measures for STGD1 trials, the current analysis used the longitudinal data of ProgStar participants enrolled in the SMART study and aimed to compare cross-sectional scotopic versus mesopic macular sensitivity loss, to report the longitudinal rate of scotopic macular sensitivity loss and to compare it to the rate of mesopic macular sensitivity loss.

### **Methods**

The study designs of the ProgStar and the SMART studies were reported previously<sup>[22 23](#)</sup>. In brief, the international ProgStar study enrolled 259 STGD1 patients from 9 clinical centers in the US, United Kingdom, France and Germany during 2014-

15. Eligibility of participants and inclusion criteria of study eyes particularly relevant here included: age $\geq$  6 years and having 2 pathogenic mutations in the ABCA4 gene, or having 1 pathogenic mutation in the ABCA4 gene together with a typical STGD1 phenotype; and eyes having a best corrected visual acuity (BCVA) of 20 or more Early Treatment Diabetic Retinopathy Study (ETDRS) letters (i.e., 20/400 Snellen equivalent or better), and having at least 1 well-demarcated area of atrophy on fundus autofluorescence (FAF) imaging with a diameter of 300  $\mu$ m or more and the sum of all lesions of 12 mm<sup>2</sup> or less (further inclusion/exclusion criteria are described in ProgStar report no. 1 [22](#)). All participants provided written informed consent before enrollment in the study. ProgStar participants were followed every 6 months for 2 years. At each study visit, participants underwent a detailed ophthalmic exam, BCVA testing using the ETDRS protocol [24](#), and mesopic MP testing. FAF and spectral domain ocular coherence tomography (SD-OCT) images were also obtained.

After the ProgStar study started enrollment, the SMART study was initiated and 6 ProgStar sites participated. Informed consents were obtained for ProgStar participants to undergo scotopic MP testing during their remaining ProgStar visits. Only one eye was selected to undergo scotopic testing. The selection was determined by the site principal investigators and they were recommended by the study protocol to select the eye with the smaller lesion from FAF imaging, better BCVA, and/or better fixation[23](#). Mesopic and scotopic MP tests were repeated at each subsequent available ProgStar follow-up visit using the follow-up function of the MP device.

Nidek MP-1S (Nidek Technologies, Inc., Gamagori, Japan) and the associated NAVIS Software (v.1.7.7 or higher) (Nidek Technologies S.R.L.) were used for both

mesopic and scotopic MP testing. Procedures for the mesopic testing and fixation assessment were described previously [19<sup>25</sup>](#). Briefly, under dim room lighting, mesopic macular sensitivity was tested at 68 retinal locations using a pattern comparable to the Humphrey 10-2 protocol (**Error! Reference source not found.** Left). A white stimulus (0.43 degree diameter; comparable to Goldmann III) was used with a duration of 200 ms and on a dim white background (1.27 candela (cd)/m<sup>2</sup>). Both cones and rods should contribute to the sensitivity response under such testing conditions. The maximum stimulus luminance in MP-1S was 127 cd/m<sup>2</sup> [26](#).

Under the SMART study protocol, testing under scotopic conditions was added where macular response was predominantly driven by rod photoreceptors in normal eyes. The scotopic testing was performed in a fully darkened room. Sensitivity was tested at 40 retinal locations distributed in a region that extended 4-10 degrees from the foveal center in a custom pattern shown in **Error! Reference source not found.** Right. Before scotopic testing, the eye selected for SMART was occluded for dark adaptation using a double pad and eye patch for at least 30 minutes. To maintain the dark-adapted state, the test background was changed to dim red, and a short-pass (<500 nm, blue) filter (NT52532, Edmund Optics, Barrington, NJ) was inserted into the light path. The dynamic range of the test stimuli was tuned to the typical range of the rod threshold by inserting neutral density (ND) filters. Considering the already impaired visual functions in STGD1 patients, the SMART protocol requested a 1.0-log unit ND filter (1.0ND) which attenuated the luminance of the test stimuli by a factor of 10, although the commonly used 2.0ND filter was also provided. The ND filter used was recorded in the data collection form.

In both the scotopic and the mesopic tests, the SD-OCT image was used to center the test pattern on the anatomical fovea as accurately as possible by the photographer. The sensitivity at each retinal location was determined using the 4-2 threshold strategy. A sensitivity value of 0 dB corresponded to the brightest stimulus applied and a 20 dB value corresponded to the dimmest stimulus. To ensure optimal tracking of the fundus during the testing, participants were instructed to fixate on a fixation target throughout testing (a red cross for the mesopic test and a “white” circle that appeared blue due to the inserted blue filter for the scotopic test). All examinations were performed monocularly with the contralateral eye patched. The mesopic test always preceded the scotopic test for SMART study eyes, and both MP tests always preceded FAF imaging.

The pattern placement on the fovea was categorized as “adequate”, “fair”, “poor”, or “cannot grade” by the study central reading center. When the center of the grid was  $\leq 1^\circ$  from the anatomical fovea this was graded as “adequate”. Distances from  $1^\circ$  to less than  $2^\circ$  were “fair”. “Poor” pattern placement meant that the grid was improperly placed at a distance  $\geq 2^\circ$  and these test results were excluded from subsequent analyses.

#### Macular Sensitivity

The dB output from the MP-1 is a unit relative to the maximum luminance of the MP test which was different between mesopic and scotopic conditions. The thresholds under mesopic and scotopic conditions in normal eyes are also different. Therefore, direct comparison of the numerical output from the above mesopic and scotopic MP tests would not be meaningful. For mesopic response, the dB output from the MP-1 (denoted as  $X_m$ ) was converted to the mean deviation (MD) from the maximum

measurable normal mesopic sensitivity, that is, the mesopic MD was calculated as (20- $X_m$ ) [27](#). For scotopic response, the dB value (denoted as  $X_s$ ) was converted to the MD from the maximum measurable normal scotopic mean sensitivity under the scotopic testing condition using the 2ND filter (i.e. 20dB)[28](#), that is, the scotopic MD was calculated as (20- $X_s$ ) for eyes where the 2ND filter was used and as [20-( $X_s$ -10)] for eyes where the 1ND filter was used.

The scotopic MP test pattern covered the extrafoveal region extending 4°-10° eccentricity. To compare mesopic and scotopic macular function losses in this same region (Figure 1 Left), the mesopic MD (mMD) of the 64 test loci in this region was calculated (Figure 1 Right), and the scotopic MD (sMD) was calculated from the 40 test loci. Fovea mMD was also calculated using the 4 loci within 2° eccentricity from the mesopic MP test.

#### Statistical Analyses

At the first visit for the SMART study, participants' corresponding mesopic MP test results were abstracted from the ProgStar study database. Paired t-test was used to compare the extrafoveal mMD and sMD, and Pearson correlation coefficient was estimated for the extrafoveal mMD and sMD. A linear regression model was also used to assess whether the difference between extrafoveal mMD and sMD was associated with the level of scotopic function loss (i.e. sMD). In addition, the foveal mMD and extrafoveal sMDs were estimated for the subgroups of eyes that had "approximately normal" extrafoveal mesopic sensitivity and of eyes that had "approximately normal" foveal mesopic sensitivity (operationally defined as mesopic mean sensitivity from MP1- $S \geq 12$ dB, i.e. mMD<8 dB [29](#)).

Longitudinally, the rate of change of each MP parameter was estimated using a linear mixed effect model (LMEM), where the mean of the parameter was modeled as a linear function of time since the first visit, with the intercept and slope parameter assumed to be random effects following normal distributions. Such modeling implicitly accounted for the correlation from repeated measurements. To compare between the rates of changes of extrafoveal sMD and mMD, the LMEM was extended to include an indicator of MP test type and its interaction with time. The coefficient of the interaction term quantifies the difference in the rates of changes of extrafoveal MDs between the scotopic MP and mesopic MP tests.

All analyses were conducted in SAS 9.4, and two-sided p-values from Wald-tests were reported. The model fit for LMEMs was inspected visually and based on plots of scaled residuals [30](#).

## **Results**

### Participants Disposition and Demographics

The SMART study enrolled 130 participants (eyes) and scotopic MP was tested for a total of 497 eye-visits. Among these eye-visits, 4 scotopic tests (0.8%) and 6 mesopic MP tests (1.2%) were deemed ungradable by the reading center, and 41 scotopic tests (8.3%) and 60 mesopic tests (12.1%) had poor pattern placement. Among the 442 eye-visits with scotopic MP of adequate or fair pattern placement, 401 eye-visits had mesopic MP tests of adequate or fair pattern placement. These 401 eye-visits were from 127 of the SMART participants (97.7%), and their baseline data (i.e., their first SMART visit) were used to compare between mesopic and scotopic sensitivity impairments cross-sectionally. There were 116 participants for whom data for at least 2

visits were available for the longitudinal data analysis. Seventy-three eyes contributed data at the 6-months follow-up, 89, 83, and 29 eyes contributed data at the 12, 18 and 24-months follow-ups, respectively.

Table 1 summarizes the baseline characteristics for the 127 SMART participants: 52% were female, 83% were white, mean age was 34.5 (standard deviation [SD]=15.1) years, and mean BCVA was 52 ETDRS letters (20/91 Snellen equivalent) (SD=19 letters). The mean age at symptom onset was 24.7 (SD=14.5) years and the mean duration of symptoms was 10.1 years (SD=7.0); with 41.7% of the cohort reporting symptom onset at an age  $\leq$ 18 years (Table 1).

#### Mean Deviations (MD) at the Baseline Visit

At SMART baseline (Table 1), the median mesopic MD (mMD) of the overall central 20° test field was 8.4 dB (interquartile range [IQR] 4.8 to 10.8). The median mMD for the foveal region was 16.8 (IQR 15.0 to 20.0) dB. The median mMD and scotopic MD (sMD) for the extrafoveal region was 7.9 dB (IQR 4.2 to 10.3) and 18.6 dB (IQR 14.9 to 21.8), respectively. Figure 2A shows the cross-sectional distribution of sMD and mMD for the extrafovea. The mean difference between extrofoveal sMD and mMD was 10.7 dB (95%CI 10.1 to 11.3,  $p<.001$ ) (positive [sMD-mMD] value indicates larger scotopic impairment). The correlation coefficient between extrafoveal sMD and mMD was 0.82 ( $p<.001$ ). All eyes had mMD smaller than sMD, indicating larger impairment of scotopic sensitivity in the extrafoveal region. Figure 2B is the scatterplot showing the difference between extrafoveal scotopic and mesopic MDs as a function of extrafoveal sMD: the positive linear trend had a slope of 0.32 (95%CI: 0.20 to 0.43,  $p<.001$ ), suggesting that every 1dB larger scotopic sensitivity loss was significantly

associated with 0.32 dB greater difference between mesopic and scotopic sensitivity impairments.

At baseline, there were 81 eyes that had “approximately normal” extrafoveal mMP ( $MD < 8\text{dB}$ ). Among these eyes, the median foveal mMD was 19.0 (IQR 11.5 to 20; range 0 to 20) dB; and the median extrafoveal sMD was 15.6 (IQR 13.7 to 17.8, and range 0.6 to 29.1) dB (Table 1).

There were 11 eyes that had “approximately normal” foveal mMP (i.e. sensitivity  $\geq 12\text{dB}$ ) (Supplemental table 1). Among these eyes, the median extrafoveal mMD was 1.7 dB (IQR 0.6 to 3.7, range 0 to 9.3); the median extrafoveal sMD was 12.6 dB (IQR 10.4 to 15.6, range 0.6 to 25.7); and the median difference between extrafoveal sMD and mMD was 11.6 dB (IQR 6.7 to 12.6, range 0.5 to 17.6) dB (sMD-mMD positive indicates larger scotopic impairment).

#### Longitudinal Changes in Mean Deviations (MD)

Figure 3 shows the longitudinal data of foveal mMD and extrafoveal mMD and sMD of individual eyes. Large within-eye variability across visits for foveal mMD was observed for some eyes (Figure 3A). Nevertheless, the average of all eyes showed a statistically significant worsening trend and the rate of foveal mMD change was 0.72 (95%CI: 0.37 to 1.07) dB/year (Table 2). The rates of extrafoveal mMD and sMD changes were 0.86 (95%CI: 0.58 to 1.14) dB/year and 1.12 (95%CI: 0.66 to 1.57) dB/year (Figure 3 B-C) (Table 2), respectively.

Comparing between the rates of mesopic and scotopic changes, the model using both extrafoveal mMD and sMD data estimated that the difference in the rate of change

of extrafoveal sMD compared to mMD was 0.41 (95%CI: -0.07 to 0.89) dB/year ( $p=0.10$ ).

For the subgroup of 81 eyes with “approximately normal” extrafoveal mMD at baseline, during follow-up, the rates of foveal mMD and extrafoveal mMD and sMD changes were 0.96 (95%CI: 0.47 to 1.46), 0.96 (95%CI: 0.61 to 1.32) dB/year, and 1.31 (95%CI: 0.74 to 1.89) dB/year, respectively (Table 2).

For the subgroup of 11 eyes with “approximately normal” foveal mMD at baseline, during follow-up, the rates of foveal mMD and extrafoveal mMD and sMD changes were 2.98 (95%CI: -0.21, 6.18) dB/year, 1.20 (95%CI: -0.06 to 2.46) dB/year, and 1.58 (95%CI: -0.84 to 4.00) dB/year, respectively (Table 2). This subgroup also included 2 eyes that had both extrafoveal mesopic and scotopic sensitivity responses that were normal at baseline (MDs~0dB). Their baseline and last follow-up data are shown in Table 3. At baseline Eye 1 had nearly normal foveal mMD (2.0 dB), and normal extrafoveal mMD (0.94 dB) and sMD (1.53 dB). Its BCVA was 20/110 and had a lesion involving the fovea shown in FAF imaging (area of decreased autofluorescence [DAF] = $2.19\text{mm}^2$ ) (Figure 4A left). At the last visit (18 months later), its extrafovea mMD was normal, but its extrafovea sMD declined to 10.2 dB loss and foveal mMD declined to 10 dB associated with the area of DAF slightly increased to  $2.25\text{ mm}^2$  (Figure 4A right). Eye 2 at baseline had normal foveal and extrafoveal mesopic and scotopic sensitivity responses, but the BCVA was 20/126. A DAF of size  $0.57\text{mm}^2$  involved the foveola (Figure 4B left). At the last visit (18 months later) (Figure 4B right), the foveal mMD, extrafoveal mMD and sMD increased to 20 dB, 4.41 dB, and 11.28 dB, respectively. The DAF grew to  $1.59\text{mm}^2$ .

## **Discussion**

Emerging therapeutic options for STGD1 need to be tested in clinical trials.

Identifying appropriate outcome measures that are clinically meaningful and can reflect disease progression in a relatively short time such as 1 or 2 years is important for the design of treatment trials aimed to slow loss of visual functions in STGD1. This led to the ProgStar study the primary aims of which were to estimate the rates of disease progression of retinal structural parameters measured by FAF and SD-OCT and visual functions measured by BCVA and macular sensitivity from microperimetry. The SMART study was further added to estimate scotopic macular function loss and to test the hypothesis that rod function loss was faster than cone loss.

Previous findings of the ProgStar study have repeatedly shown that BCVA decline is slow in STGD1 and the change in BCVA does not show a statistically significant association with growth of area of atrophy measured by FAF.[31-34](#) The ProgStar and SMART study protocols provide natural history data on functional declines in mesopic and scotopic macular sensitivities (MS) over a follow-up of up to 24 months. The MSs were measured as the mean deviations from the maximally measurable normal using the MP-1S device. The rates of MS declines were statistically significant and clinically relevant, thus mesopic and scotopic MSs were more sensitive to reflect functional change than BCVA in STGD1.

Our data suggest that the loss of scotopic macular function is faster than the loss of mesopic macular function in the extrafovea. Thus, sMD may be a more sensitive outcome measure compared to mMD. However, the scotopic MP testing requires dark adaption of at least 30 minutes and thus imposes more testing burden for patients and

clinical staff. Additionally, because sMD had larger variability than mMDs (Table 1), using sMD may not yield a smaller sample size compared to using mMD when designing a clinical trial. Nevertheless, because the loss of scotopic macular function precedes the loss of mesopic macular response in the extrafovea, sMD provides a candidate functional outcome measure that allows trials to enroll earlier staged STGD1 patients where mesopic macular function in the extrafovea has yet to show impairment. For such early stage patients, the foveal mesopic macular function may also provide a sensible functional outcome measure. Scotopic MS also provides a direct functional outcome if the immediate therapeutic target is to preserve rod function.

At the SMART baseline, in all eyes, the scotopic macular function loss was greater than the mesopic function loss in the extrafovea. This suggests that in extrafovea, scotopic function loss could have started prior to mesopic function loss, and/or that loss of scotopic function was faster than the loss of mesopic function. The observation that at baseline, the greater the loss of scotopic function the larger discrepancy between extrafoveal scotopic and mesopic losses (Figure 2B) implies that scotopic function loss must have occurred at a faster speed than the loss of mesopic function. This is further supported by the rates of extrafoveal sMD and mMD changes estimated from the longitudinal data (sMD 1.12 dB/year vs. mMD 0.86 dB/year). Statistically these rates of extrafoveal scotopic and mesopic function losses were not significantly different ( $p=0.10$ ), but the parameter estimates themselves are informative especially in observational studies<sup>35</sup>. The lack of statistical significance could be due to an insufficient sample size. The length of follow-up time ( $\leq 2$  years) was also limited and this length of period may be insufficient to statistically differentiate the trajectories of

scotopic and mesopic function losses given the sample size, especially that only 29 eyes contributed data over 24-months of follow-up.

There is also evidence that scotopic function loss precedes mesopic function loss in the extrafovea. In the subgroup of 81 eyes with approximately normal extrafoveal mesopic response at baseline, concurrently there were already profound losses of scotopic MP response (median sMD=15.6 dB). Moreover, despite such losses at baseline, longitudinally, the rate of scotopic MP function loss was greater than the rate of extrafoveal mesopic MP loss.

STGD1 is predominantly a central disease, thus expectedly only a small proportion of eyes had approximately normal fovea macular function (N=11). This group of eyes, especially those with no or mildly impaired acuity (Supplemental table 1), may include the clinical phenotype of fovea sparing<sup>36</sup>. In this group, there was minimal to no loss of extrafoveal mesopic function (median mMD=1.7 dB), but the concurrent scotopic function loss was much larger (median sMD=12.6 dB). This may suggest that in certain STGD1 genotypes, moderate to large loss in extrafoveal scotopic response may be present while fovea mesopic function remains relatively preserved. Longitudinally, however, this group of eyes in average showed a fast decline in foveal mesopic function. This group also included the 2 eyes that had normal extrafoveal scotopic and mesopic responses at baseline (Table 2). They both had poor BCVA. Over an 18 months period, both eyes had a large loss of foveal mesopic function as well as extrafoveal scotopic function, whereas the extrafoveal mesopic function loss was much smaller.

One of the SMART study's major goals was to test the hypothesis that rod function loss was faster than that of cones. In normal eyes, the extrafoveal scotopic MP response would be primarily mediated by rods, and both rods and cones would mediate sensitivity response under the mesopic condition though the exact contribution from each photoreceptor system is difficult to determine<sup>37</sup>. The rod mediated scotopic response however can be altered in diseased retinas<sup>28</sup>, and the exact contribution due to rod mediation versus cone mediation cannot be deduced without comparison of dark-adapted responses under two colors of perimetry<sup>18:20:38:39</sup>. Therefore, a direct comparison of rod versus cone function loss could not be made with Progstar and SMART data. Nevertheless, the cross-sectional comparisons at baseline showed significantly different levels of deviations of scotopic compared to mesopic MP tests. This suggests that the sensitivity responses under the scotopic and mesopic conditions were mediated by different combinations of photoreceptors.

A previous study of 66 STGD1 patients reported a rate of rod function loss of 1.1 dB/year using two-color dark adapted perimetry and a rate of cone function loss of 0.45 dB/year using light-adapted perimetry with an orange (600 nm) target.<sup>20</sup> Our longitudinal data estimated a rate of sMD change of 1.12dB/year and a rate of mMD change of 0.86 dB/year (Table 2). The sMD rate is comparable to the previously reported 1.1 dB/year of rod function loss, and the mMD rate is in between the previously reported rates of rod and cone function losses. Hence it is reasonable to assume that in the SMART study more rods were mediating the scotopic response and a combination of cones and rods were mediating the mesopic response. It thus can be inferred that, although direct quantitative comparisons of rod and cone function losses cannot not be

made with SMART and ProgStar data, our results for the extrafovea sensitivity responses support the conclusion that rod function loss precedes and declines faster than cone function loss in STGD1. Our results obtained in the fovea and extrafovea also support the observation of “cone loss predominates centrally and rod loss increases peripherally” based on AOSLO imaging of photoreceptors in 2 STGD1 patients<sup>10</sup>.

Strengths of our study include that the data were from a large prospective multi-center international cohort of patients with molecularly confirmed ABCA4 associated STGD1. All testing and data collection was performed under standardized protocols. ProgStar had an independent data coordinating center which assured the data entry and management quality.

There are important limitations of our studies. First, SMART and ProgStar results do not generalize to STGD1 in general because of the ProgStar enrollment criteria. Only eyes with at least 1 well-demarcated area of atrophy with a minimum diameter of 300  $\mu\text{m}$  and the total area of all atrophic lesions  $\leq 12 \text{ mm}^2$  as determined by the clinical examiner were eligible. Thus, the ProgStar sample may have excluded both early stage and late stage STGD1 patients. Additionally, although the rates of mesopic sensitivity loss in the SMART study sample were comparable to the rate of mesopic sensitivity loss during 12-months reported in the ProgStar study sample of 359 eyes from 200 participants <sup>40</sup>, the SMART study sample was a subsample of ProgStar, which may further reduces the generalizability of study findings. Nevertheless, generalizing to the complete ProgStar sample is less meaningful, rather, it is more important to interpret any ProgStar and SMART study finding in consideration of the characteristics of the patients sample specifically used in that analysis. Second, two-color perimetry tests

were not adopted in ProgStar or SMART, thus the exact loss of cone or rod function could not be determined. Third, at the time of our studies, all the Nidek MP-1S devices were running version number 1.7.7 of the NAVIS software, which required manual placement of the test pattern. This led to non-trivial proportions of tests deemed as poor pattern placement by the reading center and their data were not used in the analyses. Recent improved versions of the software allow automatic placement of the test pattern to match the foveal center as marked in an OCT B-scan passing through the foveal center, which should allow better centering of MP and most data being usable. Fourth, the longitudinal analysis was limited by the length of follow-up time ( $\leq 2$  years) which may be insufficient to statistically differentiate the trajectories of scotopic and mesopic function losses, especially most eyes only contributed data over 12 or 18-months of follow-up. Normal aging is associated with faster loss of scotopic versus photopic sensitivity and faster loss of rod- versus cone-photoreceptor phototransduction<sup>41,42</sup>. But the differences are small especially during 1 to 2 years, and thus aging is unlikely to explain the faster loss of scotopic macular function in STGD1 observed here. Fifth, there were limits of detection using the Nidek MP1-S device. The MDs calculated here were the deviations from the maximally measurable normal using the MP-1S device. The mesopic MDs had a range of 0 to 20 dB. Such mMDs may be underestimating the deviations from normals due to the limit of detection for normals from the MP-1S device. The scotopic test used 1ND and 2ND filters which allowed a dynamic range of 0 to 40 dB in sMD. Following the study protocol, most of the scotopic tests used 1ND filter to accommodate the impaired visual function in STGD1 patients. However, for test loci that could afford the normal response of 20dB under the 1ND filter, their values of deviations

were calculated as 10dB because of the limit of detection under the 1ND filter. Of the 386 scotopic tests that used 1ND filter, a quarter of the tests had more than 20% of the test loci that had a normal response of 20dB. Therefore, it was possible that the sMD may be overestimated for eyes that had better scotopic macular function, resulting an over estimation of the difference between scotopic and mesopic function losses. Nevertheless, we conducted a sensitivity analysis to correct for the overestimation by imputing the deviation values to be 0dB for the test loci that had a normal response of 20dB under 1ND filter, and the study conclusions remain unchanged (Supplemental Figure 1 shows the comparison of imputed extrafoveal sMDs to mMDs).

In conclusion, our data suggest that in STGD1, in the extrafovea, scotopic macular function loss preceded mesopic function loss and proceeded at a faster speed. Using sMD as a primary outcome measure may help enroll earlier stage patients and sMD provides a direct functional outcome for testing the efficacy of therapeutics that target on preserving rod photoreceptors. Scotopic and mesopic macular sensitivity measures provide alternative visual function outcomes that are more sensitive to detect change than BCVA for designing future treatment trials aimed to slow progression in STGD1.

#### **Table of Contents Statement**

This multicenter study followed 127 patients with molecularly confirmed *ABCA-4* gene mutation associated Stargardt disease for up to 2 years. The study found that scotopic macular function loss was earlier and faster than mesopic macular function loss. Scotopic and mesopic macular sensitivities using microperimetry provide viable visual function outcomes for treatment trials of *ABCA-4* gene associated Stargardt disease.

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### b. Financial Disclosures

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Hendrik Scholl is member of the Scientific Advisory Board of: Astellas Institute for Regenerative Medicine; Gensight Biologics; Ionis Pharmaceuticals, Inc.; Gyroscope Therapeutics Ltd.; Janssen Research & Development, LLC (Johnson & Johnson); and Pharma Research & Early Development (pRED) of F. Hoffmann-La Roche Ltd; Novartis Pharma AG (CORE). Dr. Scholl is paid consultant of: Boehringer Ingelheim Pharma GmbH & Co; Gerson Lehrman Group; and Guidepoint.

Hendrik Scholl is member of the Data Monitoring and Safety Board/Committee of Belite Bio and ReNeuron Group Plc/Ora Inc. and member of the Steering Committee of Novo Nordisk (FOCUS trial).

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**Appendix: The SMART Study Group Members**

The SMART studies consist of the Chair's Office, Data Coordinating Center, Image Reading Center, and six clinics with the following members:

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## References

1. Michaelides M, Hunt DM, Moore AT. The genetics of inherited macular dystrophies. *J Med Genet.* 2003;40(9):641-650.
2. Stone EM, Andorf JL, Whitmore SS, et al. Clinically Focused Molecular Investigation of 1000 Consecutive Families with Inherited Retinal Disease. *Ophthalmology.* 2017;124(9):1314-1331.
3. Sears AE, Bernstein PS, Cideciyan AV, et al. Towards Treatment of Stargardt Disease: Workshop Organized and Sponsored by the Foundation Fighting Blindness. *Transl Vis Sci Technol.* 2017;6(5):6.
4. Hanany M, Rivolta C, Sharon D. Worldwide carrier frequency and genetic prevalence of autosomal recessive inherited retinal diseases. *Proc Natl Acad Sci U S A.* 2020;117(5):2710-2716.
5. Fishman GA, Farber M, Patel BS, Derlacki DJ. Visual acuity loss in patients with Stargardt's macular dystrophy. *Ophthalmology.* 1987;94(7):809-814.
6. Rotenstreich Y, Fishman GA, Anderson RJ. Visual acuity loss and clinical observations in a large series of patients with Stargardt disease. *Ophthalmology.* 2003;110(6):1151-1158.
7. Allikmets R, Singh N, Sun H, et al. A photoreceptor cell-specific ATP-binding transporter gene (ABCR) is mutated in recessive Stargardt macular dystrophy. *Nat Genet.* 1997;15(3):236-246.
8. Illing M, Molday LL, Molday RS. The 220-kDa rim protein of retinal rod outer segments is a member of the ABC transporter superfamily. *J Biol Chem.* 1997;272(15):10303-10310.
9. Weng J, Mata NL, Azarian SM, Tzekov RT, Birch DG, Travis GH. Insights into the function of Rim protein in photoreceptors and etiology of Stargardt's disease from the phenotype in abcr knockout mice. *Cell.* 1999;98(1):13-23.
10. Song H, Rossi EA, Latchney L, et al. Cone and rod loss in Stargardt disease revealed by adaptive optics scanning light ophthalmoscopy. *JAMA ophthalmology.* 2015;133(10):1198-1203.
11. Nishida Y, Murata T, Yoshida K, Sawada T, Kani K. An automated measuring system for fundus perimetry. *Jpn J Ophthalmol.* 2002;46(6):627-633.
12. Muller PL, Birtel J, Herrmann P, Holz FG, Charbel Issa P, Gliem M. Functional Relevance and Structural Correlates of Near Infrared and Short Wavelength Fundus Autofluorescence Imaging in ABCA4-Related Retinopathy. *Transl Vis Sci Technol.* 2019;8(6):46.
13. Tanna P, Georgiou M, Aboshiha J, et al. Cross-Sectional and Longitudinal Assessment of Retinal Sensitivity in Patients With Childhood-Onset Stargardt Disease. *Transl Vis Sci Technol.* 2018;7(6):10.
14. Anastasakis A, McAnany JJ, Fishman GA, Seiple WH. Clinical value, normative retinal sensitivity values, and intrasession repeatability using a combined spectral domain optical coherence tomography/scanning laser ophthalmoscope microperimeter. *Eye (Lond).* 2011;25(2):245-251.
15. Mori F, Ishiko S, Kitaya N, et al. Scotoma and fixation patterns using scanning laser ophthalmoscope microperimetry in patients with macular dystrophy. *American journal of ophthalmology.* 2001;132(6):897-902.
16. Park SP, Chang S, Allikmets R, et al. Disruption in Bruch membrane in patients with Stargardt disease. *Ophthalmic genetics.* 2012;33(1):49-52.
17. Testa F, Rossi S, Sodi A, et al. Correlation between photoreceptor layer integrity and visual function in patients with Stargardt disease: implications for gene therapy. *Invest Ophthalmol Vis Sci.* 2012;53(8):4409-4415.
18. Cideciyan AV, Swider M, Aleman TS, et al. Macular function in macular degenerations: repeatability of microperimetry as a potential outcome measure for ABCA4-associated retinopathy trials. *Invest Ophthalmol Vis Sci.* 2012;53(2):841-852.
19. Schonbach EM, Wolfson Y, Strauss RW, et al. Macular Sensitivity Measured With Microperimetry in Stargardt Disease in the Progression of Atrophy Secondary to Stargardt Disease (ProgStar) Study: Report No. 7. *JAMA ophthalmology.* 2017;135(7):696-703.
20. Cideciyan AV, Swider M, Aleman TS, et al. ABCA4 disease progression and a proposed strategy for gene therapy. *Hum Mol Genet.* 2009;18(5):931-941.
21. Salvatore S, Fishman GA, McAnany JJ, Genead MA. Association of dark-adapted visual function with retinal structural changes in patients with Stargardt disease. *Retina.* 2014;34(5):989-995.
22. Strauss RW, Ho A, Munoz B, et al. The Natural History of the Progression of Atrophy Secondary to Stargardt Disease (ProgStar) Studies: Design and Baseline Characteristics: ProgStar Report No. 1. *Ophthalmology.* 2016;123(4):817-828.

23. Strauss RW, Kong X, Bittencourt MG, et al. Scotopic Microperimetric Assessment of Rod Function in Stargardt Disease (SMART) Study: Design and Baseline Characteristics (Report No. 1). *Ophthalmic Res.* 2019;61(1):36-43.
24. Early Treatment Diabetic Retinopathy Study design and baseline patient characteristics. ETDRS report number 7. *Ophthalmology.* 1991;98(5 Suppl):741-756.
25. Schönbach EM, Ibrahim MA, Strauss RW, et al. Fixation Location and Stability Using the MP-1 Microperimeter in Stargardt Disease: ProgStar Report No. 3. *Ophthalmology Retina.* 2017;1(1):68-76.
26. 2019 NIDEK Inc. Nidek MP-1S Specifications <https://usa.nidek.com/products/microperimeter-mp-1s/>. Published 2019. Accessed2021.
27. Midena E, Vujosevic S, Cavarzeran F, Microperimetry Study G. Normal values for fundus perimetry with the microperimeter MP1. *Ophthalmology.* 2010;117(8):1571-1576, 1576 e1571.
28. Birch DG, Wen Y, Locke K, Hood DC. Rod sensitivity, cone sensitivity, and photoreceptor layer thickness in retinal degenerative diseases. *Invest Ophthalmol Vis Sci.* 2011;52(10):7141-7147.
29. Chen FK, Patel PJ, Xing W, et al. Test-retest variability of microperimetry using the Nidek MP1 in patients with macular disease. *Invest Ophthalmol Vis Sci.* 2009;50(7):3464-3472.
30. SAS Institute Inc. . *SAS/STAT® 14.2 User's Guide.* Cary, NC: SAS Institute Inc; 2016.
31. Kong X, Strauss RW, Cideciyan AV, et al. Visual Acuity Change over 12 Months in the Prospective Progression of Atrophy Secondary to Stargardt Disease (ProgStar) Study: ProgStar Report Number 6. *Ophthalmology.* 2017;124(11):1640-1651.
32. Kong X, Strauss RW, Michaelides M, et al. Visual Acuity Loss and Associated Risk Factors in the Retrospective Progression of Stargardt Disease Study (ProgStar Report No. 2). *Ophthalmology.* 2016;123(9):1887-1897.
33. Kong X, Fujinami K, Strauss RW, et al. Visual Acuity Change Over 24 Months and Its Association With Foveal Phenotype and Genotype in Individuals With Stargardt Disease: ProgStar Study Report No. 10. *JAMA ophthalmology.* 2018;136(8):920-928.
34. Kong X, West SK, Strauss RW, et al. Progression of Visual Acuity and Fundus Autofluorescence in Recent-Onset Stargardt Disease: ProgStar Study Report #4. *Ophthalmol Retina.* 2017;1(6):514-523.
35. Harrington D, D'Agostino RB, Sr., Gatsonis C, et al. New Guidelines for Statistical Reporting in the Journal. *N Engl J Med.* 2019;381(3):285-286.
36. Bax NM, Valkenburg D, Lambertus S, et al. Foveal Sparing in Central Retinal Dystrophies. *Invest Ophthalmol Vis Sci.* 2019;60(10):3456-3467.
37. Stockman A, Sharpe LT. Into the twilight zone: the complexities of mesopic vision and luminous efficiency. *Ophthalmic Physiol Opt.* 2006;26(3):225-239.
38. Birch D, Fish G. Rod ERGs in Retinitis Pigmentosa and Cone-Rod Degeneration. *Invest Ophthalmol Vis Sci* 1987;28:140-150.
39. Massof RW, Finkelstein D. Two forms of autosomal dominant primary retinitis pigmentosa. *Doc Ophthalmol.* 1981;51(4):289-346.
40. Schonbach EM, Strauss RW, Munoz B, et al. Longitudinal Microperimetric Changes of Macular Sensitivity in Stargardt Disease After 12 Months: ProgStar Report No. 13. *JAMA Ophthalmol.* 2020;138(7):772-779.
41. Jackson GR, Owsley C. Scotopic sensitivity during adulthood. *Vision Res.* 2000;40(18):2467-2473.
42. Cideciyan AV, Jacobson SG. An alternative phototransduction model for human rod and cone ERG a-waves: normal parameters and variation with age. *Vision Res.* 1996;36(16):2609-2621.

## Figure captions

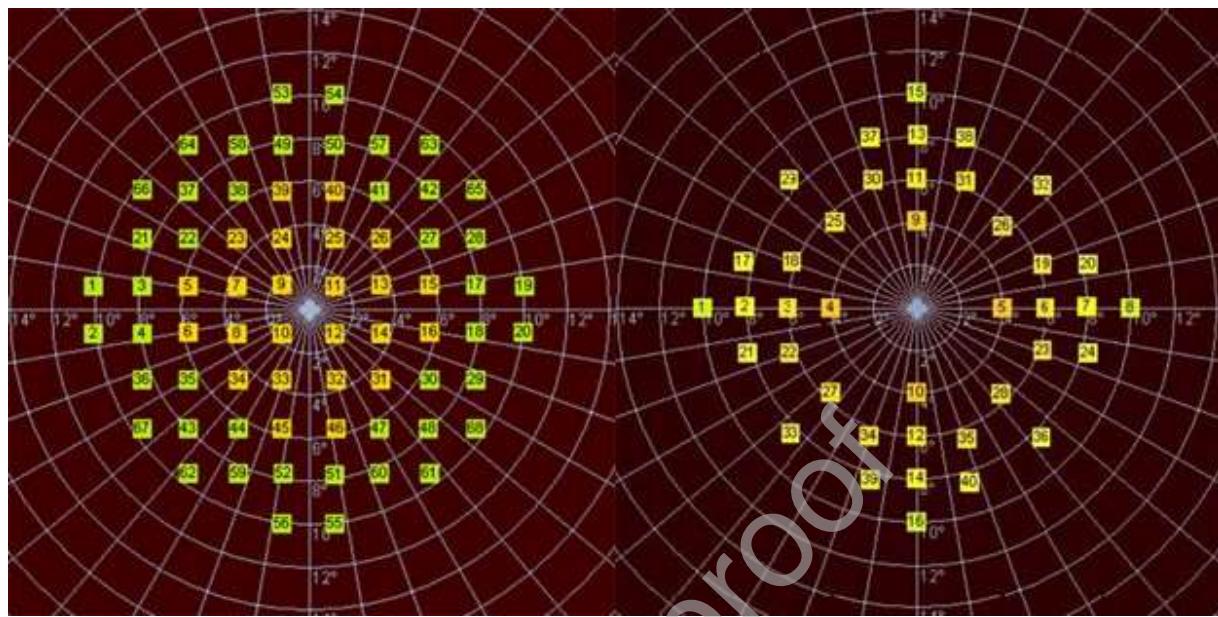


Figure 1. Distributions of microperimetry (MP) test loci in ProgStar and SMART studies. Left: Mesopic MP test pattern including 68 loci from 2° to 10°. Right: Scotopic MP test pattern including 40 loci from 4° to 10°.

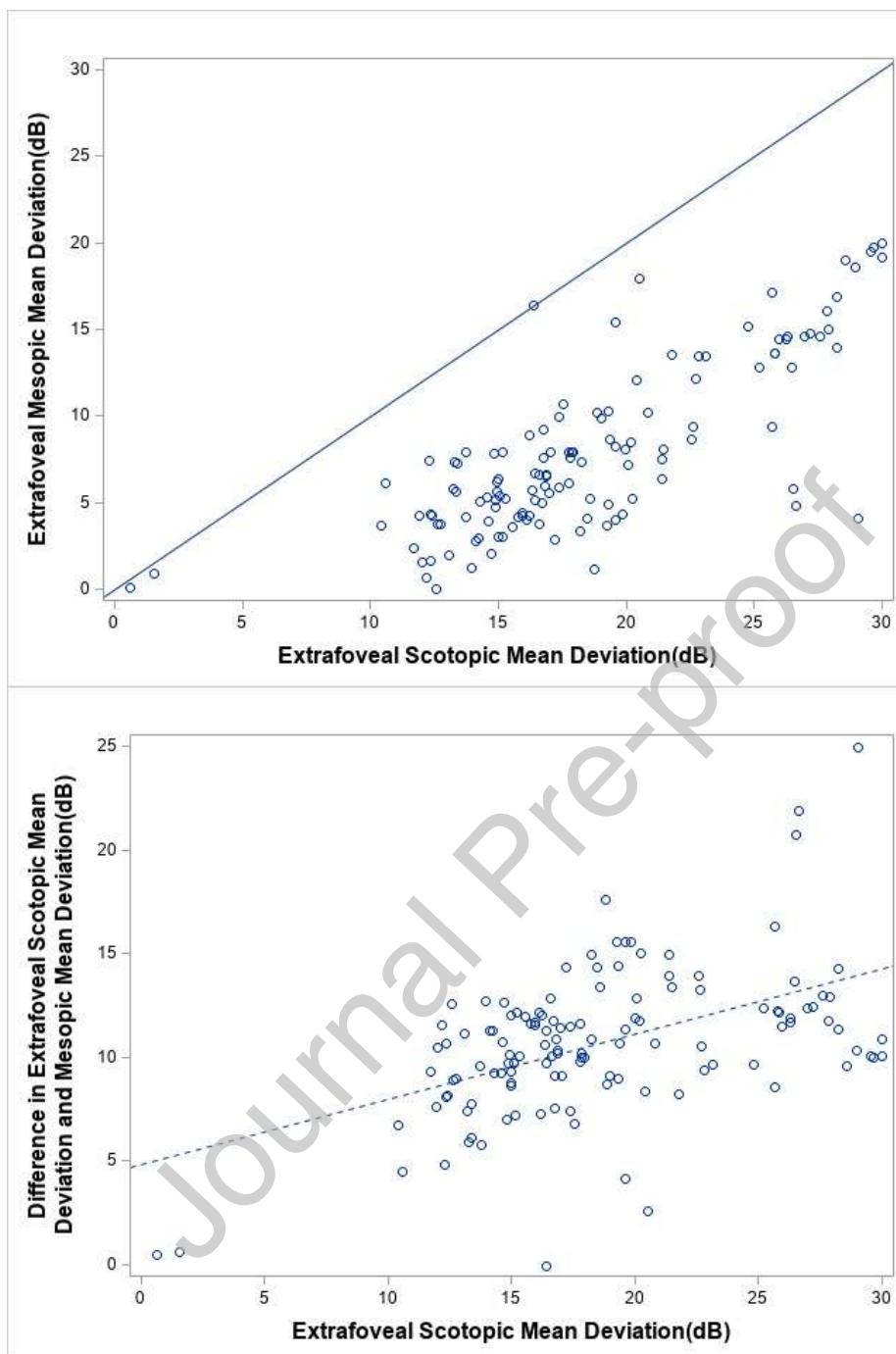
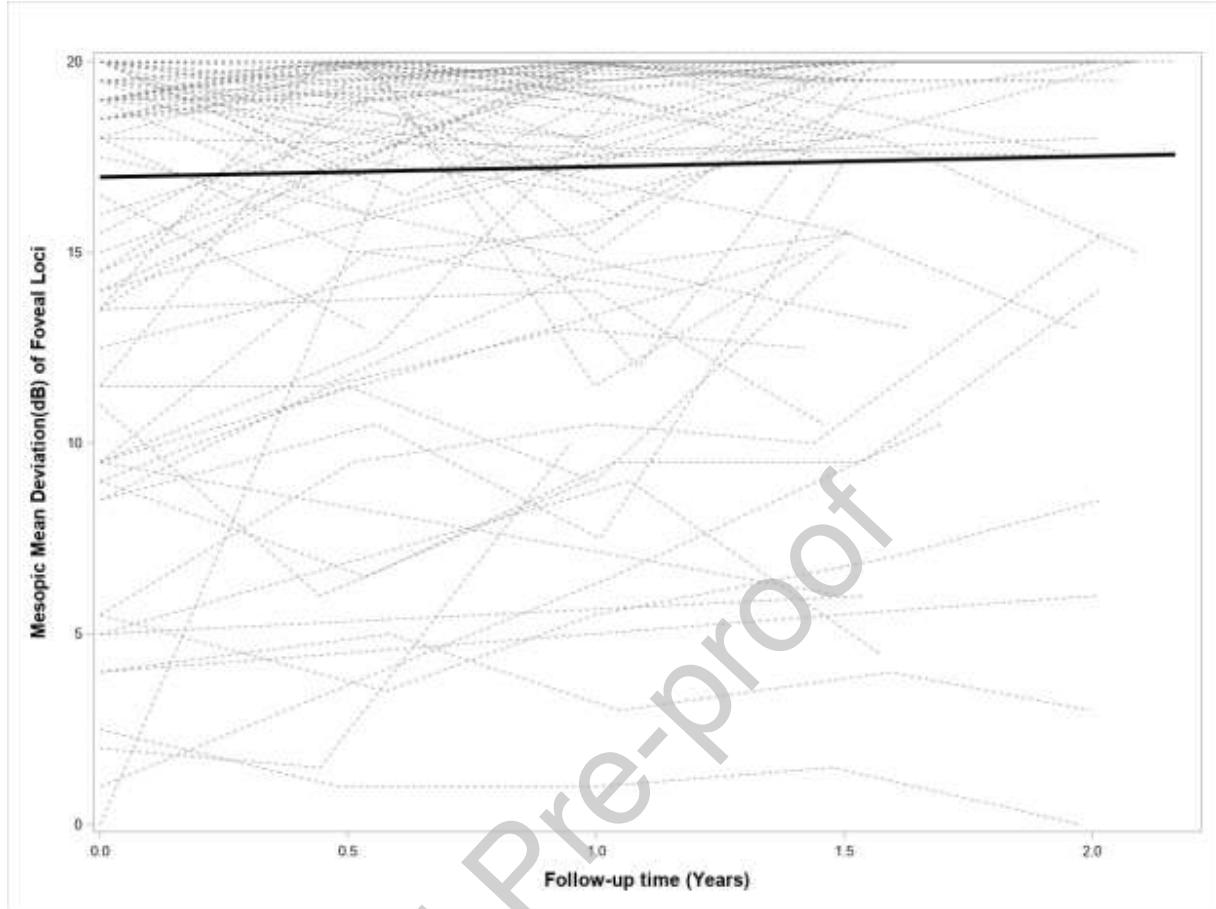


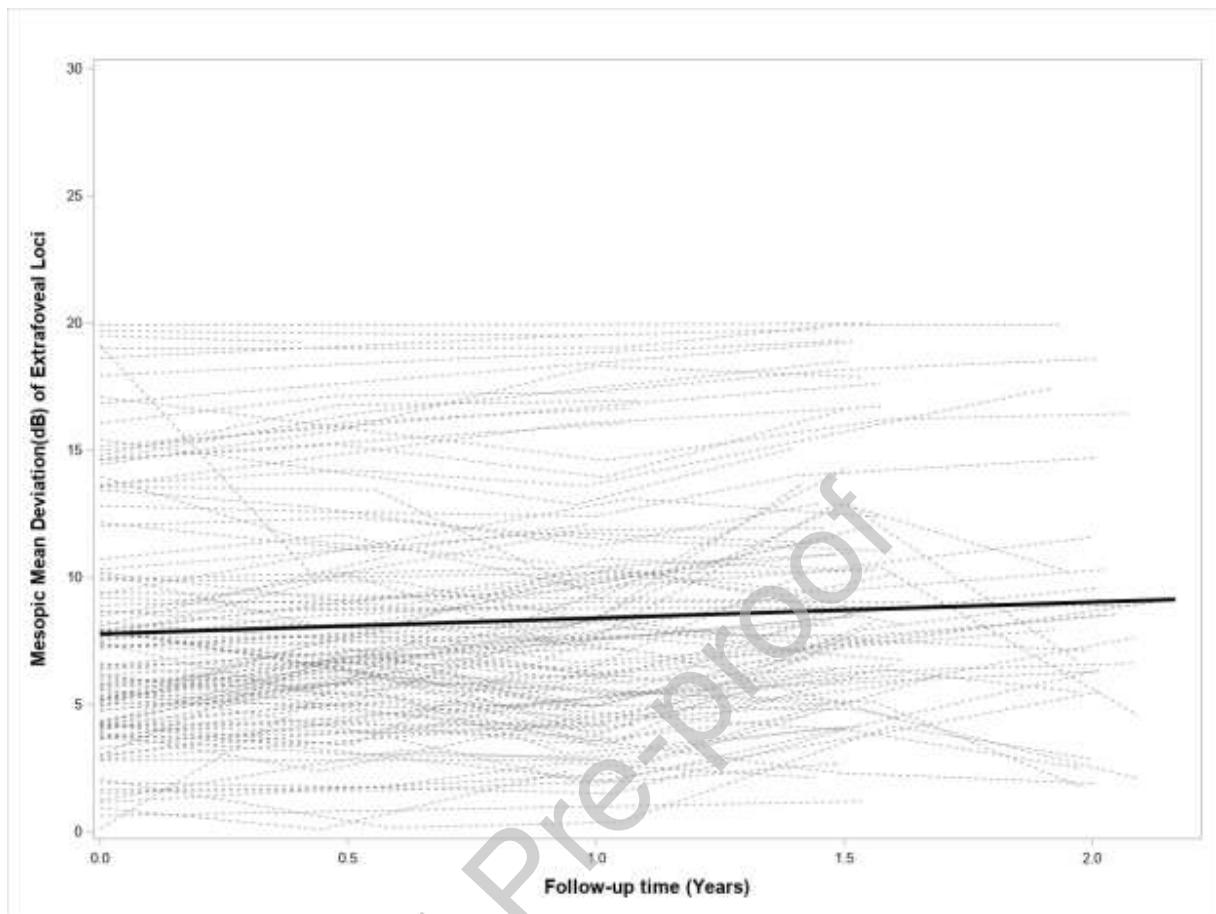
Figure 2. Plots of cross-sectional data at the SMART baseline visit.

A: Scatter plot of the cross-sectional extrafoveal scotopic mean deviation (MD) versus extrafoveal mesopic MD at SMART baseline visit. The line is the diagonal indicating equal x-axis and y-axis values.

B: Scatter plot of the difference between extrafoveal mMD and sMD versus sMD at the baseline visit. The dotted line is the fitted line from the linear regression 0.32 (95%CI: 0.20 to 0.43,  $p < .001$ ). The positive linear trend means that the larger the

scotopic MD, the greater difference between extrafoveal mesopic and scotopic responses and loss of scotopic response was always greater than loss of mesopic response.





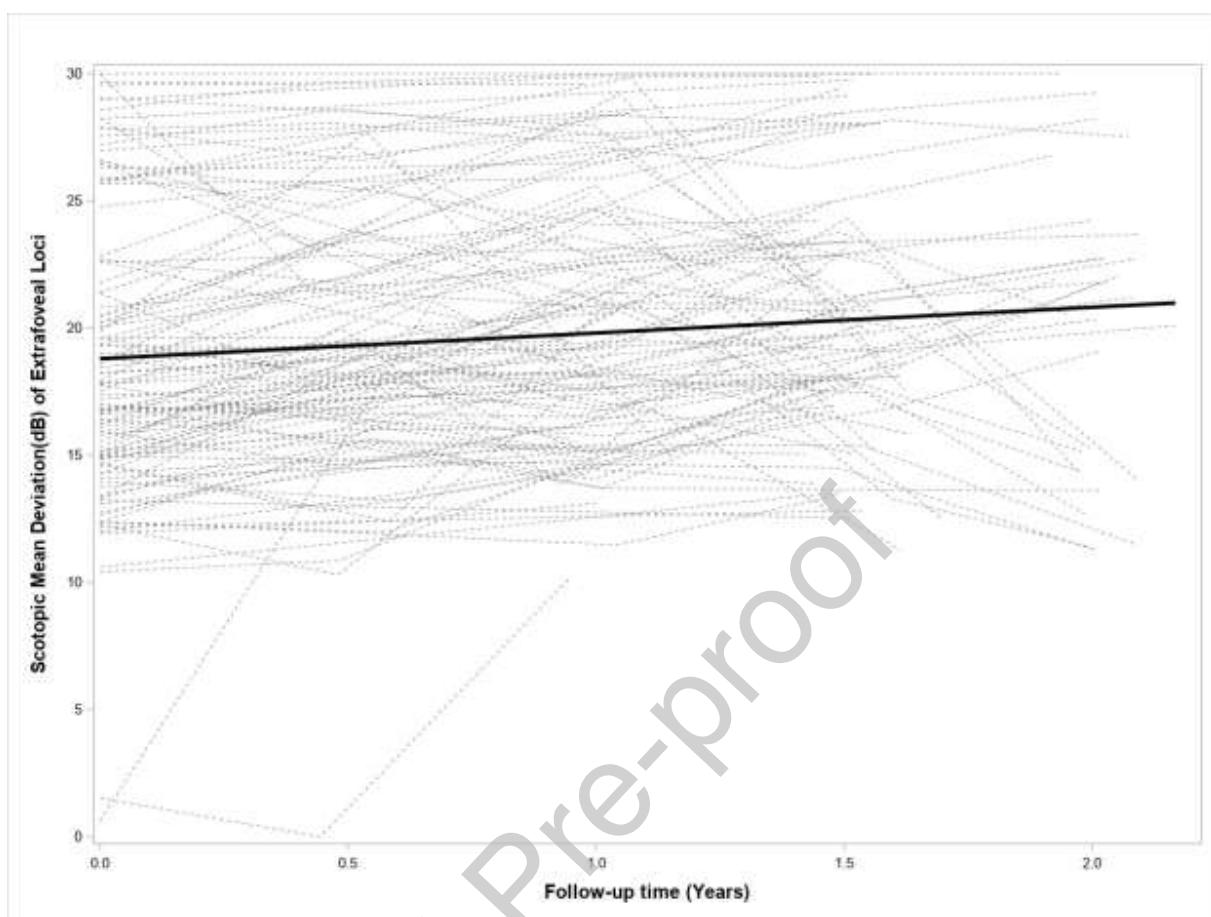


Figure 3. Spaghetti plots showing the longitudinal trajectories of mean deviations (MD) during the follow-up. Each dotted line shows observed data for one eye. The real line is the trend of average MD over time estimated using linear mixed effects model.

- A. Mesopic MD of the fovea loci. The slope estimate is 0.72 (95%CI: 0.37 to 1.07) dB/year.
- B. Mesopic MD of the extrafoveal loci. The slope estimate is 0.86 (95%CI: 0.58 to 1.14) dB/year.
- C. Scotopic MD of the extrafoveal loci. The slope estimate is 1.12 (95%CI: 0.66 to 1.57) dB/year.



Figure 4. Fundus autofluorescence (FAF) images of the 2 eyes that had normal extrafoveal mesopic and scotopic sensitivity responses at SMART baseline.

- A. FAF image of Eye 1 in Table 3 acquired at the SMART baseline visit (left) and the image acquired approximate 12-months after the SMART baseline visit (right).
- B. FAF image of Eye 2 in Table 3 acquired at the SMART baseline visit (left) and image acquired at approximate 18-months after the SMART baseline visit (right).

Table 1. Baseline demographics and clinical characteristics of participants enrolled in the SMART study.

		Number (n=127)	%	
<b>Gender</b>	Female	66	52.0	
<b>Race</b>	White/Caucasian	105	82.7	
	Black/African American	12	9.5	
	Asian	7	5.5	
	Unknown	3	1.6	
<b>Age at onset</b>	≤ 18	53	41.7	
	>18	68	53.5	
	unknown	6	4.7	
		Mean	SD	Median IQR Range
<b>Age at Baseline (years)</b>		34.5	15.1	33.0 22.0-45.0 11.0-70.0
<b>Duration of symptoms (years)</b>		10.1	7.0	8.5 5.5-13.5 0-30.0
<b>Age at onset among known (years)</b>		24.7	14.5	21.0 13.0-36.0 4.0-64.0
<b>Best corrected visual acuity (ETDRS Letters) (Snellen equivalent)</b>		52.0 (20/91)	18.6 (20/126)	45.0 (20/166- 20/40) (20/417 to 20/14)
<b>Mean Deviations (MD) (dB) in all eyes (N=127)</b>				
Overall Mesopic MD (central 20°)		8.36	4.79	7.20 4.80 to 0 to 19.90 10.80
Mesopic MD in the fovea (loci of 2° eccentricity)		16.78	5.32	19.50 15.00 to 0 to 20.00 20.00
Mesopic MD Extrafovea (loci of 4-10° eccentricity)		7.91	4.90	6.59 4.22 to 0 to 19.94 10.31
Scotopic MD Extrafovea (loci of 4-10° eccentricity)		18.59	5.60	17.40 14.90 to 0.63 to 21.78 30.00
Extrafovea Difference*: Scotopic MD-Mesopic MD		10.68	3.53	10.69 9.09 to -0.03 to 12.21 24.93
<b>Mean Deviations (MD) in eyes with approximately normal extrafovea mMD at baseline (N=81)</b>				
Mesopic MD in the fovea		15.74	6.00	19.0 11.5 to 20.0 0 to 20

Scotopic MD Extrafovea	15.72	4.07	15.55	13.73 to 17.75	0.63 to 29.05
<b>Mean Deviations (MD) in eyes with approximately normal fovea mMD at baseline (N=11)</b>					
Mesopic MD in the extrafovea	2.67	2.89	1.66	0.63 to 3.69	0-9.34
Scotopic MD Extrafovea	12.67	7.06	12.58	10.43 to 15.56	0.63 to 25.68

IQR= interquartile range, SD = standard deviation. MD=Mean deviation from maximally measurable normal under Nidek MP-1S

\*: Positive difference between scotopic and mesopic MDs means larger impairment in sMD.

Table 2. Rates of declines in foveal mesopic and extrafoveal mesopic and scotopic mean deviations. mMD: mesopic mean deviation. sMD: scotopic mean deviation. CI: confidence interval.

	Rates		
	Foveal mMD (dB/year) (95%CI)	Extrafovea mMD (dB) (95%CI)	Extrafovea sMD (dB) (95%CI)
<b>Among all eyes (N=127)</b>	0.72 (0.37 to 1.07)	0.86 (0.58 to 1.14)	1.12 (0.66 to 1.57)
<b>Among the eyes with approximately normal extrafovea mMD at baseline (N=81)</b>	0.96 (0.47 to 1.46)	0.96 (0.61 to 1.32)	1.31 (0.74 to 1.89)
<b>Among the eyes with approximately normal fovea mMD at baseline (N=11)</b>	2.98 (-0.21, 6.18)	1.20 (-0.06 to 2.46)	1.58 (-0.84 to 4.00)

Table 3. Detailed data for the 2 eyes that had both scotopic and mesopic extrafoveal sensitivity above average normal at the baseline.

		<b>Foveal mMD (dB)</b>	<b>Extrafovea mMD (dB)</b>	<b>Extrafovea sMD (dB)</b>	<b>Atrophic lesion size in FAF (mm<sup>2</sup>)*</b>	<b>BCVA(ETDRS Letters) (Snellen equivalent)</b>
<b>Eye 1</b>	First visit	2.0	0.94	1.53	2.19	48 (20/110)
	Last visit (12- months follow-up)	10	2.41	10.20	2.25	49 (20/100)
<b>Eye 2</b>	First visit	0	0.13	0.63	0.57	45 (20/126)
	Last visit (18- months follow-up)	20.0	4.41	11.28	1.59	43 (20/138)

mMD: mesopic mean deviation.

sMD: scotopic mean deviation.

\*: Atrophic lesion size was measured as the area of decreased autofluorescence

FAF: fundus autofluorescence

BCVA: best corrected visual acuity

ETDRS: Early Treatment Diabetic Retinopathy Study