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Let's Talk About Sex: Differences in Drug Therapy in Males and Females

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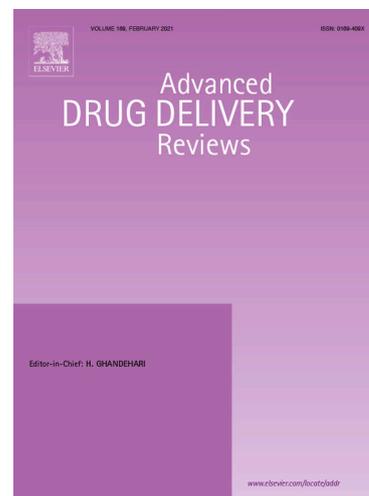
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Let's Talk About Sex: Differences in Drug Therapy in Males and Females

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Abstract

Professor Henry Higgins in *My Fair Lady* said, ‘Why can’t a woman be more like a man?’. Perhaps unintended, such narration extends to the reality of current drug development. A clear sex-gap exists in pharmaceutical research spanning from preclinical studies, clinical trials to post-marketing surveillance with a bias towards males. Consequently, women experience adverse drug reactions from approved drug products more often than men. Distinct differences in pharmaceutical response across drug classes and the lack of understanding of disease pathophysiology also exists between the sexes, often leading to suboptimal drug therapy in women. This review explores the influence of sex as a biological variable in drug delivery, pharmacokinetic response and overall efficacy in the context of pharmaceutical research and practice in the clinic. Prospective recommendations are provided to guide researchers towards the consideration of sex differences in methodologies and analyses. The promotion of disaggregating data according to sex to strengthen scientific rigour, encouraging innovation through the personalisation of medicines and adopting machine learning algorithms is vital for optimised drug development in the sexes and population health equity.

Keywords

Sex and gender differences; Gastrointestinal pharmacokinetics and pharmacodynamics; Drug response and side effects; Personalized pharmaceuticals and medicines; Artificial intelligence and machine learning; 3D printing drug delivery systems; In silico and PBPK modeling; Cell lines; Pharmaceutical drug product design and development; Health equity

1.0 Introduction

In general, women are prescribed more drugs than men, require increased access to health care services but suffer from more adverse drug reactions (ADRs) and are hospitalised more often due to ADRs than men (even when adjusted for age-related differences) [1]. The lack of consideration of potential sex differences exists in nearly all areas of research and development and has seeped into the mainstay of society. For example, research from van Hoof et al. observed that office building thermostats are based on male metabolic rates with temperatures set too low for many women [2]. Some consequences, however, can be life threatening. In engineering, many devices and machines have been designed to fit male bodies; military and commercial cockpits were traditionally based on male anthropometry. As a consequence, it was potentially dangerous for some women or small men to become pilots [3]. Appropriate female representation has vastly been ignored in scientific research including immunology, pharmacology and neuroscience. Interestingly, in the preclinical field of behaviour and reproduction, sex as a biological variable is of particular interest [4].

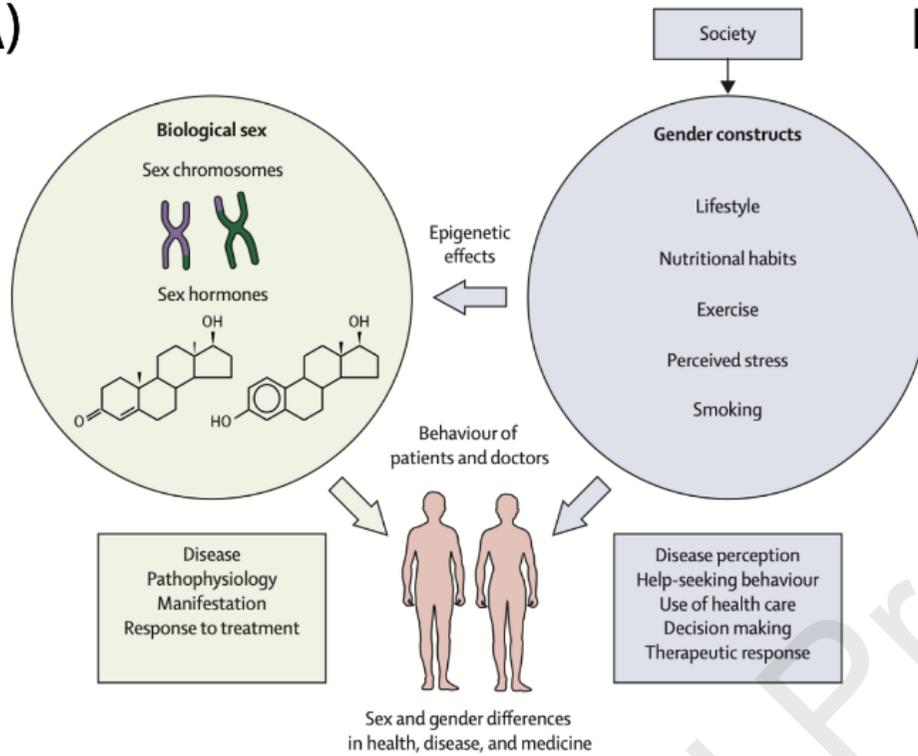
Our cells are innately infused with sex differences that cannot be ignored. The phrase “every cell has a sex” captures the essence of how fundamentally different men and women are when it comes to health and disease (Figure 1a). In fact, every nucleated cell has a sex containing the sex chromosomes (in its simplest form, XX in females or XY in males [5]). A female-predominance to chronic disease is seen in epidemiology, pathology, clinical course and diagnosis of Alzheimer’s disease, influenza and pneumonia to name a few (Figure 1b). The onset and development of heart disease, specific cancers and chronic pulmonary disease, however, are leading causes of death in men [6]. In light of the COVID-19 pandemic, the global number of confirmed cases, severe symptoms, differing immune response and mortality rate due to the disease are higher among men [7-10]. However, recent epidemiology data from the COVID Symptom Study application revealed that women were more likely to develop “long COVID” where symptoms persist for longer than 12 weeks [11].

In biomedical research, women and non-human female mammals have often been under-represented. Although there is some recognition today of the need for appropriate female representation in clinical trials, in previous decades, the consideration and inclusion of males overshadowed females in clinical research design and conduct. Figure 1c demonstrates a male

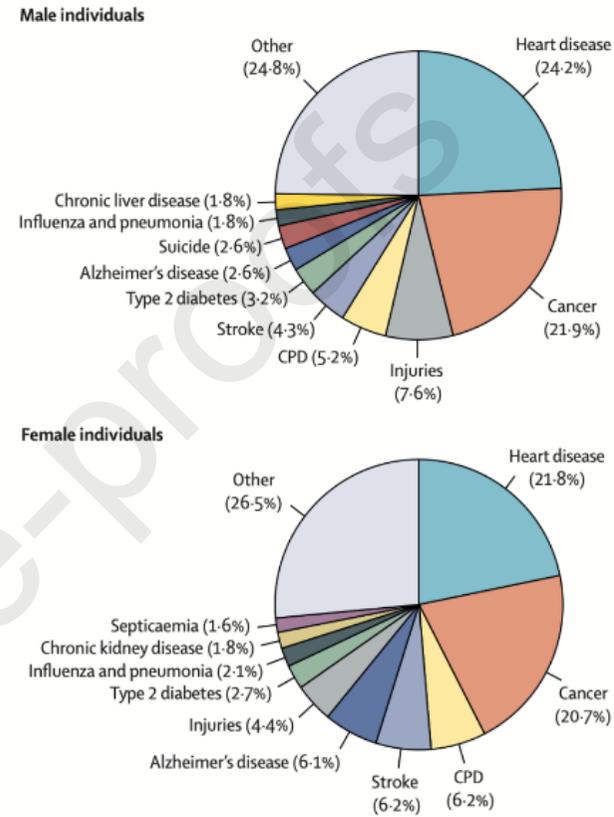
bias in articles involving interdisciplinary research including biology, neuroscience, physiology, pharmacology and behaviour. Females, however, were the sex of interest in subject areas spanning reproduction, endocrinology and behavioural physiology [4]. Although Figure 1c outlines the employment of both sexes, it is clear that results are seldom disaggregated according to sex which can potentially skew or even conceal sex-specific differences in biomedical research (Figure 4) [4]. The leading assumption is that i) results from male studies appropriately apply to females or ii) hormonal cycles decrease the homogeneity of study populations and complicate experimental designs to such an extent that it may not be worth studying females from the outset [4]. In addition, the risk of adverse effects such as teratogenicity outweighed other considerations and thus, females of child-bearing potential were largely excluded from clinical trials [12]. In some cases, little evidence exists for the safety profiles of drugs in pregnant or breastfeeding females, therefore such females and their healthcare professionals are advised to consider the risk-benefit ratio of therapeutic use. The COVID-19 vaccination programme is such an example [13]. In addition, when studying diseases prevalent in both sexes, Caucasian males were considered to be the typical study population [14] highlighting the lack of consideration of potential ethnic differences too [15].

To address the historical overrepresentation of male subjects in biomedical research, a 10-year follow-up study of sex inclusion across interdisciplinary research was conducted by Woitowich et al. [16]. The work identified that there was a significant increase in the number of studies that included both sexes across general biology, immunology, neuroscience, physiology, pharmacology, endocrinology, reproduction, behavioural physiology and behaviour. However, in all subject areas bar pharmacology, there was no change in the proportion of studies that included data specifically analysed by sex. In addition, the studies failed to provide rationale for single-sex studies or the lack of sex-bases analyses outlining a clear sex gap in biological research disciplines [16].

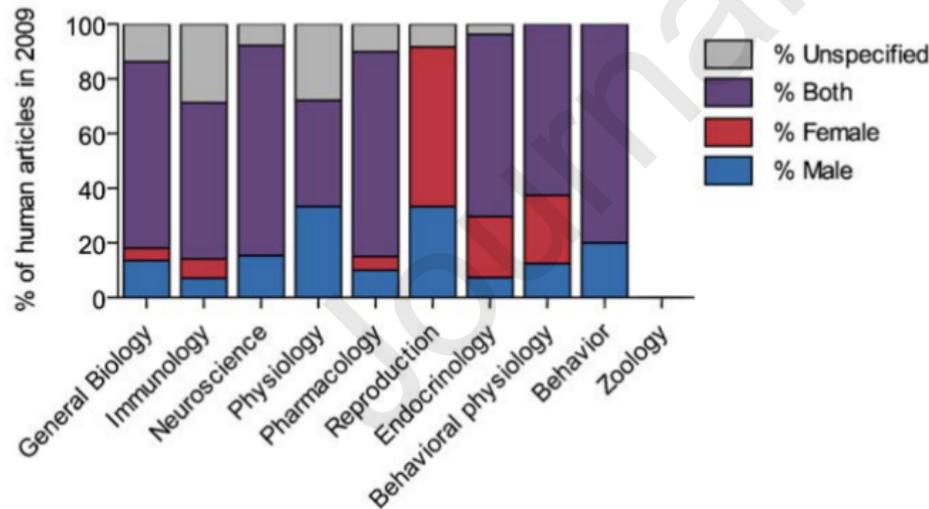
A)



B)



C)



D)

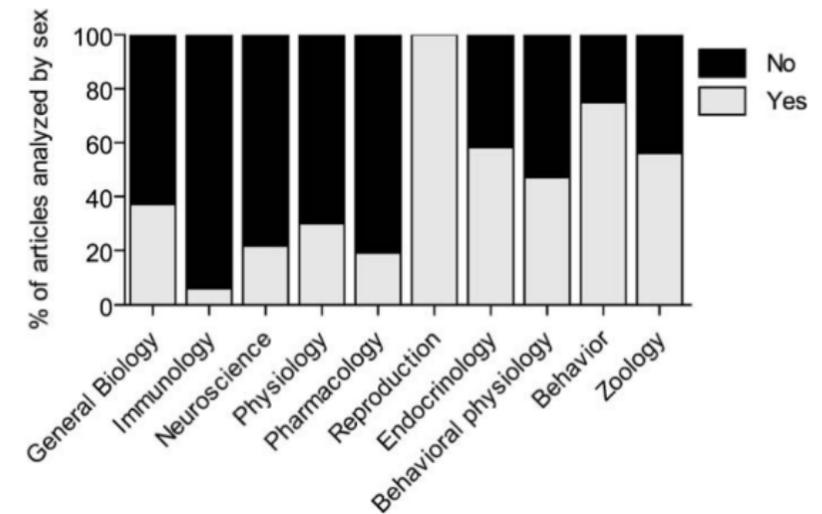


Figure 1. Sex as a biological variable as modifiers of health, disease and research outcomes. **A)** Inter-relation between biological sex and societal gender in health, disease pathophysiology and clinical manifestations. **B)** Distribution of the ten leading causes of death disaggregated by sex in the US in 2017. (CPD = chronic pulmonary disease). **C)** Percentage of articles that used male subjects, female subjects, both male and female subjects or did not specify sex of sample population. **D)** Percentage of articles disaggregating pre-clinical animal results according to sex in different scientific disciplines. Adapted with permission from [4] and [6].

To clarify nomenclature, this review will go forward with ‘sex’ and not ‘gender’ in its terminology. Sex refers to the biology of living things, i.e. as male or female according to reproductive organs of functions based on the chromosomal complement [5]. Gender, however, refers to sociocultural attributes, behaviours or personal identification [17]. As such, this review will comprehensively report on how the lack of pharmaceutical analyses considering sex differences in systems biology, sex-specific needs and behaviour, regulatory affairs and post-marketing surveillance has led to a disparity in optimum drug development. As peroral administration is the main route of drug delivery due to convenience and consequently patient medicine adherence [18], a key focus is dedicated towards sex differences in the gastrointestinal (GI) tract and drug absorption in male and female human adults or equivalent preclinical models. Recommendations will also be proposed on how scientists should rethink standards and reference models across the drug delivery pipeline with the aim to integrate sex analyses into research and innovation.

2.0 Sex bias in pharmaceutical research

Government reports from 1980s to 90s indicated that women had lower representation in federally funded studies investigating diseases that affected both sexes [19-22]. In 1992, the United States of America’s Food and Drug Agency (FDA) and Food and Drug Law Institute concluded that young women needed to be included in clinical trials in order to understand female response to pharmaceuticals [12]. The regulation and guidance published by the FDA on female participation in industry-sponsored clinical trials has transformed over the half century, instigated by the thalidomide tragedy in pregnant women [23]. Thalidomide was developed by the Swiss company CIBA in 1953 and introduced to the pharmaceutical market in 1956 by German pharmaceutical company, Chemie Grunenthal [24]. Initially marketed with the brand name Contergan, thalidomide was prescribed as a non-barbiturate sedative to induce deep sleep. Pre-clinical testing, however, failed to establish a median toxic dose and the drug was believed to be non-toxic to humans [25]. In that era, testing for harmful teratogenic effects were not considered. The drug was used as a sedative but soon became popular for its anti-emetic effects in pregnant women suffering with morning sickness [26]. In 1961, observations linked thalidomide use in pregnancy to congenital malformations in multiple cases worldwide [25, 27], and thalidomide was ultimately withdrawn from the pharmaceutical market in the same year.

In 1977, a guidance document from the FDA advised that women of child-bearing potential (females capable of becoming pregnant, including pre-menopausal single abstinent women, women using contraceptives or women with sterile partners) should be excluded from Phase I and early-Phase II trials. If a drug was deemed to have a positive risk-benefit ratio, women could then be included in late-Phase II and Phase III trials providing animal teratogenicity and fertility studies were completed [20]. In 1993, however, the FDA reversed the 1977 FDA guidance which lifted the ban on women of child-bearing potential to be excluded from early clinical trials research. The guidance further specified that clinical trial participants should be representative of the patient population likely to be prescribed the drug once regulated for market approval [20]. In the same year, the National Institutes of Health (NIH) formalised an NIH Revitalisation Act entitled Women and Minorities as Subjects in Clinical Research where four main issues were addressed; 1) Women and minorities are to be included in all clinical research, 2) Numbers in Phase III clinical trials be sufficient to allow for valid analyses of potential sex and ethnic differences, 3) Such groups should not be excluded due to trial costs and, 4) Programmes and support outreach efforts should be created to enrol women and minorities in clinical trials [28]. Although progress has been made towards the appropriate representation of females across the whole clinical arena in the last decade (Figure 2), sex differences in drug response are still demonstrated following regulatory approval and entry of a drug into market [29].

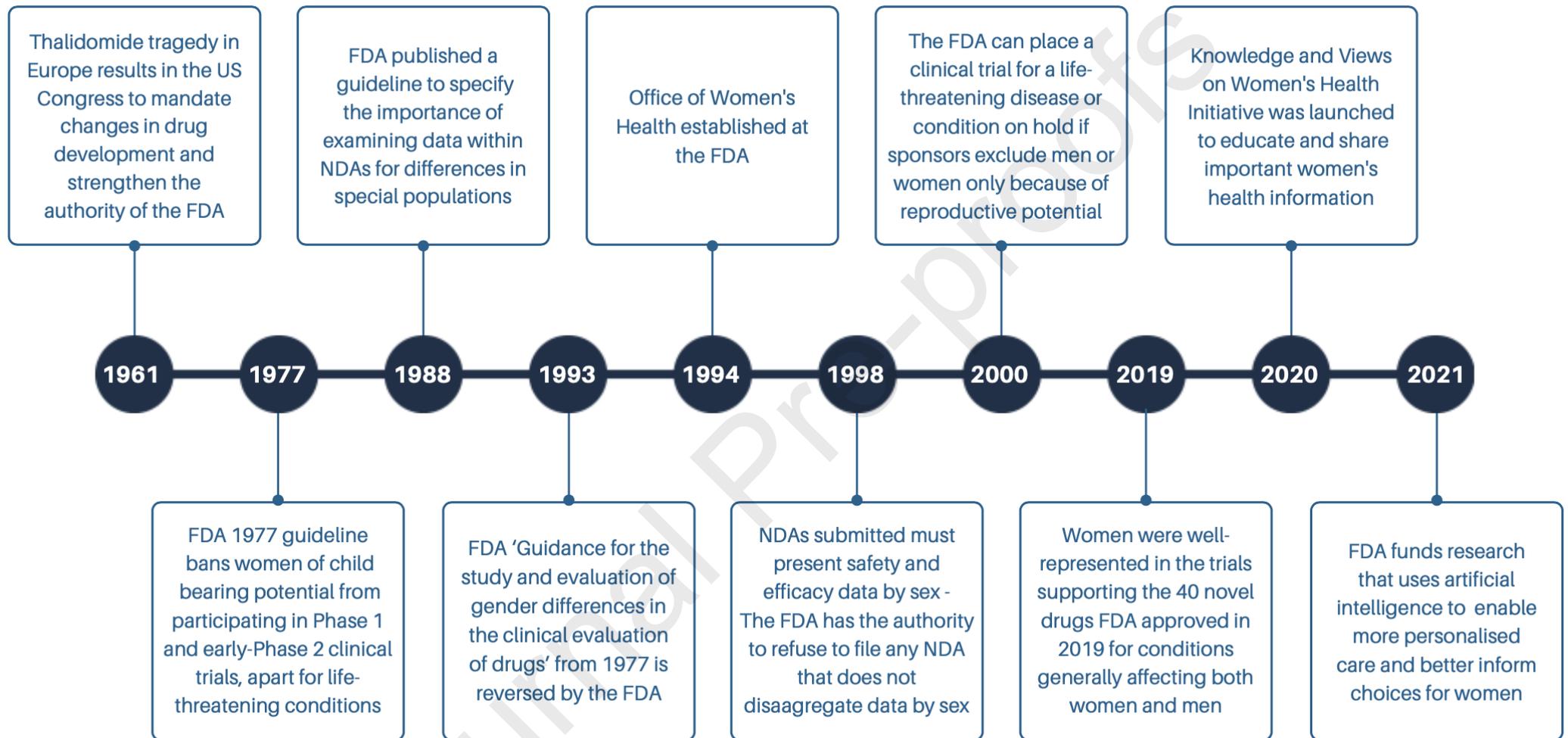


Figure 2. Significant events in the history of female participation in clinical trials in the United States in line with guidelines from the US FDA.

Adapted from [30, 31].

A 2018 review of 107 NIH funded randomised control trials that enrolled both men and women found that only 26% reported even one outcome disaggregated by sex or included both sexes as a co-variate [32]. 72% simply did not include sex as a factor in their analyses. NIH policies mandated over a quarter century ago have yet to yield the intended increases in reporting by sex. A consequence of this sex inequality hides in plain sight today: most drugs are prescribed to women and men at the same dose. Many currently prescribed drugs were approved by FDA prior to 1993, with inadequate inclusion of female animals in preclinical research and of women in clinical trials [33]. The existing knowledge base on sex-aware prescribing lacks information on sex differences for one-third of all drugs [34, 35]. Pharmaceutical companies responsible for generating pre-approval data often fail to include information on sex differences in New Drug Applications (NDA) documents, and the FDA has previously failed to enforce its own requirements before approving new drugs [36]. Consequently, potential sex differences in pharmacokinetics, pharmacodynamics and their relation to adverse side effects often remain unknown. Most of the data submitted to the FDA by drug companies are not publicly available and not subject to peer-review by the broader scientific community [35]. Regulatory agencies have historically paid insufficient attention to differences between women and men in terms of both sex and gender which perpetuates inequalities by neglecting drug safety problems that are sex-specific. In addition, this disparity allows for misleading drug marketing [36]. For example, irritable bowel syndrome (IBS) is disproportionately diagnosed in females, despite recent evidence that males equally both suffer and access health advice for IBS symptoms [37, 38]. Tegaserod was approved for IBS in females first, followed by a FDA-approved extension to males for chronic constipation based on two clinical trials with over 85% females participants [39, 40]. Tegaserod was later removed from market following a meta-analysis of 29 trials reporting an increased risk of cardiac adverse events [41].

In the preclinical arena, routine *in vitro* models, namely cell lines, are not sufficient to study and understand sex differences in early drug development as the cells are often derived from a single animal or human subject to reflect a specific organ. The sex of cell lines is often not reported, failing to acknowledge potential sex differences in the *in vitro* mechanisms. To overcome these shortcomings, scientists should state the sex of their *in vitro* models in their publications. Cvitanović Tomaš et al. have created a computational model LiverSex [42], taking sex differences in the liver into account, adapted from SteatoNet *in silico* model [43]. Data from oestrogen and androgen receptor responses are included which includes sex-related effects on growth hormone release. Currently, the model has been validated in mice but not in

humans. A step further, Thiele et al., have created two validated, sex-specific, whole body metabolic models called Harvey and Harvetta [44]. Here, the male and female physiologies have been represented with 20 organs, 6 sex organs, 6 types of blood cells, the systemic blood circulation, the blood-brain barrier and the GI lumen, including the microbiome. These sex-specific models represent systems biological approaches to precision medicine.

3.0 Sex differences in human physiology

Men Are From Mars, Women Are From Venus by John Gray in 1992 outlined that differences in communication tactics between males and females stem from fundamental differences in psychological processes between the sexes. Indeed, such sex differences are not limited to psychology but extend to the complete physiological system and anatomy itself. For many years - except for studies related to the physiology of reproduction - physiological principles contained in classical physiological and medical textbooks have been based on the androcentric model of 70 kg healthy Caucasian males between 18 to 40 years of age [45]. In addition, thousands of genes differ in their expression between males and females in the liver, adipose tissue and muscle with the brain being less sexually dimorphic [42]. The appreciation of this led to the US Institute of Medicines declaring in 2001 that biological sex will considerably affect the course and prevention of disease [46].

Significant physiological differences exist between men and women such as percentage of body fat, body water volume, plasma volume and organ blood flow, in addition to body weight (Figure 3). As such, women are not small men. These parameters, however, are often overlooked in the drug development process and can consequently lead to differing response to medicines [15]. These have been reviewed elsewhere [47, 48], however herein, focus will be invested towards sex differences in the processes involved following solid oral drug administration, i.e. the GI tract.

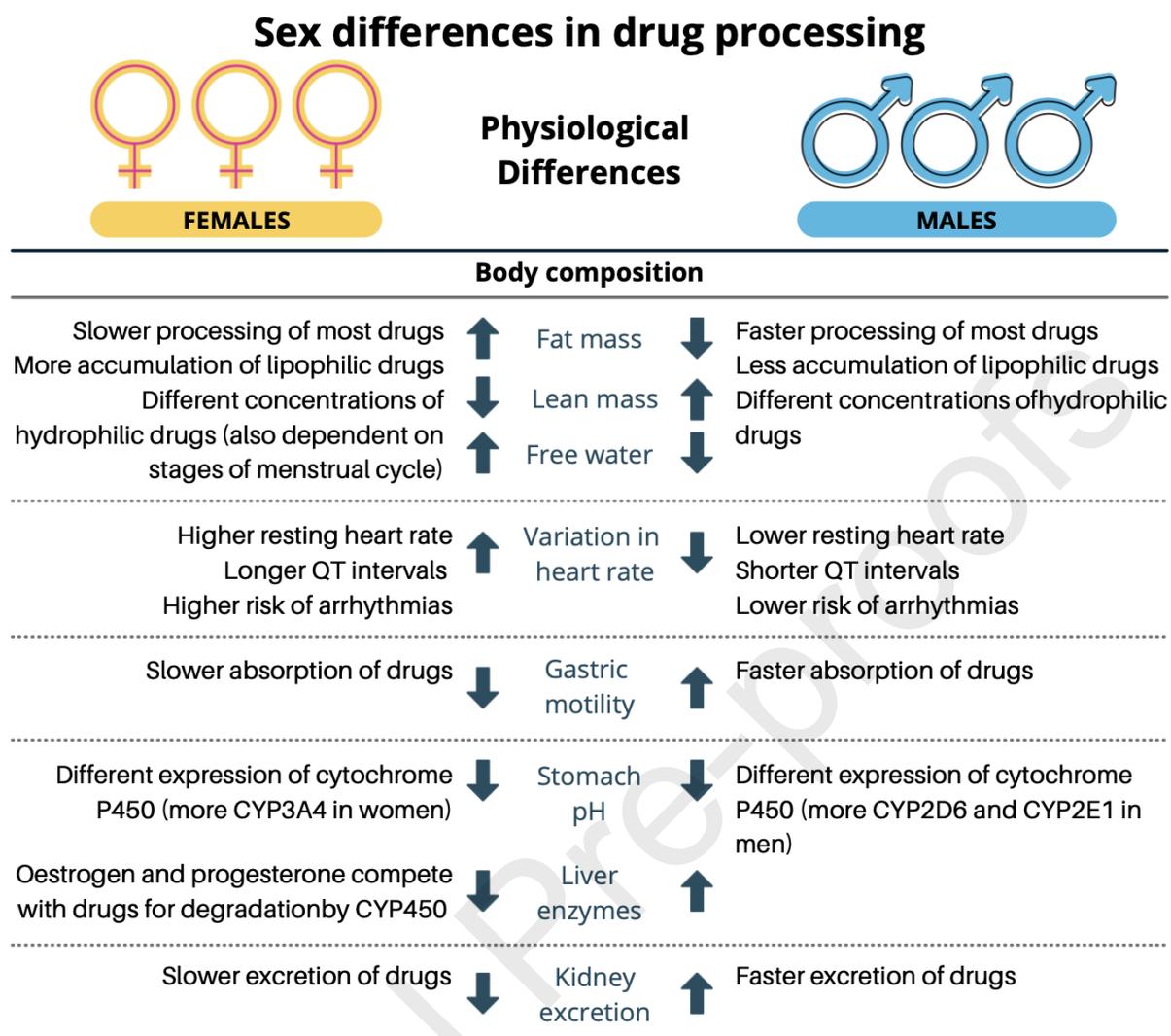


Figure 3. Physiological differences in males and females that can affect drug processing. Adapted with permission from [49].

3.1 Sex differences in gastrointestinal physiology and the influence on oral drug performance

There are significant sex-specific differences in terms of drug bioavailability and pharmacokinetics which can, in turn, differentially affect drug efficacy and safety. Underlying reasons for sex-related variations in drug performance include obvious differences in physiological parameters such as body fat content and hormonal control [50]. Fundamental differences at the level of the GI tract, liver and kidneys can further influence drug absorption, distribution, metabolism and elimination, and consequently lead to variability in drug therapy and potential toxicity [51]. It is difficult to envisage that differences in drug performance and adverse effects are linked to a single pharmacokinetic parameter and governed by a single

organ. Instead, it is much more likely that sex differences may be a result of the interplay of the complete system following oral drug administration. Herein, focus on the GI tract, potential differences between the sexes and its influence on oral drug variability will be discussed.

Tissue exposure of orally administered medication is affected by variability in gastric fluid pH and volumes, gastric emptying time (GET) and intestinal transit time (ITT), competition and/or regulation of intestinal transporters and drug metabolising enzymes, and the potential interactions of sex steroids on drug PK [52] (Figure 4). In terms of gastric and small intestinal fluid volumes, males have been reported to have higher volumes than females [53] which may affect the extent of drug dissolution. Average fasted gastric pH is significantly higher in females (2.79 ± 0.18) than in males (2.16 ± 0.09) ($p < 0.05$) which may be attributed to reduced acid secretion due to the smaller stomach size seen in females [54]. The basal acid output in the fasted state was nearly half in females than in males, 2.1 ± 0.2 and 4.0 ± 0.2 mmol/h, respectively [55]. Lowered gastric acid secretion may influence drug ionisation, particularly of weak bases, the solubility of pH-sensitive drugs and the degradation of acid-labile drugs, thereby affecting absorption and consequently, oral drug bioavailability. Sex differences have also been reported in bile acid composition with higher concentrations of cholic acid being reported in males and higher concentrations of chenodeoxycholic acid reported in females [56].

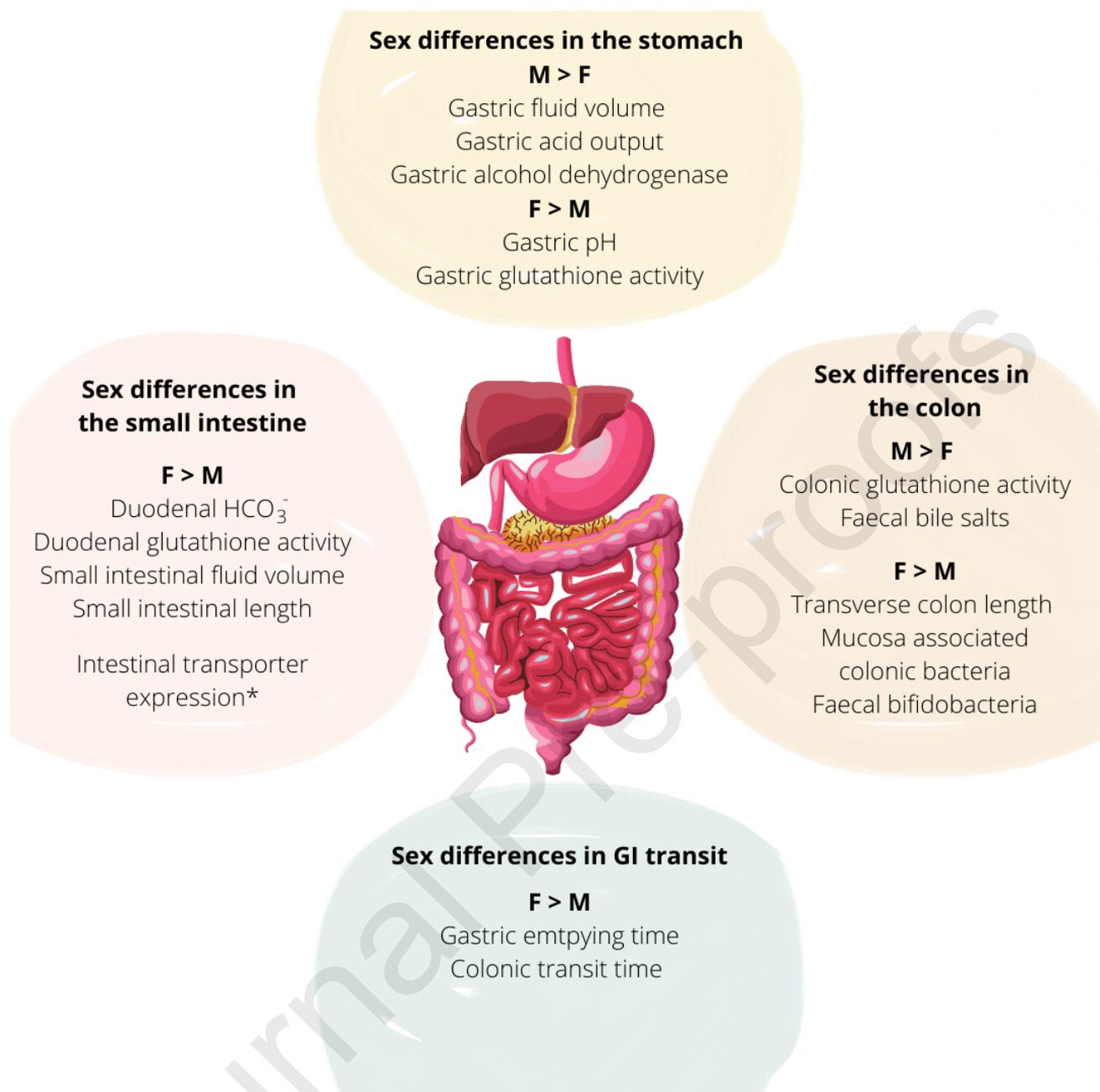


Figure 4. Key sex differences at the level of the gastrointestinal tract that impact oral drug delivery and bioavailability, (M = Male; F = Female). Adapted with permission from [57]. *Denotes variable transporter expression in the regions of the small intestine between males and females.

Females have a significantly longer GET for solids and calorific liquids (118.0 ± 8.1 min) compared with males (91.4 ± 7.5 min), however, GET decreases in post-menopausal women (97.9 ± 7.6 min) becoming similar to that in men of the same age [58]. Variabilities in drug pharmacokinetics can be attributed to differences in GET; for example, peak plasma concentration of orally administered carbidopa was achieved 22 min later in women than men

due to longer GET [59]. Sex differences in the oral bioavailability of a gastro-resistant ketoprofen formulation has also been demonstrated. Males showed a higher c_{\max}/AUC than females (0.468 ± 0.094 versus $0.361 \pm 0.087 \text{ h}^{-1}$) and a significantly lower t_{\max} (3 – 5 h versus 5 – 10 h) respectively. Such differences were attributed to the faster SITT in males which allowed for ketoprofen to reach the appropriate site of absorption in the intestinal environment for absorption to occur more rapidly [60], therefore leading to t_{\max} to occur at an earlier time point. Females have longer transverse and descending CTT, but shorter rectosigmoid transit time compared to males [61]. The longer GET and CTT and so the overall GI residence of sustained-release dosage forms may facilitate enhanced drug absorption in women, as demonstrated with diltiazem which is sensitive to GI transit time [62]. This, however, may be further affected by the regulation of intestinal membrane transporters and metabolising enzymes located in the GI mucosa.

Distinct sex differences in drug performance have been further demonstrated in treatments for GI syndromes. For example, alosetron, a 5-hydroxytryptamine (5-HT) receptor 3 antagonist, is a drug that is effective in females but has low performance in males [63]. At identical plasma concentrations, alosetron achieves therapeutic levels only in females. Sex differences may partially contribute towards variability in the activity of serotonergic receptors in the colon. Serotonergic type III receptors are involved in postprandial colonic responses in health and diarrhoea and findings reported that with alosetron treatment, females displayed a significantly greater overall colonic transit compared to males (a colonic geometric centre mean at 24h of -1.45 and -0.32, respectively) [64]. Viramontes et al. proposed that the pharmacogenomics of 5-HT₃ may be a factor. Although, additional studies into serotonin synthesis and genotypic serotonin synthesis and metabolism were suggested to further understand the sex difference [64]. Alosetron, however, was withdrawn from the pharmaceutical market in 2000 due to significant side effects but was reintroduced in 2002 in the US under restrictive conditions of use only for females suffering from severe diarrhoea-related IBS [65].

The gut microbiota adds further to the complexity to GI physiology and varying drug response in males and females. For example, levodopa, a treatment for Parkinson's disease, has been subject to increased metabolism in the presence of *Helicobacter pylori*, which consequently reduced the drug bioavailability. The eradication of *Helicobacter pylori* infection, however, improved levodopa action and clinical symptoms. The prevalence of *Helicobacter pylori*

infection, however, is more prevalent in male than female individuals [66] and as such, may lead to differences in levodopa pharmacokinetics between the sexes.

Research in the understanding of differences between the sexes and the clinical performance of drugs continues to be limited. It is clear that males and females respond differently to medicines due to the dynamic interplay of GI physiology, drug pharmacokinetics and contributions from other associated organs. A single pharmacokinetic parameter cannot be considered as the rate-limiting step as this may occur in a drug-by-drug basis. For a better understanding of the basic mechanisms of sex differences, future studies should be designed with this primary focus in mind to determine the extent that these differences may have on clinical management [67].

4.0 Sex differences in pharmacokinetics

Many drugs show distinct pharmacokinetic differences between the sexes in humans (Table 2) and preclinical animal models (Table 3). A hallmark example displaying significant sex differences in drug response is the sedative zolpidem. It was approved by the FDA one year before the NIH Revitalisation Act and marketed under several names including Ambien, Edluar and Zolpimist, where males and females were prescribed the same dose of 10 mg and 12.5 mg for immediate-release and extended-release products, respectively [68]. During decades of post-marketing drug surveillance, women were found to be more susceptible to next-day effects due to a slower rate of drug elimination, with emergency department visits from exclusively females with cognitive defects [69]. The FDA subsequently recommended that the dose of zolpidem be reduced by half for women [70]. Many other drugs administered in equal doses to males and females likely require re-evaluation for sex-specific dose adjustment. An analysis of 10 prescription drugs that were withdrawn from the market from 1997 – 2001 found that eight posed “greater health risks for women”, mainly because of adverse drug events due to known pharmacodynamic differences (e.g. 3 drugs withdrawn due to risk of Torsades de Pointes) or because women are more prone to polypharmacy [71].

Apart from the innate differences in physiology, chemical and biological processes, pharmacokinetic and pharmacodynamic processes add further to the complications of varying drug response. In addition, endogenous steroid hormone exposure (from peripubertal to adulthood) and sex differences in exogenously administered steroids, the higher rates of

polypharmacy in women and sex differences in reporting rates contribute to the manifestation of sex differences in drug response [72]. Up to 6 – 7% of new drug applications that include sex analysis report at least a 40% difference in pharmacokinetics between males and females [73]. In general, drug disposition occurs through separate phases including absorption, distribution, metabolism and elimination. Sex differences have been demonstrated for each phase [50, 74].

4.1 Absorption

Absorption of drug products across the gut epithelium depends on a number of highly complex mechanisms [75]. Absorption can be a rate-limiting step for an orally administered drug to reach its target site of action and is drug- and mechanism-dependent. Sex differences in absorption can be seen for a number of drugs. In addition, sex hormones were recently found to affect the passive diffusion and active transport of drugs to different extents in males and females [76].

A key case that illustrates sex differences in drug absorption is aspirin. For example, one study showed that oral administration of aspirin in young healthy adult males ($n = 9$) and females ($n = 9$) resulted in faster oral absorption in females than in males (statistically significant terminal $t_{1/2}$ [16.2 and 20.6 mins] and mean residence times [33.5 and 39.9 mins], respectively) [77]. Whilst the females were lighter in weight than the males, which resulted in different dose per kilogram body weight, weight was not considered a major factor in absorption as the pharmacokinetics of aspirin was reported to be independent of dose [78]. A further study on aspirin disposition in seven young females, six young males, six elderly females and six elderly males found statistically significant higher plasma levels (C_p max) in young and elderly females compared with the male counterparts. Age, on the other hand, did not show a statistically significant effect on the pharmacokinetics of aspirin [79, 80].

A population pharmacokinetic analysis ($n = 449$ for learning and $n = 247$ for validation with similar clinical and biological characteristics except for weight) showed a longer time of absorption (t_{max}) in males, with medians of over 3 hrs for men, compared with 40 mins in females, for the antihistamine mizolastine [81]. The absorption of copper was reported to be significantly ($p = 0.02$) higher in females (71%) than in males (64%) aged 20 – 59 years ($n = 127$) [82]. Interestingly, the permeability of lactulose and sucralose was reported to decline

with ageing in females ($p = 0.05$, $r^2 = 0.24$ and $p = 0.01$, $r^2 = 0.41$, respectively) but not in males in healthy adults ($n = 17$) and children ($n = 15$), with a suggestion that the age- and sex-related deterioration was mediated by glucocorticoid hormones [83].

Drugs may compete for intestinal membrane transporters into cells that affect the downstream metabolism or availability of the drug at its target site. The influx pump OATP1B1, encoded by the gene *SLCO1B1*, transports oestrogens including estrone-3-sulfate and oestradiol 17 β -D-glucuronide, as well as statins. Competition for the transporter may occur when multiple substrates are present [84], which may limit the efficacy of statin treatment [85]. Sex-specific effects of *SLCO1B1* genetic variants (*SLCO1B1* rs4149056 (T > C) polymorphism) were reported in a pilot study, which showed homozygous males displayed the lowest decrease ($\Delta - 21.2 \pm 7.2\%$) of total cholesterol, compared with females where the same genotype was associated with the highest ($\Delta - 33.5 \pm 7.6\%$) decrease ($P = 0.04$) [86]. Females of older age were associated with an increased risk of statin-related myopathy (relative risk of 2.0 (95% confidence interval, 1.0 to 3.9)), especially amongst carriers of the *SLCO1B1* c.521C allele with impaired renal function and diabetes and those who take amiodarone [87].

In addition, there is an increasing body of literature evidence that report the inherent sex-specific expression of a number of efflux transporters (Table 1) [88, 89] that result in differential treatment outcomes (Figure 5a). P-glycoprotein (P-gp) is the most studied drug efflux pump, encoded by the *MDR1* gene. Polymorphisms in the *MDR1* gene is linked with higher levels of neutropenia with docetaxel [90]. Sex hormones are believed to modulate P-gp expression and inhibit drug absorption by P-gp-mediated efflux at the intestinal epithelia [91, 92]. In a similar manner, multidrug-resistant protein transporters (MRPs), display sex differences in their expression, modulated by sex hormones [93].

Table 1. Sex differences in efflux transporter mRNA expression between male and female research models in the kidneys, liver, lung, brain and intestinal tract. Adapted from [94].

Tissue and Transporter mRNA	Efflux Membrane Protein Transporter	Model	Sex difference	References
Kidneys				
Ent1	Equilibrative nucleoside	Mice	F > M	[95]

	transporter 1	Rat	M = F	
Ent2	Equilibrative nucleoside	Mice	F > M	
	transporter 2	Rat	M = F	
Ent3	Equilibrative nucleoside	Mice	M > F	
	transporter 3	Rat	M = F	
Mrp3	Multidrug resistance protein 3	Mice	F > M	[96]
Mrp4	Multidrug resistance protein 4	Mice	F > M	
Mdr1a	P-glycoprotein isoform	Mice	F > M	[97]
Mdr1b	P-glycoprotein isoform	Mice	F > M	
Bcrp	Breast cancer resistance protein	Mice	M = F	[98]
		Rat	M > F	
Mate1	Multidrug and toxin extrusion 1	Mice	M > F	[99]
Liver				
Mrp3	Multidrug resistance protein 3	Mice	M = F	[96, 100]
		Rat	F > M	
Mrp4	Multidrug resistance protein 4	Mice	F > M	[96]
Bcrp	Breast cancer resistance protein	Mice	M > F	[98, 101]
		Rat	M = F	
Mate1	Multidrug and toxin extrusion 1	Mice	F > M	[99]
Lungs				
Mdr1b	P-glycoprotein isoform	Mice	F > M	[97]
Mdr2	Multidrug resistance 2	Mice	F > M	
Intestine				
Abca1	Cholesterol efflux regulatory protein	Mice	F > M	[102]
Mate2	Multidrug and toxin extrusion 2	Mice	M > F	[99]
Mdr1a	P-glycoprotein isoform	Rats	M > F	[89]
		Humans	M > F	

4.2 Distribution

Most drugs will bind to plasma proteins in the systemic circulation that are specific to the drug. The distribution of a drug is affected by several body composition parameters (Figure 4). Sex differences in these parameters may account for differences in the concentration of a drug at the target site and result in varying responses.

It has been reported that males have higher content of total body water (i.e. extracellular water and intracellular water), total blood volume and plasma volume than females. The higher percentage of body fat in females, especially in pregnant people, may also alter the distribution of lipid-soluble, slowly metabolised or toxic substances in the body. For example, differences in increased organ blood flow and body fat in females accelerated the onset of action but prolonged the duration of vecuronium and rocuronium bromide in females (e.g.) [103, 104] (Figure 5b). Differences in body fat content and in protein binding are responsible for sex-related pharmacokinetic differences in the distribution of diazepam, where females have been shown to have larger volumes of distribution than males due to higher free fraction [105]. The degree of plasma protein binding is affected by sex hormones with wider variation seen during the time of menstruation [106].

4.3 Metabolism

The majority of drug metabolism occurs in the liver, but biotransformation can also occur in the intestinal tract, lung, kidney and skin. Despite intra-individual variations in drug metabolism following normalisation for height, bodyweight and body surface area, differences in drug metabolism can be dependent on the sex of the individual due to transporters and enzymes expressed in hepatocytes. Drugs metabolised by Phase I and Phase II are cleared faster in males when compared with females [107, 108]. For example, the activity of gastric alcohol dehydrogenase (ADH) is lower in females than in males [109], distinct in younger age (20 – 40 years). As age increases (41 – 60 years), the opposite is found, females show higher gastric ADH activity than males. In older age (61 – 80 years) [110], no sex differences are found. These differences in ADH are believed to be caused by the lower first pass metabolism of alcohol in females, leading to higher blood levels in females than male.

Cytochrome (CYP) enzymes are responsible for the metabolism of a number of drug substrates of which CYP2C and 3A are most commonly expressed in the small intestine. Significant sex

differences are observed in the expression of hepatic drug metabolising enzymes which contribute variabilities in clinical drug performance [111]. Numerous studies have shown that females have higher rates of CYP3A substrate metabolism compared with men [112-114]. A large retrospective analysis into sex dimorphic drug pharmacokinetics found statistically significant sex differences; an average of 20 – 30% higher clearance for drugs that are CYP3A substrates, compared with males [115]. Endogenous and oral exogenous oestrogen are shown to alter hepatic enzymes [116]. The nuclear receptor transporters pregnane X receptor (PXR) and constitutive androstane receptor (CAR) regulate the expression of cytochrome P450 enzymes, including CYP3A4. These receptors are activated by a variety of compounds including steroid hormones [117]. The nuclear hormone receptor ER α has been shown to modulate CYP1B1 expression directly which could affect its drug substrate levels [118].

Interestingly, dose-related sex differences were found in some drug metabolisms. Using zolmitriptan as an example, the bioavailability of zolmitriptan was significantly higher in women than in men after both 5 mg oral dosing and intravenous dosing. However, there were no reported sex differences in oral bioavailability with a dose of 2.5 mg [119]. This sex-related variation was smaller than the finding in the previous report which also demonstrated sex difference in the bioavailability of zolmitriptan after 10 mg oral administration [120] (Figure 5c). It therefore stands a dose-related manner in the bioavailability of zolmitriptan. The reason for this sex-dependent difference was assumed to be most likely explained by a difference in first-pass metabolism [121], as the plasma concentrations of zolmitriptan in women were higher than in men with relatively higher levels of the active metabolite in men.

4.4 Elimination

The kidney is the main site of excretion of waste products following metabolism, xenobiotics, parent drug compounds and their metabolites. In addition, the kidneys are responsible for the maintenance of the water/electrolyte balance and of the synthesis, metabolism and secretion of hormones. There are known sex differences in all three major renal functions (namely glomerular filtration, tubular secretion and tubular reabsorption) resulting in generally higher renal clearance in men than in women [122-124].

A number of transporters present in the kidney show sex-bias in their expression. From investigations into the mRNA expression in human kidneys Joseph et al., found 21 genes with

male dominance and 2 transporter genes with female dominance [125]. Sex differences in drug transporters expression has been suggested for the differential induction of renal diseases via sex-specific toxicities in the kidney. Sex hormones may mediate these differences through alterations in the renin-angiotensin system [126]. Renal sex differences are also seen in the subunits of glutathione-S-transferase (GST) isoenzyme [127]. GST plays a role in cellular detoxification [128] and polymorphisms and sex differences may influence its activity [129]. Female rats showed greater levels of subunits 3 and 4, whereas subunits 1 and 2 showed greater levels in male rats [130]

Aspirin is more rapidly cleared from women and its metabolite, salicylate, has an increased rate of absorption in women. On the other hand, the clearance of acetaminophen, gemcitabine and heparin is slower in females than in males [50, 131], with 71% of patients admitted to hospital for acetaminophen overdose being women [131, 132]. It may be due to the increase in renal blood flow and glomerular filtration in men, which increase the elimination rate of drugs cleared by the kidneys. Renal blood flow, glomerular filtration, tubular secretion and tubular reabsorption are all greater in men than in non-pregnant women, however, changes in renal blood flow, the glomerular filtration rate, hepatic blood flow, bile flow and pulmonary function may alter elimination of a drug in women during gestation.

Such differences have already resulted in sex-specific dosing. Desmopressin (Figure 5d), which activates vasopressin receptors in the kidneys to regulate water homeostasis, is such an example. Women have been found to be more sensitive to the antidiuretic effects of desmopressin than men due to the gene coding for the arginine vasopressin receptor. This gene is found on the X chromosome and in humans, several other genes involved in water homeostasis are located on the X chromosome [133]. As males only have one X chromosome, only one copy of the vasopressin receptor gene is likely to escape X chromosome related-inactivation, unlike in females, having two copies of the gene [134]. It has been reported that older females taking desmopressin are more likely to have lowered sodium concentration leading to unwanted side effects such as weakness, dizziness and fainting. To prevent adverse reactions, both the EU and Canadian medical agencies have recommended lower dosages of desmopressin be used by women [49]. Lower doses of desmopressin have also elicited good response in female paediatric patients [135], consistent with research in adults [134]. The drug is consequently marketed with different recommended doses on the labelling package for men

and women. A comprehensive portfolio of sex-specific differences in the pharmacokinetics is outlined in Table 2.

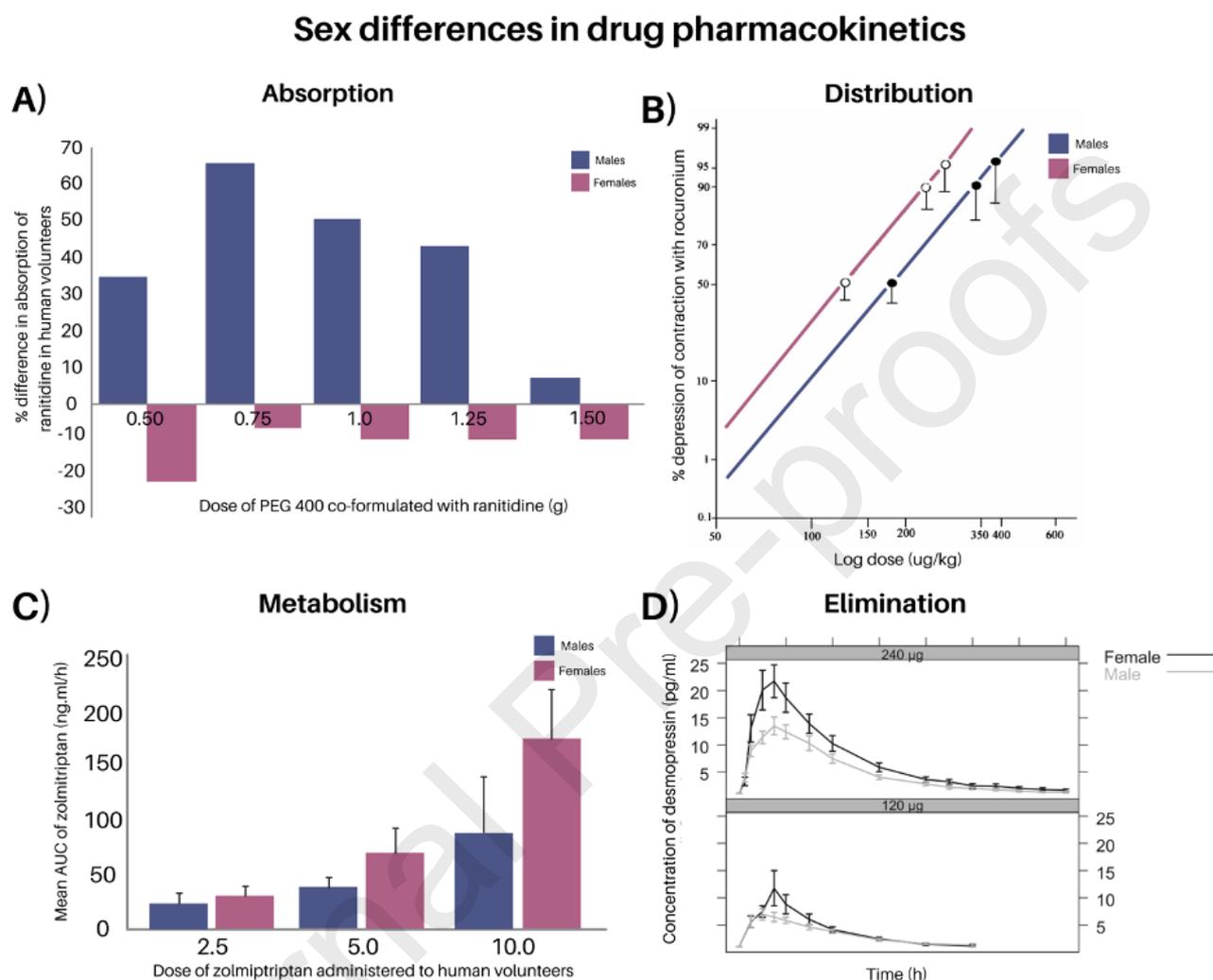


Figure 5. Hallmark examples of sex differences in drug pharmacokinetics. **A)** Sex differences in drug absorption have been observed with the co-formulation of different doses of PEG 400 with ranitidine in the human volunteers. Adapted with permission from [136]. **B)** Dose-response curves of rocuronium in male and female anaesthetised patients. Adapted with permission from [137]. **C)** Mean $AUC_{0-\infty}$ after 2.5 mg, 5 mg and 10 mg zolmitriptan in males and females. Adapted with permission from [119, 138]. **D)** Mean desmopressin concentration profiles by dose (120 μ g and 240 μ g) and sex. Reproduced with permission from [134].

Table 2. Sex-specific differences in the drug pharmacokinetics in humans and pre-clinical rat models following oral administration.

Drug	Sex differences in pharmacokinetic effect	References
Human data		
Aspirin	Higher clearance in females than males and shorter $t_{1/2}$ in females	[77]
Chlordiazepoxide	Higher AUC in males than females	[139]
Cefotaxime	Longer duration in females than males	[140]
Daidzein	Higher AUC ₀₋₄₈₀ in females than in males; no differences reported between pre- vs. post-menopausal females in $t_{1/2}$, t_{max} and AUC due to influence from gut microflora	[141]
Diazepam	Higher free fraction and larger distribution in females than males	[105]
Fentanyl	Lower urinary excretion in females than males due to a higher glomerular filtration rate in males	[123]
Fluconazole	Higher plasma levels in females than males due to sex differences reported in CYP3A4, CYP2C9, CYP2C19 expression	[142]
Flurazepam	Higher AUC of its major metabolites (N-1-hydroxyethylfurazepam and N-1-desalkylflurazepam) in females than males	[143]
Glucose	Fluctuating absorption levels in females than males; lower absorption is reported during the first hour but increased in absorption in the last hour of a three-hour oral glucose tolerance test in females	[144]
	Early glucose absorption lower in females with impaired glucose intolerance	[145]
Heparin	Longer duration in drug distribution in females than males	[146]

Levofloxacin	Higher C_{max} in females but larger AUC_{0-480} in males	[147]
Lignocaine	Higher distribution in females than males Plasma binding of lignocaine was almost completely attributed to changes in α_1 -acid glycoprotein concentration which is reduced by oestrogens	[148, 149]
Losartan	Larger AUC in females than males Lower total body clearance and volume of distribution in girls as higher content of water in men which can influence water-soluble drugs such as losartan	[150]
Methylprednisolone	Lower distribution, higher clearance and shorter $t_{1/2}$ in males than females Differences in IC_{50} values for cortisol secretion, basophil and helper T-lymphocyte trafficking are sensitive to methylprednisolone suppressive effects	[151]
Metoprolol	Lower distribution, higher clearance and shorter $t_{1/2}$ in males than females Stereoselectivity in oral clearance appeared to be greater in subjects with higher clearance (i.e. males) thus increasing metoprolol exposure in females	[152]
Metronidazole	Higher distribution in males than females	[153]
Midazolam	Lower AUC in females than males due to lower level of CYP3A expression in males	[154]
Ofloxacin	Higher AUC and C_{max} but lower total body clearance and volume of distribution in young females Significantly higher AUC_{0-480} in younger females than younger males	[155]
Quinine	Lower distribution, higher clearance and shorter $t_{1/2}$ in males than females	[140]
Rocuronium	Rocuronium is a lipid-soluble drug and a longer duration of rocuronium distribution was reported in females than males due to a higher content of fat	[104]

Torasemide	Lower clearance in females with higher AUC and C_{max} due to a higher glomerular filtration rate in males	[156]
Vecuronium	Longer C_{max} in females than males	[104]
Verapamil	Clearance of oral verapamil was accelerated in females, $t_{1/2}$ and mean residence times were significantly shorter in females than males Sex difference not evident when administered intravenously suggesting that intestinal processes likely influence sex-specific differences in drug clearance	[157-159]
Zolmitriptan	Higher bioavailability in females after both 5 mg oral dosing and intravenous dosing	[119]

Table 3. Sex-specific differences in the drug pharmacokinetics in pre-clinical rat models following oral administration.

In pre-clinical rat models

Drug	Sex differences in pharmacokinetic effect	References
Celastrol	Higher oral bioavailability in female rats due to altered mechanism in absorption and metabolism	[160]
Clindamycin	Higher plasma levels in female rats due to sex differences in CYP3A4, CYP2C9, CYP2C19 expression	[161]
Diltiazem	Higher C_{max} and longer t_{max} in female rats than males	[162]
Letrozole	Higher C_{max} and AUC_{0-480} in female rats; AUC and $t_{1/2}$ in females were 3-fold and 4-fold higher than in males Tissue/plasma drug concentration in female rats 24 h after dosing was significantly higher in female rats than in males in the heart, spleen, brain and genital glands	[163]
Letrozole	Letrozole metabolism more extensive in male rats	[163]

Nimodipine	Higher AUC ₀₋₄₈₀ and C _{max} in female rats	[164]
Ranolazine	Significantly higher plasma concentrations in female rats than male rats; C _{max} and AUC in female rats were roughly 2-to 3-fold greater	[165]
Ranolazine	t _{1/2} in male rats were shorter than in female rats	[165]
Schizandrin	Higher C _{max} and AUC ₀₋₄₈₀ in female rats	[166]
Taurocholate	Lower renal clearance in males	[167]

5.0 Adverse drug reactions in women

The occurrence of ADRs is 50 – 75% higher in females than men [168] and 60% of all patients hospitalised for adverse drug effects were women [169, 170]. This may be due to the interplay of differences in physiology, sex hormones, pharmacodynamics and pharmacokinetic response in the processing drugs. In addition, women may be more frequently overdosed and more commonly polymedicated than men [171]. Males and females also display different non-adherence behaviours. A cross-sectional questionnaire in the Swedish population did not find sex differences in the reporting of non-adherence. However, males were more likely to forget to take their dose or change their dose. Whereas in contrast, females were reported to collect their prescription medicine and not take it and omit their medication due to ADRs [172]. An extensive table of sex-specific differences in ADRs is outlined in Table 4.

Males and females appear to respond differently to pain and opioid analgesics. Women are reported to experience more severe postoperative pain and need a greater dose (+11%) of morphine than men postoperatively [173]. Greater analgesic effects were reported with opioid analgesic in females compared with males, with more adverse side effects than males [174]. In addition, pain response are more variable in females and more painful diseases are more commonly reported among females. Sex hormones and different density and modulation of the endogenous opioid system may contribute towards these sex differences [175]. Cepeda et al., showed that women had a 60% higher risk of nausea and vomiting than men following opiates use, though response did not differ between the sexes [176].

Largely ignoring female participants or not powering for sex in clinical trials has resulted in a distinct female-bias in ADRs [68], even to the point that pharmaceuticals have been withdrawn from the market due to a greater risk of side effects in women [71]. For example, Posicor (mibefradil dihydrochloride) approved for hypertension and angina, lowered the heart rate of elderly women and interacted with 26 other drugs [71]. Although the FDA outlined that both sexes should be represented in all phases in clinical trials to avoid undetected sex differences in drug efficacy and side effects [50, 177, 178], there is still a long way to go. Labots et al. conducted a cross-sectional, structured research into the publicly available registration dossiers of the FDA-approved drugs that are frequently prescribed. For 38 of the drugs where sufficient data was publicly available, a clear disparity in male and female representation between the phases of clinical trials was identified. For example, only 22% of female participation was

demonstrated in Phase I in comparison to 48% and 49% in Phase II and III trials respectively [179].

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Table 4. Drugs with adverse drug reactions experienced more commonly by females (risk factors and statistical differences provided in brackets). Adapted from [68]. The symbols F refers to females, M to males and ↑ to greater or higher.

Drug	Indication	Adverse drug reactions	Reference
Cardiovascular system			
Aliskiren	Hypertension	Diarrhoea (rates ↑ in F at a dose of 150 mg daily, whereas rates only ↑ in M at dose of 300 mg)	[180]
Amlodipine	Angina and hypertension	Oedema (F 14.6 % and M 5.6 %), flushing (F 4.5 % and M 1.5 %), palpitations (F 3.3 % and M 1.4 %) and somnolence (F 1.6 % and M 1.3 %)	[181, 182]
Clopidogrel	Atherothrombotic and thromboembolic events	Fracture (rates ↑ in F at low doses for all fractures and hip fractures, whereas rates only ↑ in M at higher doses), bleeding (risk ↑ in F [relative risk 1.40; 95 % CI, 1.00-1.96]), GI symptoms with IBD (control 60%, F 70 %, $p = 0.0003$ and M 61%, $p = 0.8312$)	[183, 184]
Dabigatran	Prophylaxis of thromboembolic events	Bleeding (↑ in F elderly, confounded by decreased renal function, low body weights & more drug-drug interactions)	[185, 186]
Digoxin	Atrial fibrillation and heart failure	Mortality from any cause (5.8% ↑ in F)	[187-189]
Dofetilide	Arrhythmia	Torsades de Pointes (risk was 3 times ↑ in F)	[190]
Pravastatin	Hypercholesterolaemia	Coronary heart disease (incidence ↑ in older F)	[191]

Propranolol	Thyrotoxicosis, migraine prophylaxis, arrhythmias, hypertension, angina and anxiety	Dizziness, muscle pain, headache and dry mouth (incidence substantially ↑ in F)	[192]
Torasemide	Oedema and hypertension	Hospitalisation (66% of cases occurred in F)	[193]
Warfarin	Anticoagulant	Major bleed (3.35 times more likely in F than M)	[194, 195]
Nervous system			
Aripiprazole	Schizophrenia	Heart rate (significantly ↑ bpm in F), elongated QTc (significantly ↑ in F than M) and nausea and vomiting (F 42.6% M 20.2%, $p = 0.037$)	[196]
Buprenorphine	Severe pain and treatment of opioid dependence	Sleep disturbance (M significantly less likely to report sleep disturbances than F)	[197]
Bupropion	Smoking cessation	EEG abnormalities (EEG sharp waves in F ↑ by a factor of 2.53 compared to M), seizure (F 1.5-fold ↑ likelihood)	[198, 199]
Carbamazepine	Epilepsy	Cognitive impairment (reaction time significantly more impaired in F), elevated LDL/HDL (significantly ↑ in F than M and control)	[200, 201]
Citalopram	Depressive illness, panic disorder	Elevated antidiuretic hormone (F is a risk factor)	[202]
Clozapine	Schizophrenia and psychosis in Parkinson's disease	Increase in blood glucose, type II diabetes (fasting blood glucose ≤ 6.0 mmol/L M 88%, F 41%, $p < 0.0001$), laxative use (F 49.1% and M 29.1%, $p < 0.01$), ileus (F odds ratio (OR): 1.60 confidence interval (CI): 1.10–2.31), neutropenia (F↑ OR 1.45 95% CI 1.28 to 1.67),	[203-212]

		leukopenia (F ↑, $p = 0.026$), obesity, weight gain (F +5.5% and M +1.3%, $p = 0.01$)	
Diazepam	Muscle spasm, anxiety, dystonic reactions, sedation and panic attacks	Psychomotor impairment (F reportedly felt clumsier)	[213]
Eszopiclone	Insomnia	Dysgeusia (lasted longer and more intense in F [66%] than M [53%])	[214]
Fluoxetine	Major depression	Hypercortisolemia (F 98% and M 68%), elevated albumin (F only 23%, $p < 0.05$), elevated tryptophan (F 83% and M 32%), suicidal ideation (F risk factor)	[215-218]
Gabapentin	Epilepsy and neuropathic pain	Dizziness, somnolence, nausea (probability F 0.6 and M 0.4, and F 1.9 times more likely to report ADRs)	[219]
Imipramine	Depressive illness and nocturnal enuresis	Dry mouth, constipation, sweating, tremor, treatment discontinuation (F 27.8% and M 11.5%)	[220]
Methylphenidate	Attention deficit hyperactivity disorder	Anxiety disorder (F 20.8% and M 5.9%)	[221]
Morphine	Pain	Respiratory depression (F 52% and M 32%), emesis (F 18% and M 0%), nausea (F 35% and M 3%)	[173, 222]
Nortriptyline	Depressive illness and neuropathic pain	Dry mouth (In a 6-week clinical trial, self-rated dry mouth was present for 6 weeks in F and 2 weeks in M)	[223]
Oxycodone	Pain	Nausea (F 24% and M 12%), pruritus (F 9% and M 5%), functional impairment (F $t[584] = 3.02, p < 0.01$), psychiatric severity (F $t[636] = 3.99, p < 0.001$)	[224-226]

Perampanel	Epilepsy	Dizziness, headache, treatment discontinuation (F 10.9% and M 6.8%)	[227]
Pramipexole	Nausea, fatigue	Nausea (F 20.8% M 6.7%), fatigue (F 10.5% and M 7.3%)	[228]
Risperidone	Schizophrenia, psychosis and mania	Hyperprolactinemia (F 127 ng/ml and M 54 ng/ml), headache (F 31% M 11%), hypotension (F 17% and M 0%)	[229-231]
Sertraline	Depressive illness, obsessive compulsive disorder, panic disorder	Cholesterol (F $\chi(2)(1) = 7.15, p = 0.008$), nausea (F 36.7% and M 21%), dizziness (F 19.3% and M 10.5%), delusions (F $t(257) = -2.10, p = .04$)	[232, 233]
Respiratory system			
Terfenadine	Antihistamine	Torsades de Pointes (F \uparrow susceptibility)	[71, 234]
Infection			
Erythromycin	Antibiotic	Torsades de Pointes (F 58% and M 32%)	[235]
Primaquine	Malaria	Nausea (F 35% and M 12%)	[236]
Endocrine system			
Liraglutide	Diabetes	Headache, vomiting, nausea (F 44% and M 6.3%, $p = 0.02$)	[237]
Prednisone	Steroid – inflammatory conditions	Depression (F 24.4% and M 16.1%, $p = 0.09$), fatigue (F 34.4% and M 29.2%, $p = 0.6$), hair loss (F 28.1% and M	[238]

Rosiglitazone	Diabetes	3.6%, $p < 0.0001$), mood swings (F 43.1% and M 30.7%, $p = 0.03$), weight gain (F 68.8% and M 56.2%, $p = 0.03$) Fractures (F OR 2.23, 95% CI 1.65–3.01; $p < 0.001$)	[239]
Genito-urinary system			
Trospium	Urinary frequency, urgency and incontinence	Cognitive impairments (F 2 times more likely)	[240]
Malignant disease			
Capecitabine	Cancer	Dose-limiting toxicity (F 68% and M 52%)	[241]
Fluorouracil	Cancer	Stomatitis, leukopenia, alopecia, diarrhoea, mucositis	[242, 243]
Paclitaxel	Cancer	Lower lesion revascularization (F 11.5% and M 22.6%, $p < 0.001$)	[244, 245]
Musculoskeletal system			
Infliximab	Inflammatory diseases	Allergic reactions (F 38% and M 22%, $p = 0.009$; OR 2.2, 95% CI 1.2-4.1)	[246]

6.0 Sex differences in pharmacodynamics

Pharmacodynamic differences based on sex have not been reported as extensively as pharmacokinetic differences due to difficulties in the quantification of such effects. Pharmacodynamic differences occur when the same plasma concentration of a drug in both sexes does not cause the same pharmacological response between the sexes [247]. Signalling pathways are believed to be similar in structure between the sexes, but differently expressed and regulated by sex hormones [248]. Soldin and colleagues proposed that women may be more pharmacodynamically sensitive than men [171], with sex differences in the binding affinity and number of receptors and differences in signal transduction pathway following receptor binding.

A significant pharmacodynamic sex difference is the increased prevalence of QT interval prolongation in women which is reported for many drugs (Table 4), leading to an increased incidence of ventricular tachyarrhythmias, syncope and increased risk of the cardiac arrhythmia Torsades de Pointes. An early study investigated 32 cases of arrhythmia induced Torsades de Pointes and found 70% of all cases occurred in women [249]. The antiarrhythmic agent, quinidine, shows greater QT prolongation in females than men for the same plasma concentration [250]. Several drugs were removed from the market due to this sex-specific pharmacodynamic effect which includes terfenadine, astemizole and cisapride [71]. The ICH E14 guidance [251] requires all new compounds to be tested for effects on the QT interval according to the Thorough QT (TQT) protocol, with standardised approaches for males and females. In mice, testosterone appears to be the main influence for lower risk of Torsades de Pointes which increases the rapid repolarisation of potassium channels [252].

Dofetilide, another antiarrhythmic agent, can increase the risk of QT prolongation in males and showed a 14 – 22% higher exposure in females compared with males, after adjustments for weight and creatinine clearance [250]. Higher dofetilide exposure in females may be due to lower creatinine clearance with higher sensitivity and longer QT interval at baseline. Sex differences exist in cortisol pharmacodynamics with women showing increased sensitivity to cortisol suppression and may therefore be more sensitive to basophils and helper T lymphocytes [253]. Sex differences were also found for the pharmacodynamics of prednisolone with a significantly smaller 50% inhibitory concentration (IC_{50}) value, which may be mediated

by endogenous oestrogens with increased sensitivity found at higher oestradiol concentrations [151].

Sex differences in pharmacodynamics have been investigated in anaesthetic agents response with males showing a 30 – 40% greater sensitivity to the effects of propofol than females [254]. Females showed an increase pharmacodynamic sensitivity to diazepam [213]. Women were reported to require a smaller dose of olanzapine to achieve a 70% binding of the dopamine D₂ receptor for therapeutic efficacy [255]. Adjusting for weight, height, age or concomitantly administered medicines did not affect olanzapine clearance and testosterone and/or oestrogen may modulate the pharmacodynamics of olanzapine [256].

7.0 Female-specific states that affect oral drug performance

Fluctuations in endogenous steroid sex hormones naturally occur throughout the menstrual cycle, pregnancy and in the transition towards menopause; such continuous variation in biological females has the potential to manipulate drug efficacy and ADRs [257]. Women can also take exogenous hormones for use as contraceptives and for the symptoms of the menopause. A bi-directional relationship can be observed; exogenous hormones can influence other drug products by altering metabolism whilst at the same time, drug metabolism pathways may also impact exogenous hormones used for therapy. Ritz et al., suggest adding a single sex hormone, oestrogen or testosterone for example, to *in vitro* cell cultures to investigate the effect on a particular outcome. The action of sex steroid hormones can exert epigenetic changes such as DNA acetylation and methylation on cell behaviour which can cause sex differences in human physiology and consequently on the mechanism of drug action [258].

7.1 Menstrual cycle

The menstrual cycle is a 28-day process and can be divided into three distinct phases; the follicular, ovular and luteal phases whereby plasma hormone concentration widely varies due to fluctuations in oestradiol and progesterone concentrations. High levels of both oestradiol and progesterone can encourage water retention which may affect body composition through the menstrual cycles [259, 260]. An overview of the known physiological variations that occur

during the menstrual cycle which may impact drug pharmacokinetics is shown in (Figure 6), although several findings need further research.

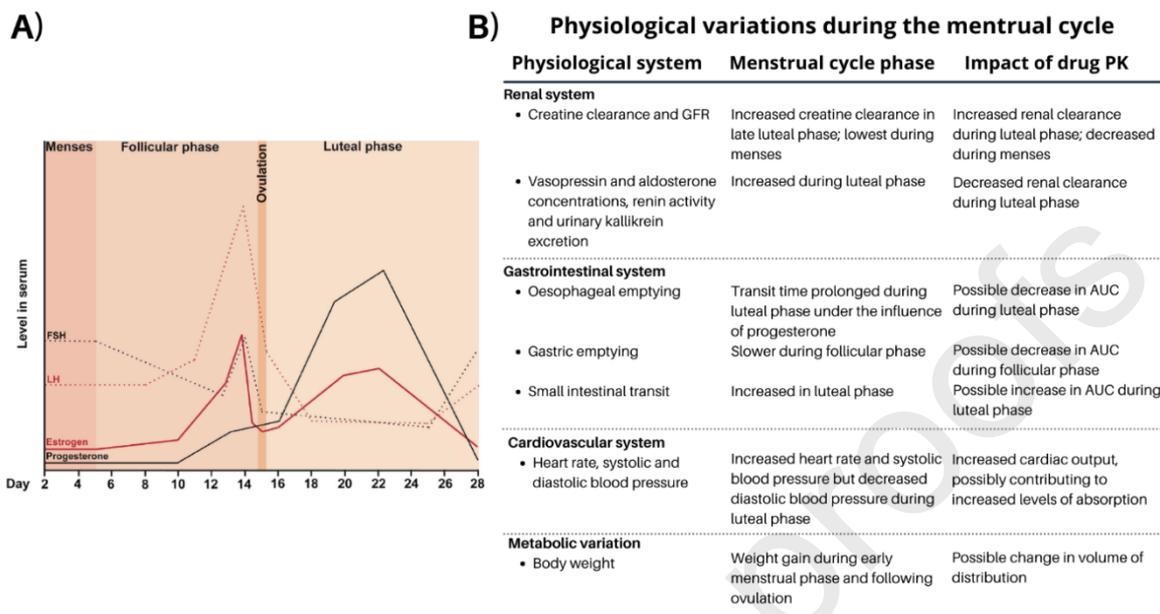


Figure 6. A) Endogenous hormone changes during the menstrual cycle. B) Physiological variations during the menstrual cycle and its potential influence on drug pharmacokinetics. Adapted with permission from [261].

Females taking ibutilide were reported to be more susceptible to QT prolongation particularly during the ovulatory phase of the menstrual cycle, compared with the luteal phase [262]; testosterone has been proposed to have protective effects by shortening the action potential duration and diminishing the QT response as seen in the luteal phase [263]. The effect of hormones is suggested for pharmacokinetics, pharmacodynamics and ADRs sex differences, although contrary to expectations, the activity of many drugs is not influenced by the menstrual cycle [264, 265].

7.2 Pregnancy

Evidently, several physiological changes can occur during the gestation which affect drug pharmacokinetics including i) volume of distribution: increased plasma volume, extracellular fluid space and total body water; ii) cardiovascular systems: plasma volume expansion, increased in carbon monoxide, altered regional blood flow (i.e. increased uterine, renal, skin and mammary blood flow but decreased skeletal blood flow), increase in stroke volume and

heart rate; iii) respiratory changes: compensated respiratory alkalosis; iv) decreased plasma albumin and; v) GI absorption: altered activity in uridine diphosphate glucuronosyltransferase (UGT) isoenzyme [116]. Changes in endogenous hormones associated with pregnancy are known to modify drug efficacy and with some drug products, may have adverse effects of foetal development. 64% of pregnant women take medication and is expected that two thirds of which may not have been tested in pregnant women [266, 267].

Pregnancy is able to modify the distribution and clearance of drugs due to the increase of blood and extracellular fluid volume. Hormonal changes, however, are further capable of influencing enzyme activity; for example, both CYP2C19 and CYP1A2 activity is decreased during pregnancy with the latter affecting the drug metabolism of caffeine and theophylline [268]. An increase in CYP2C9, CYP3A4 and UGT1A4 activity, however, is observed in the second and third trimester of pregnancy which will affect the drug processing of such CYP substrates. The effect of hormonal changes with pregnancy on drug transporter genes, however, is not fully understood but may involve the activity of oestrogen and androgen receptors [269].

7.3 Oral contraceptives

Oral contraceptives are the most common form of contraception between the ages of 15 – 49 years old, with 28% using its as their main method of contraception [270]. Most oral contraceptives contain a combination of oestrogen and progesterone to suppress ovulation and luteinising hormone secretion, respectively [271]. These exogenous hormones can impact the metabolism of a multitude of other medications through the inhibition of multiple cytochrome P450 enzymes, with moderate inhibition of CYP1A2 and CYP2C19 and weak inhibition of CYP3A4 [272-275]. Decreased CYP activity as a result of oral contraceptives is believed to be due to competitive inhibition. Although there is some evidence that oestradiol may downregulate CYP2C19 expression by the interaction of oestrogen receptor- α with a binding site in the CYP2C19 promoter [269]. A study found that oral contraceptive-induced CYP2B6 inhibition led to higher plasma concentration of bupropion, partly metabolised by CYP2B6 [276]. For the commonly administered drug product ibuprofen, the phase of the menstrual cycle did not affect the pharmacokinetics of *S*-ibuprofen or *R*-ibuprofen, but women treated with oral contraceptives had lower AUC and higher clearance than women not taking oral contraceptives [277]. Significantly, the use of oral contraceptives is not often considered in the prescribing of concomitant medicines.

7.4 Menopause

Menopause is the natural permanent cessation of menstruation, after the loss of ovarian follicular development [278]. With menopause, circulating oestrogen can decrease by up to 90% [279]. Women use exogenous hormones, such as combined oestrogen and progestogen tablets, skin patches and gels to control the symptoms of menopause which include hot flashes, night sweats, sleep disturbances and vaginal dryness. Adipose tissue and skin become the predominant source of oestrogen, where androgens are converted to oestrogen by aromatase, encoded by *CYP19A1* [279]. Changes to other drug metabolising enzymes are reported, such as a 20% reduction in the activity of intestinal CYP3A4 [280]. Conflicting results are found in the literature on pharmacokinetic changes in women relating to menopausal status. Several studies on the pharmacokinetics of erythromycin and prednisolone in pre- and postmenopausal women, considering changes in intestinal or hepatic CYP3A4 activity, found no significant differences in drug metabolism according to menopausal status [281].

Reports suggest that postmenopausal women respond differently to antidepressants compared with premenopausal women [232], for example, the response of postmenopausal women to antidepressant treatment was in general worse than those in premenopausal women showing an association with high basal levels of follicle-stimulating hormone (FSH) in the postmenopausal women [282]. Sex hormones are known to interact with serotonergic, noradrenergic and dopaminergic systems [283].

7.5 Transgender people

Transgender is a term for those whose gender identity differs from the sex assigned at birth [284]. Long-term testosterone or oestrogen treatments are standard practice for transgender people, taken to align secondary sex characteristics with gender identity [285]. Due to potential physiological differences related to the XX or XY chromosomes and the various effects of administering endogenous and exogenous hormones, it can be difficult to predict the drug response in transgender people [267]. There are few studies on hormone-drug interactions in transgender patients [286]. Clinicians may use drug-drug interaction data from the general adult population. However, this does not consider the pharmacodynamics effects of hormone therapy [287].

The limited investigations have however found that in transmen with XX chromosomes who followed testosterone treatment, increases in serum triglycerides and low-density lipoprotein cholesterol were found which caused an increased risk of venous thromboembolism [288, 289]. For transwomen with XY chromosomes who used exogenous oestrogen treatment, there was a reported increase in the risk of stroke and myocardial infarction [290, 291]. Trans patients who receive sickle cell disease treatment combined with hormone therapy poses a challenge as the symptoms of sickle cell disease and the side effects of hormonal therapy can both cause cardiovascular complications [286]. In transgender women with human immunodeficiency virus (HIV), antiretroviral therapy can interact with oestradiol, which may result in lower rates of virologic suppression and higher HIV-related mortality [292]. In addition, trans patients may require adjustments prior to starting steroid hormones [267]. The authors direct the reader towards a recent comprehensive review article on the clinical pharmacology in transgender people, which includes pharmacokinetic and pharmacodynamics considerations associated with hormonal treatments [287].

8.0 Sex differences in Bioequivalence Studies

Generic formulations are by far the most prescribed drugs [293]. Before entry into the pharmaceutical market, the manufacturer of the generic drug is expected to prove bioequivalence (BE) to the marketed, reference drug [293]. This is achieved by comparing the systemic AUC of the generic formulation to that of the reference drug in a crossover clinical trials design with individuals acting as their own control. To achieve BE, the AUC and peak concentrations of the generic drug need to be within 80 – 120% of the reference drug [294]. In BE studies, the US FDA guidelines states that ‘if the drug product is intended for use in both sexes, the sponsor should attempt to include similar proportions of males and females in the study.’ [295]. Unsurprisingly, BE studies, however, are typically conducted in healthy, young adult male volunteers [296]. The argument in favour for carrying out BE studies exclusively in males is that though the sexes may pharmacokinetically respond differently, there is an assumption that intra-individual variabilities in BE are similar between males and females.

Research from the FDA Center for Drug Evaluation and Research reviewed 26 BE studies submitted to the FDA which compared original and generic drug formulations in men and women. The study found that five generic drug products (22%) were statistically different

between the sexes with respect to variability in AUC and in four (18%) variability in peak plasma concentration (Table 5) [297]. The incorporation of sex as a biological variable will have major implications for the management and interpretation of BE studies. The BE results of alprazolam in men show negligible intrasubject variability, i.e. a small number of individuals will only be needed to show BE. However in women, intra-subject variability in alprazolam was 6-fold. This consequently means that a larger number of individuals will need to be studied in order to power for BE. This distinct drug example is one of many to show that drug variability is much larger in women, precluding the ability to generalise results from men to women.

The fluctuating hormonal status of females along the menstrual cycle may also affect drug pharmacokinetics. Ranitidine, for example, is subjected to varying pharmacokinetic response according to menstrual period. In the follicular phase, AUC was 7312 ng/ml/h although was 29% lower in the luteal phase at 5195.83 ng/ml/h. In men, AUC was 11,471.94 ng/ml/h [298]. This highlights that studies of BE of drugs targeting women must compare the reference and generic drug formulation during similar stages in the menstrual cycle.

Table 5. Drugs with statistically significant sex differences in bioequivalence. Adapted with permission from [297].

Drug	Variability in AUC (%CV)		Variability in C _{max} (%CV)	
	Males	Females	Males	Females
Alprazolam	4.9	29.4	-	-
NAPA	9.0	4.4	-	-
Nitroglycerin	21.3	39.5	13.6	24.4
Phenylacetate	4.3	9.9	6.1	17.4
Cimetidine	-	-	26.8	11.8
Ketoprofen	-	-	22.2	51.5

AUC, area under the concentration-time curve; C_{max}, peak plasma concentration; CV, coefficient of variation; NAPA, *N*-acetylprocainamide

8.1 Sex differences in excipient effects

In generic drug manufacturing, not only is the drug synthesis process likely to be different to that of the reference drug product, but the formulation itself. As such, a generic drug may differ

in concentrations or nature of pharmaceutical excipients from the reference formulation. Studying only men in BE studies requires the hypothesis that both males and females respond to excipients similarly. Prior to the 1990s, many excipients were generally regarded as inert with the majority comprising molecules that were structurally simple, biologically inactive and of natural origin such as wheat, minerals and sugars. However, in the present day, the number of excipients has substantially increased with over 1000 excipient types from 40 functional categories being used in commercialised drug products [299]. Excipients are considered a reliable source of safe chemical matter, co-formulated only to carry out their intended function in a dosage form (i.e. disintegrants, glidants, lubricants, binders, taste masking and colouring agents). However, an exponentially growing body of research and clinical reports contest their biologically inert character [300]. In addition, it is also being found that excipients affect males and females differently which further complicates BE studies and suggests that certain populations may experience adverse reactions to pharmaceutical excipients more commonly [299].

For example, polyethylene glycol (PEG) 400, a commonly used solubility enhancer, is osmotically active at pharmaceutically relevant concentrations. Following a 10 g dose, PEG 400 reduced GI transit time by 35% [301] and when co-formulated with ranitidine, accelerated small intestinal transit which consequently reduced drug absorption by 31% [302]. A further study evaluated the effect of different PEG 400 concentrations (1 g, 2.5 g and 5 g) on liquid GI transit and ranitidine bioavailability and found ranitidine bioavailability to be reduced by 38% at higher PEG concentrations, which was thought to be due to PEG 400 stimulating GI motility [302]. Conversely, at the lowest concentration (1 g), drug bioavailability was significantly increased by 41% possibly due to the modulation of intestinal permeability. These studies, however, were conducted in male participants.

The activity of PEG 400 was then investigated in a human study conducted in both males and females at pharmaceutically relevant doses. In male volunteers, the co-formulation of PEG 400 at 0.5 g, 0.75 g, 1.0 g, 1.25 g or 1.5 g increased bioavailability by 34%, 63%, 49%, 43% and 6% in comparison to the control treatment of ranitidine alone. At equivalent drug-excipient formulations administered to female subjects, however, ranitidine bioavailability decreased by 24%, 8%, 13%, 13% and 13% against the control. As such, all doses of PEG 400 enhanced the bioavailability of ranitidine in male subjects but not in females. The most pronounced effect was noted with the 0.75 g dose of PEG 400 attributed to a 63% increase in bioavailability in

males [136]. Recent research has shown that the bioavailability-modifying effect of PEG 400 is not only seen in the presence of ranitidine but extends to another BCS Class III drug, cimetidine. Cimetidine co-formulated with PEG 400 at 0.5 g, 0.75 g, 1.0 g and 1.5 g significantly increased cimetidine bioavailability in male participants by 34%, 58% and 41% respectively. No such enhancement, however, was seen in female participants, similar to what was observed in the presence of ranitidine. At 1.5 g PEG 400, however, both sexes displayed a reduction in ranitidine bioavailability [303] (Figure 7).

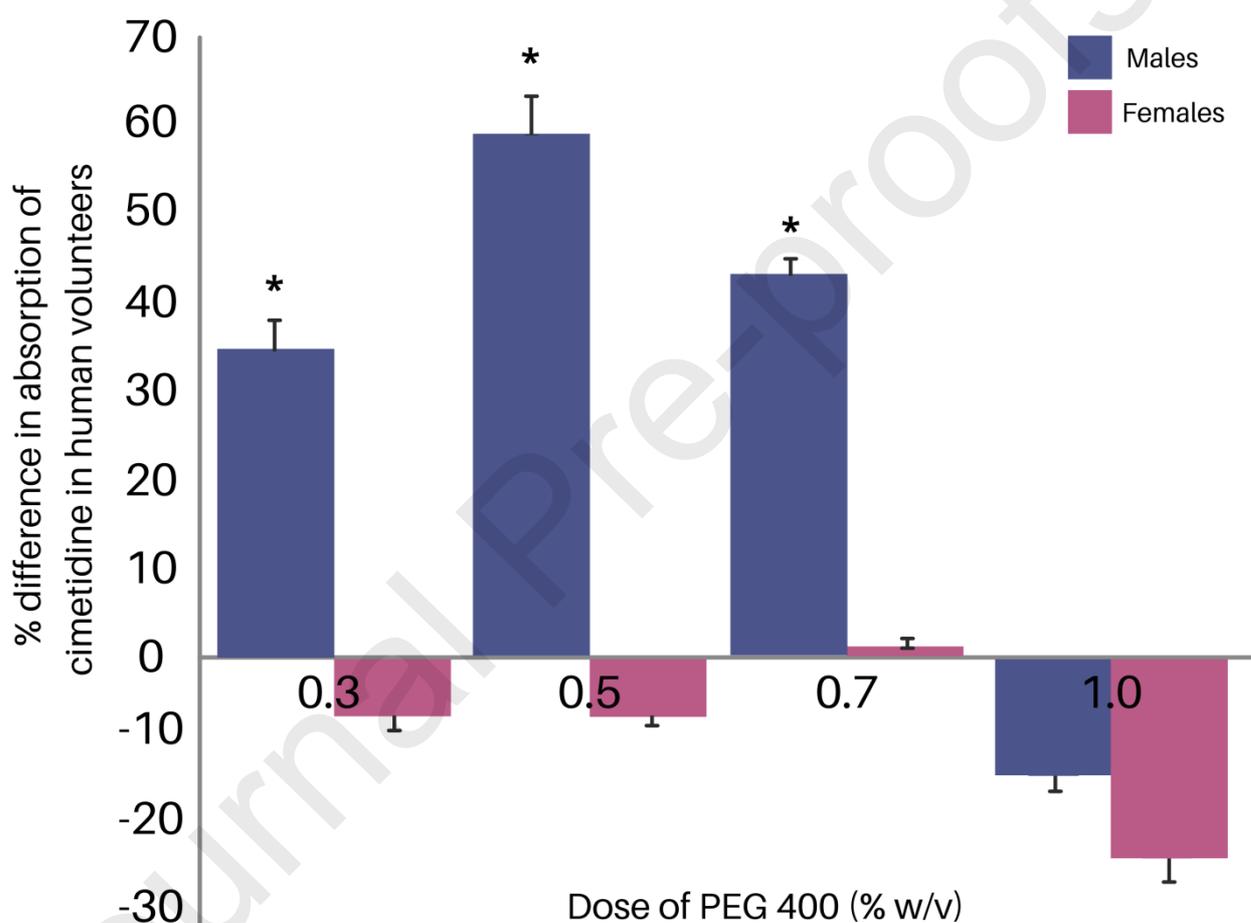


Figure 7. Percentage difference in cimetidine bioavailability when co-formulated with different doses of PEG 400 against the control (cimetidine alone) in male and female human volunteers (mean \pm S.D., $n = 6$). *Denotes statistical significance against the control ($p < 0.05$). Adapted with permission from [303].

As such, if a ranitidine and cimetidine BE study were to be conducted in men only, the dose of the generic product if using PEG 400 in its generic drug formulation would be significantly different when compared with the reference drug. The above examples illustrate very clearly

that, for any drug product, BE in women cannot be extrapolated from BE studies conducted in males only, i.e. if a generic formulation is to be taken by women, it must be tested with sufficient power in women as well as men [304].

8.2 Sex differences in food effects

Sex differences exist in the processing of food. For example, female bodies tend to take longer to digest food. As aforementioned, the concomitant intake of food and drugs may affect the oral drug performance between the sexes. In preclinical and clinical studies, diets affect weight, metabolism, hormone and immune function, therefore diet formulation should be stated [304, 305]. Following a food intake, the secretion of acid is significantly higher in males than females [306].

Animal models such as the rat are used in preclinical studies, although potential sex differences are often not considered by pooling males and females or using male animals due to ease of handling and faster elimination of some drug products. Dou and colleagues recently reported sex-differences in the P-gp expression in both prandial states in rats. In the fasted state, male rats exhibited a significantly higher P-gp expression than females. In the fed state, however, the P-gp expression was significantly higher in females, 77% higher in the jejunum than the male counterparts [307]. Sex differences were also identified in male and female human jejunal and ileal tissues via mRNA and protein quantification via real-time polymerase chain reaction (RT-PCR), Western blot and liquid chromatography-tandem mass spectrometry (LC-MS/MS) respectively. Small intestinal P-gp was higher in human males than females with an increasing trend from the proximal to distal regions which was closely reflected in a pharmaceutically common preclinical model, Wistar rats [89]. Additionally, an *ex vivo* Ussing chamber found that the P-gp substrates ganciclovir and ranitidine demonstrated sex differences in their intestinal permeability [308]. Sex differences in the bioavailability of cyclosporine A, a P-gp substrate, was reported after a fat-rich meal; decreased bioavailability in females and increased bioavailability in male humans [309]. Diets rich in phytoestrogens, a component in soy, which is often included in rodent diet, may have sex-specific effects of cardiac health. In male humans, soy-based diets significantly decreased cardiac function and associated heart failure, observed to a lesser extent in females [310].

9.0 Suggestions for sex-informed scientific approaches

Historically, drug development has followed a ‘one size fits all’ approach. The incorporation of sex-informed perspectives, however, increases rigor, promotes drug discovery and expands the relevance of biomedical research. Thoughtful and deliberate methodology can improve study design and progress towards identifying potential sex differences in research. Promoting sex-as-a-biological-variable approaches in drug prescribing can start with relatively simple yet powerful steps with the use of female and male cells, tissues and organisms throughout the preclinical and clinical drug development, powering for any sex-related influences to be determined. Drug development should also consider the physiological nuances between males and females for effective drug delivery and the active inclusion of women of childbearing age and of pregnant women in drug clinical trials and diagnostic tools (Figure 8).

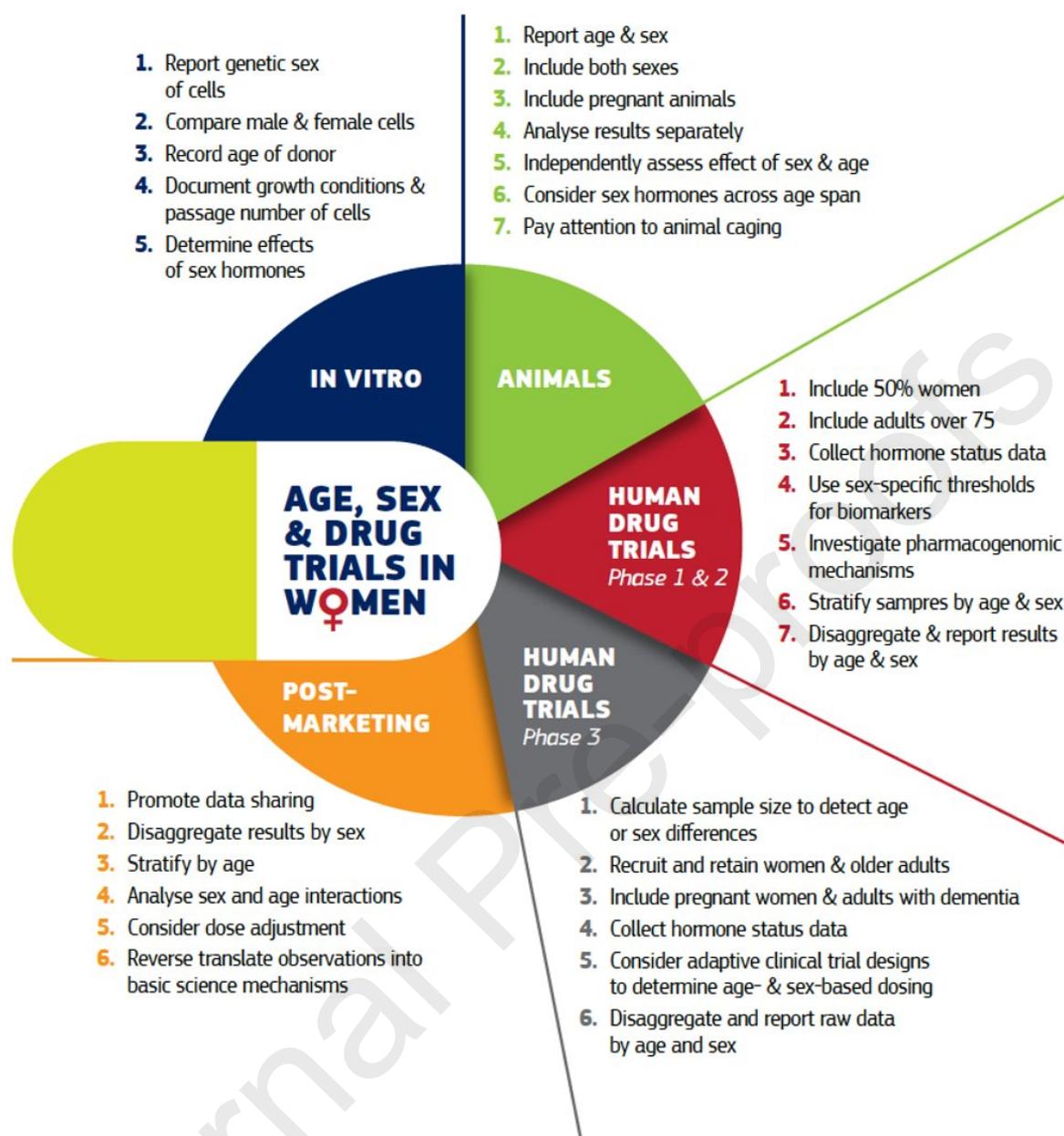


Figure 8. Suggestions towards sex-specific approaches in preclinical and clinical research, and post-translational approaches in drug development. Reproduced with permission from [311].

9.1 Sex-specific recommendations towards pharmaceutical research

If female cells and animals are not included in early phases of drug development, sex-specific differences in efficacy and toxicity will not be detected. Female and male cells are affected by their sex chromosomes and influenced by hormones in their environment. Certain types of cells such as those found in the liver produce different amounts of metabolic enzymes (Table 6). If sex differences are not considered in preclinical trials, experimental results can be irreproducible. As such, analysing the cellular response to medicines in a sex-specific manner

can offer early indications of potential differences that could influence the subsequent processes of drug development [312, 313]. If women are not included in early clinical trials, real-world effects of a medicine, such as adverse side effects, will subsequently not be detected before its release into market. It is known that women experience more unwanted side effects than men and the magnitude of the problem is difficult to assess as many countries have not included sex information in their statutory reports of side effects [314]. New reporting guidelines such as Prisma-Equity Extension [315] and Consort Equity 2017 [314] advocate for the disaggregation of data by sex in large comparative studies and meta-analyses aimed at predicting unwanted side effects better.

Table 6. Most commonly used cell lines in the pharmaceutical preclinical arena for drugs and formulations intended for oral administration. Adapted from [316].

Cell line	Sex	Description	Species	Year derived	Origin
A6	Male	Kidney epithelial	Xenopus	1965	Non-cancerous tissue
AGS	Female	Stomach epithelial	Human	1979	Cancerous tissue
AML-12	Male	Liver epithelial	Mouse	1994	Non-cancerous tissue
C2BBel	Male	Colonic epithelial cell (a Caco-2 subclone)	Human	1988	Cancerous tissue
Caco-2	Male	Colonic epithelial	Human	1977	Cancerous tissue
Capan-1	Male	Pancreatic epithelial	Human	1N/A	N/A
CFPAC-1	Male	Pancreatic epithelial	Human	1990	Cancerous tissue
CV-1	Male	Kidney fibroblast	African green monkey	1964	Non-cancerous tissue

H4TG	Male	Liver epithelial	Rat	1964	Cancerous tissue
HEP 3B	Female	Liver epithelial	Human	1983	Cancerous tissue
HEP G2	Male	Liver epithelial	Human	1994	Cancerous tissue
HK2	Male	Kidney epithelial	Human	1994	Non-cancerous
HPAF-II	Male	Pancreatic epithelial	Human	1982	Cancerous tissue
HT-29	Female	Colonic epithelial	Human	1964	Cancerous tissue
HuTu80	Male	Duodenal epithelial	Human	N/A	Cancerous tissue
IEC-6	Male	Small intestinal epithelial	Rat	1978	Non-cancerous tissue
KATO III	Male	Gastric carcinoma mixed	Human	1978	Cancerous tissue
LLC-PK1	Male	Kidney epithelial	Human	1977	Cancerous tissue
LS 174T	Female	Colonic epithelial	Human	1976	Cancerous tissue
MDCK	Female	Kidney epithelial	Dog	1958	Non-cancerous tissue
MIA-PaCa-2	Male	Pancreatic epithelial	Human	1975	Cancerous tissue
MKN45	Female	Gastric carcinoma	Human	N/A	Cancerous tissue

9.2 Sex-specific recommendations towards the application of physiologically based pharmacokinetic (PBPK) modelling and simulation

Analyses have found that the majority of attrition in drug candidate is due to unfavourable pharmacokinetic behaviours [317]. Therefore, prediction of the absorption, distribution, metabolism, and excretion properties could help to de-risk the drug development pathway. Physiologically based pharmacokinetic (PBPK) models can be used to predict inter-individual variability in the pharmacokinetic profile of drugs between the sexes. Here, female or male-specific physiological and pharmacokinetic differences as well as drug-specific physicochemical data can be inputted from *in vitro* and *in vivo* preclinical and clinical data. These physiological sex differences can be calibrated in the PBPK models by adjusting absorption values (fraction absorbed and intrinsic clearance), rate constants, scaling factors and enzyme or transporter activity coefficients. Parameter sensitivity analyses can be tested to understand if sex-specific parameters are required to accurately predict drug response between the sexes. The population simulator feature of PBPK software can be harnessed to alter the variance associated with input parameters to show the sex differences [317, 318]. A goal of the model construction would be to provide insight into the right dose for the patient. Since the 1990s regulatory agencies such as the FDA and the EMA have encouraged pharmaceutical companies to use PBPK modelling to understand drug response. Model parameters are chosen to reflect the inter-individual variability in the physiology of patient groups, in this case, male and females.

The development of sex-specific PBPK models requires quantitative data for the physiological sex differences. However, in the literature, sex differences are often presented as a relative comparison (for example, males > females, males < females, males = females) [319]. The successes of machine learning (ML) stem from its capability to discern patterns from complex and large volume data sets [320, 321]. In the drug discovery arena, artificial intelligence (AI) is facilitating research and development in drug candidate selection in larger pharmaceutical companies [322] as well as start-ups such as Google's DeepMind [323]. However, within drug product development and the field of prediction of pharmacokinetics, pharmaceutical companies are yet to realise the potential of AI.

Personalised medicines require an understanding of inter-individual differences in drug response [324]. AI has been applied to assess patient response to oncology therapeutics [325], drug-drug interactions [326] and ADRs [68, 327]. However, the majority of ML models fail to account for sex and its influence on disease and therapeutic outcomes. Therefore, the results may be discriminatory and be sex bias. For example, the US Department of Veterans Affairs

healthcare system was used to assess the risk of acute kidney injury [328]. Female patients comprised of 6.37% of patients in the dataset and therefore algorithm performance was lower in the females, compared with the males.

Drug product development is increasingly being guided by Big Data, with large data sets computationally analysed to reveal patterns, trends and associations [329]. Clinical and pharmaceutical Big Data has the potential to provide untapped insights into health and disease, as well as to explore sex differences [324]. However, the majority of genome-wide association studies concentrate on white male subjects, ignoring potential sex differences in diseases [330]. Biomarkers are increasingly being explored to facilitate detection and diagnoses. The FDA has recently approved the use of a number of digital biomarker devices that monitoring symptoms and measurables in clinical trials [331]. Ramsey et al., found that the concentration of 56% of biomarkers varied between males and females, concluding that sex and female hormonal status should be reported when collect biomarker-related data. AI models should incorporate sex and gender differences, so effective personalised medicines and tailored treatment plans can be recommended [332]. Furthermore, algorithm validation, regulation, explanation and interpretation must be ensured as much as possible [333].

9.3 Sex-specific recommendations towards personalised medicines

Reductionism in the biomedical research has resulted to the identification and mutations in the human genome. Omics-sciences can uncover the intricacies of healthy and disease pathways, particularly of defective molecules or specific cellular phenotype responsible for the latter. Sex-specific research is a promising field in which genomics, epigenomics, transcriptomics, proteomics and metabolomics can be applied to investigate the mechanistic reasons responsible for sex-related differences in complex multi-factorial disease pathophysiology and even drug response [334]. By using multi-omics approaches, information on sex differences in gene expression and protein levels can be understood, and successfully applied towards the development of sex-specific pharmaceutical therapeutics and medical devices.

Typically based on a 'one size fits all' concept, traditional manufacturing processes are unsuitable for the production of personalised drug delivery therapies involving labour-intensive, dose-inflexible and time-consuming processes [335, 336]. Due to recent manufacturing innovations and technologies, however, the number of drugs and treatments

available for individualised therapies has increased nearly 10-fold from 13 to 113 [337, 338]. Continuous manufacturing and additive manufacturing, for example, has transformed the healthcare industry towards tailored medicines development [339]. Specifically, three-dimensional (3D) printing, an additive manufacturing technique, is set to be a major disruptive technology in healthcare by the formation of bespoke intricate objects of virtually any shape and size, layer by layer [340, 341]. Structures can be created from a digital 3D file using a computer-aided design (CAD) software to readily manufacture objects individualised to each patient [342]. Since its introduction nearly three decades ago, 3D printing has transformed manufacturing abounding all fields and applications. 3D printing, however, is set to become a revolutionary technology within the healthcare space [343]; from its capability to create individualised objects, personalised medical prosthetics, implants and devices can be tailored to the individual needs of the patient. In the arena of drug delivery, various constructs have already been prepared using 3D printing [344-346] from drug-eluting implants and personalised solid oral dosage forms with characteristics such as increased patient acceptability, orally dissolving tablets, modified drug release, polypills and novel therapeutics for orphan diseases [347-352]. This technology has been explored as a viable method for personalising medicines at the point of use with a view to expand into rapid throughout screening of new drug candidates on 3D printed-biological tissues to identify intra-individual therapeutic responses [353]. Majority of the research of 3D printing pharmaceuticals or medical devices have focused on formulation characteristics and drug performance. Less efforts, however, have been invested in larger variables of personalisation such as sex-specific formulations in terms of the wider patient population. Due to its innate unique proposition in delivering personalised medicine, 3D printing could be employed to manufacture sex- and dose-specific oral drug products to limit side effects whilst providing optimum therapeutic effect according to the individual.

9.4 Sex-specific recommendations towards regulatory agencies

Bridging the sex-gap between drug development and patient care is not possible without approval from regulatory agencies. As aforementioned, a number of guidance documents published by the FDA and NIH amongst others have formalised the inclusion of women and minority groups to be included in all clinical research. The British Journal of Pharmacology also recommends that all future studies either include both sexes in experimental designs or provide explanation as to why sex or gender perspectives are not relevant for their research

methodology [312]. Other journals and funding agencies have followed suit and adopted similar policies to promote sex analyses in drug development [76]. Despite this, sex inequality still remains. As such, regulatory agencies should *require* and not simply *recommend* sex-disaggregated data reporting of all drug trial results submitted by the pharmaceutical industry. Regulatory bodies should also ensure that sex-specific information is available to prescribers and patients on drug websites and labels. In addition, as many pharmaceuticals in the market were tested and approved in years when women were not appropriately included in clinical trials, post-market surveillance is the only avenue to obtain data on sex differences in efficacy and toxicity. Post-pharmaceutical surveillance should also disaggregate side effect reporting between males and females to identify sex-specific drug responses.

10.0 Conclusion

Recent governmental policies mandate that researchers across the drug development pipeline should collect and analyse data by sex. It is clear, however, that the onus to incorporate the study of sex differences is on investigators to address these perspectives adequately and accountably at all levels of basic, clinical and population research. In human clinical trials, it should be an imperative to investigate and aggregate data according to sex. A focus of sex differences in the innovation process will further illuminate fundamental, modifiable causes of disease and highlight potentially significant findings in optimum drug efficacy and importantly, toxicity. If sex as a biological variable is skilfully addressed and powered in experimental designs and analyses, this will decrease the prevalence of patients experiencing adverse drug reactions, better treatment options and may give rise to new insights for men and women that will be critical for next generation scientific and therapeutic discoveries in the age of precision medicine.

Declaration of Interest

The Authors declare no conflict of interest.

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References

- [1] S. Watson, O. Caster, P.A. Rochon, H. den Ruijter, Reported adverse drug reactions in women and men: Aggregated evidence from globally collected individual case reports during half a century, *EClinicalMedicine*, 17 (2019) 100188.
- [2] J. van Hoof, Female thermal demand, *Nature Climate Change*, 5 (2015) 1029-1030.
- [3] R.N. Weber, Manufacturing Gender in Commercial and Military Cockpit Design, *Science, Technology, & Human Values*, 22 (1997) 235-253.
- [4] A.K. Beery, I. Zucker, Sex bias in neuroscience and biomedical research, *Neurosci Biobehav Rev*, 35 (2011) 565-572.
- [5] N.H.G.R. Institute, *Sex Chromosomes*, 2021.
- [6] F. Mauvais-Jarvis, N. Bairey Merz, P.J. Barnes, R.D. Brinton, J.J. Carrero, D.L. DeMeo, G.J. De Vries, C.N. Epperson, R. Govindan, S.L. Klein, A. Lonardo, P.M. Maki, L.D. McCullough, V. Regitz-Zagrosek, J.G. Regensteiner, J.B. Rubin, K. Sandberg, A. Suzuki, Sex and gender: modifiers of health, disease, and medicine, *Lancet*, 396 (2020) 565-582.
- [7] T. Takahashi, P. Wong, M.K. Ellingson, C. Lucas, J. Klein, B. Israelow, J. Silva, J.E. Oh, T. Mao, M. Tokuyama, P. Lu, A. Venkataraman, A. Park, F. Liu, A. Meir, J. Sun, E.Y. Wang, A.L. Wyllie, C.B.F. Vogels, R. Earnest, S. Lapidus, I.M. Ott, A.J. Moore, A. Casanovas-Massana, C.D. Cruz, J.B. Fournier, C.D. Odio, S. Farhadian, N.D. Grubaugh, W.L. Schulz, A.I. Ko, A.M. Ring, S.B. Omer, A. Iwasaki, I.r.t. Yale, Sex differences in immune responses to SARS-CoV-2 that underlie disease outcomes, *medRxiv*, (2020).
- [8] S. Villa, A. Lombardi, D. Mangioni, G. Bozzi, A. Bandera, A. Gori, M.C. Raviglione, The COVID-19 pandemic preparedness ... or lack thereof: from China to Italy, *Glob Health Med*, 2 (2020) 73-77.
- [9] C. Wenham, J. Smith, R. Morgan, Gender, C.-W. Group, COVID-19: the gendered impacts of the outbreak, *Lancet*, 395 (2020) 846-848.
- [10] S.L. Klein, S. Dhakal, R.L. Ursin, S. Deshpande, K. Sandberg, F. Mauvais-Jarvis, Biological sex impacts COVID-19 outcomes, *PLOS Pathogens*, 16 (2020) e1008570.
- [11] C.H. Sudre, B. Murray, T. Varsavsky, M.S. Graham, R.S. Penfold, R.C. Bowyer, J.C. Pujol, K. Klaser, M. Antonelli, L.S. Canas, E. Molteni, M. Modat, M.J. Cardoso, A. May, S. Ganesh, R. Davies, L.H. Nguyen, D.A. Drew, C.M. Astley, A.D. Joshi, J. Merino, N. Tsereteli, T. Fall, M.F. Gomez, E.L. Duncan, C. Menni, F.M.K. Williams, P.W. Franks, A.T. Chan, J. Wolf, S. Ourselin, T. Spector, C.J. Steves, Attributes and predictors of Long-COVID: analysis

of COVID cases and their symptoms collected by the Covid Symptoms Study App, medRxiv, (2020) 2020.2010.2019.20214494.

[12] A. Parekh, E.O. Fadiran, K. Uhl, D.C. Throckmorton, Adverse effects in women: implications for drug development and regulatory policies, *Expert Rev Clin Pharmacol*, 4 (2011) 453-466.

[13] JCVI, Joint Committee on Vaccination and Immunisation: advice on priority groups for COVID-19 vaccination, UK Government, 2020.

[14] V.W. Pinn, Sex and gender factors in medical studies: implications for health and clinical practice, *JAMA*, 289 (2003) 397-400.

[15] C. Stillhart, K. Vucicevic, P. Augustijns, A.W. Basit, H. Batchelor, T.R. Flanagan, I. Gesquiere, R. Greupink, D. Keszthelyi, M. Koskinen, C.M. Madla, C. Matthys, G. Miljus, M.G. Mooij, N. Parrott, A.L. Ungell, S.N. de Wildt, M. Orlu, S. Klein, A. Mullertz, Impact of gastrointestinal physiology on drug absorption in special populations - An UNGAP review, *Eur J Pharm Sci*, (2020) 105280.

[16] N.C. Woitowich, A. Beery, T. Woodruff, A 10-year follow-up study of sex inclusion in the biological sciences, *Elife*, 9 (2020).

[17] J.A. Clayton, Applying the new SABV (sex as a biological variable) policy to research and clinical care, *Physiol Behav*, 187 (2018) 2-5.

[18] J.P.F. Bai, G.J. Burckart, A.E. Mulberg, Literature Review of Gastrointestinal Physiology in the Elderly, in Pediatric Patients, and in Patients with Gastrointestinal Diseases, *J Pharm Sci*, 105 (2016) 476-483.

[19] FDA, Guidance for Industry: General Considerations for the Clinical Evaluation of Drugs, Food and Drug Administration, 1977.

[20] FDA, Guideline for the Study and Evaluation of Gender Differences in the Clinical Evaluation of Drugs; Notice, Food and Drug Administration, 1993.

[21] P.H. Reports, Women's health. Report of the Public Health Service Task Force on Women's Health Issues, *Public Health Reports* (Washington, D.C. : 1974), 100 (1985) 73-106.

[22] N.I.o. Medicine, Women and Health Research: Ethical and Legal Issues of Including Women in Clinical Studies, Washington (DC), 1993.

[23] FDA, Regulation, Guidance, and Reports related to Women's Health, 2019.

[24] S.V. Rajkumar, Thalidomide: tragic past and promising future, *Mayo Clin Proc*, 79 (2004) 899-903.

[25] W. Lenz, A short history of thalidomide embryopathy, *Teratology*, 38 (1988) 203-215.

- [26] W. Rehman, L.M. Arfons, H.M. Lazarus, The rise, fall and subsequent triumph of thalidomide: lessons learned in drug development, *Ther Adv Hematol*, 2 (2011) 291-308.
- [27] W.G. McBride, Thalidomide and Congenital Abnormalities, *The Lancet*, 1358 (1961) 291-292.
- [28] NIH, NIH Policy and Guidelines on The Inclusion of Women and Minorities as Subjects in Clinical Research, NIH Policy and Compliance, 2001.
- [29] A. Holdcroft, Gender bias in research: how does it affect evidence based medicine?, *J R Soc Med*, 100 (2007) 2-3.
- [30] K.A. Liu, N.A. Mager, Women's involvement in clinical trials: historical perspective and future implications, *Pharm Pract (Granada)*, 14 (2016) 708.
- [31] FDA, FDA Insight: An Insight into Women's Health, U.S. Food and Drug Administration, 2020.
- [32] S.E. Geller, A.R. Koch, P. Roesch, A. Filut, E. Hallgren, M. Carnes, The More Things Change, the More They Stay the Same: A Study to Evaluate Compliance With Inclusion and Assessment of Women and Minorities in Randomized Controlled Trials, *Acad Med*, 93 (2018) 630-635.
- [33] I. Zucker, A.K. Beery, Males still dominate animal studies, *Nature*, 465 (2010) 690.
- [34] E.O. Fadiran, Zhang, L., Effects of sex differences in the pharmacokinetics of drugs and their impact on the safety of medicines in women., in: M. Harrison-Woolrych (Ed.) *Medicines For Women*, Springer International Publishing 2015, pp. 41-68.
- [35] L. Karlsson Lind, M. von Euler, S. Korkmaz, K. Schenck-Gustafsson, Sex differences in drugs: the development of a comprehensive knowledge base to improve gender awareness prescribing, *Biol Sex Differ*, 8 (2017) 32.
- [36] J.A. Fisher, L.M. Ronald, Sex, gender, and pharmaceutical politics: From drug development to marketing, *Gend Med*, 7 (2010) 357-370.
- [37] K.A. Gwee, Irritable bowel syndrome in developing countries--a disorder of civilization or colonization?, *Neurogastroenterol Motil*, 17 (2005) 317-324.
- [38] B.B. Toner, D. Akman, Gender role and irritable bowel syndrome: literature review and hypothesis, *Am J Gastroenterol*, 95 (2000) 11-16.
- [39] P. Layer, J. Keller, H. Loeffler, A. Kreiss, Tegaserod in the treatment of irritable bowel syndrome (IBS) with constipation as the prime symptom, *Ther Clin Risk Manag*, 3 (2007) 107-118.
- [40] FDA, Gastrointestinal Drugs Advisory Subcommittee Meeting, July 14., (2004).

- [41] L.R. Schiller, D.A. Johnson, Balancing drug risk and benefit: toward refining the process of FDA decisions affecting patient care, *Am J Gastroenterol*, 103 (2008) 815-819.
- [42] T. Cvitanović Tomaš, Ž. Urlep, M. Moškon, M. Mraz, D. Rozman, LiverSex Computational Model: Sexual Aspects in Hepatic Metabolism and Abnormalities, *Frontiers in Physiology*, 9 (2018).
- [43] A. Naik, D. Rozman, A. Belič, SteatoNet: The First Integrated Human Metabolic Model with Multi-layered Regulation to Investigate Liver-Associated Pathologies, *PLOS Computational Biology*, 10 (2014) e1003993.
- [44] I. Thiele, S. Sahoo, A. Heinken, L. Heirendt, M.K. Aurich, A. Noronha, R.M.T. Fleming, When metabolism meets physiology: Harvey and Harvetta, *bioRxiv*, (2018) 255885.
- [45] V.M. Miller, Introduction for Sex Differences in Physiology, in: G.N.a.M. Neigh, M.M. (Ed.) *Sex Differences in Physiology*, Academic Press 2016.
- [46] T.M.a.P. Wizemann, M. L., Exploring the Biological Contributions to Human Health: Does Sex Matter?, in: T.M.W.a.M.-L. Pardue (Ed.), *National Academies Press (US)*, Washington (DC), 2001.
- [47] C. Kinsley, Bardi, M., Beigh, G.N., Lambert, K. , Chromosomal and Endocrinological Origins of Sex, in: G.N.a.M. Neigh, M.M. (Ed.) *Sex Differences in Physiology*, Academic Press 2016.
- [48] K. Ethun, Sex and Gender Differences in Body Composition, Lipid Metabolism and Glucose Regulation, in: G.N.a.M. Neigh, M.M. (Ed.) *Sex Differences in Physiology*, Academic Press 2016.
- [49] L. Schiebinger, Klinge, I., *Gendered Innovations 2: How Inclusive Analysis Contributes to Research and Innovation*, 2020.
- [50] O.P. Soldin, S.H. Chung, D.R. Mattison, Sex differences in drug disposition, *J Biomed Biotechnol*, 2011 (2011) 187103.
- [51] A.M. Valodara, K.J. Sr, Sexual Dimorphism in Drug Metabolism and Pharmacokinetics, *Curr Drug Metab*, 20 (2019) 1154-1166.
- [52] G.B. Hatton, V. Yadav, A.W. Basit, H.A. Merchant, Animal Farm: Considerations in Animal Gastrointestinal Physiology and Relevance to Drug Delivery in Humans, *J Pharm Sci*, 104 (2015) 2747-2776.
- [53] F. Gotch, J. Nadell, I.S. Edelman, Gastrointestinal water and electrolytes. IV. The equilibration of deuterium oxide (D₂O) in gastrointestinal contents and the proportion of total body water (T.B.W.) in the gastrointestinal tract, *J Clin Invest*, 36 (1957) 289-296.

- [54] M. Feldman, C. Barnett, Fasting gastric pH and its relationship to true hypochlorhydria in humans, *Dig Dis Sci*, 36 (1991) 866-869.
- [55] M. Feldman, C. Barnett, Fasting gastric pH and its relationship to true hypochlorhydria in humans, *Dig. Dis. Sci.*, 36 (1991) 866-869.
- [56] J.-M. Nicolas, P. Espie, M. Molimard, Gender and interindividual variability in pharmacokinetics, *Drug metabolism reviews*, 41 (2009) 408-421.
- [57] A.C. Freire, A.W. Basit, R. Choudhary, C.W. Piong, H.A. Merchant, Does sex matter? The influence of gender on gastrointestinal physiology and drug delivery, *International Journal of Pharmaceutics*, 415 (2011) 15-28.
- [58] W.R. Hutson, R.L. Roehrkasse, A. Wald, Influence of gender and menopause on gastric emptying and motility, *Gastroenterology*, 96 (1989) 11-17.
- [59] M. Senek, D. Nyholm, E.I. Nielsen, Population pharmacokinetics of levodopa/carbidopa microtablets in healthy subjects and Parkinson's disease patients, *Eur J Clin Pharmacol*, 74 (2018) 1299-1307.
- [60] L. Magallanes, M. Lorier, M. Ibarra, N. Guevara, M. Vazquez, P. Fagiolino, Sex and Food Influence on Intestinal Absorption of Ketoprofen Gastroresistant Formulation, *Clin Pharmacol Drug Dev*, 5 (2016) 196-200.
- [61] G.K. Nandhra, E.B. Mark, G.L. Di Tanna, A.M. Haase, J. Poulsen, S. Christodoulides, V. Kung, M.W. Klinge, K. Knudsen, P. Borghammer, K.O. Andersen, L. Fynne, N. Sutter, V. Schlageter, K. Krogh, A.M. Drewes, M. Birch, S.M. Scott, Normative values for region-specific colonic and gastrointestinal transit times in 111 healthy volunteers using the 3D-Transit electromagnet tracking system: Influence of age, gender, and body mass index, *Neurogastroenterol Motil*, 32 (2020) e13734.
- [62] T. Zimmermann, H. Laufen, R. Yeates, F. Scharpf, K.D. Riedel, T. Schumacher, The pharmacokinetics of extended-release formulations of calcium antagonists and of amlodipine in subjects with different gastrointestinal transit times, *J Clin Pharmacol*, 39 (1999) 1021-1031.
- [63] K.M. Koch, J.L. Palmer, N. Noordin, J.J. Tomlinson, C. Baidoo, Sex and age differences in the pharmacokinetics of alosetron, *Br J Clin Pharmacol*, 53 (2002) 238-242.
- [64] B.E. Viramontes, M. Camilleri, S. McKinzie, D.S. Pardi, D. Burton, G.M. Thomforde, Gender-related differences in slowing colonic transit by a 5-HT₃ antagonist in subjects with diarrhea-predominant irritable bowel syndrome, *Am J Gastroenterol*, 96 (2001) 2671-2676.
- [65] A. Farkouh, T. Riedl, R. Gottardi, M. Czejka, A. Kautzky-Willer, Sex-Related Differences in Pharmacokinetics and Pharmacodynamics of Frequently Prescribed Drugs: A Review of the Literature, *Adv Ther*, 37 (2020) 644-655.

- [66] A. Cabal, Wassenaar, T.M. and Ussery, D.W., Gender Differences in the Gut Microbiome and how these affect Cardiovascular Diseases, in: J.L.a.M. Mehta, J. (Ed.) Gender Differences in the Pathogenesis and Management of Heart Disease, Springer, Switzerland, 2018.
- [67] Z. Vinarov, B. Abrahamsson, P. Artursson, H. Batchelor, P. Berben, A. Bernkop-Schnurch, J. Butler, J. Ceulemans, N. Davies, D. Dupont, G.E. Flaten, N. Fotaki, B.T. Griffin, V. Jannin, J. Keemink, F. Kesisoglou, M. Koziolk, M. Kuentz, A. Mackie, A.J. Melendez-Martinez, M. McAllister, A. Mullertz, C.M. O'Driscoll, N. Parrott, J. Paszkowska, P. Pavek, C.J.H. Porter, C. Reppas, C. Stillhart, K. Sugano, E. Toader, K. Valentova, M. Vertzoni, S.N. De Wildt, C.G. Wilson, P. Augustijns, Current challenges and future perspectives in oral absorption research: An opinion of the UNGAP network, *Adv Drug Deliv Rev*, 171 (2021) 289-331.
- [68] I. Zucker, B.J. Prendergast, Sex differences in pharmacokinetics predict adverse drug reactions in women, *Biol Sex Differ*, 11 (2020) 32.
- [69] D.M. Bush, Emergency Department Visits Attributed to Overmedication that Involved the Insomnia Medication Zolpidem, Substance Abuse and Mental Health Services Administration Rockville (MD), 2014.
- [70] J.L. Norman, D.R. Fixen, J.J. Saseen, L.M. Saba, S.A. Linnebur, Zolpidem prescribing practices before and after Food and Drug Administration required product labeling changes, *SAGE Open Med*, 5 (2017) 2050312117707687.
- [71] GAO, Drug Safety: Most Drugs Withdrawn in Recent Years Had Greater Health Risk for Women, United States General Accounting Office, Washington, 2001.
- [72] J.C. Kando, K.A. Yonkers, J.O. Cole, Gender as a risk factor for adverse events to medications, *Drugs*, 50 (1995) 1-6.
- [73] G.D. Anderson, Sex and racial differences in pharmacological response: where is the evidence? Pharmacogenetics, pharmacokinetics, and pharmacodynamics, *J Womens Health (Larchmt)*, 14 (2005) 19-29.
- [74] F. Mauvais-Jarvis, H.K. Berthold, I. Campesi, J.J. Carrero, S. Dakal, F. Franconi, I. Gouni-Berthold, M.L. Heiman, A. Kautzky-Willer, S.L. Klein, A. Murphy, V. Regitz-Zagrosek, K. Reue, J.B. Rubin, Sex- and Gender-Based Pharmacological Response to Drugs, *Pharmacol Rev*, 73 (2021) 730-762.
- [75] M. Vertzoni, P. Augustijns, M. Grimm, M. Koziolk, G. Lemmens, N. Parrott, C. Pentafragka, C. Reppas, J. Rubbens, J. Van Den Abeele, T. Vanuytsel, W. Weitschies, C.G. Wilson, Impact of regional differences along the gastrointestinal tract of healthy adults on oral

drug absorption: An UNGAP review, *European Journal of Pharmaceutical Sciences*, 134 (2019) 153-175.

[76] L. Shiebinger, Klinge, I., Paik, H. Y., Sánchez de Madariaga, I., Schraudner, M., Stefanick, M., *Gendered Innovations in Science, Health & Medicine, Engineering, and Environment*, 2011-2020.

[77] L. Aarons, K. Hopkins, M. Rowland, S. Brossel, J.F. Thiercelin, Route of Administration and Sex-Differences in the Pharmacokinetics of Aspirin, Administered as Its Lysine Salt, *Pharm Res-Dordr*, 6 (1989) 660-666.

[78] C.M. Metzler, G.L. Elfring, A. McEwen, A users manual for NONLIN and associated programs, Fotostelle der UB rev1976.

[79] P.C. Ho, E.J. Triggs, D.W. Bourne, V.J. Heazlewood, The effects of age and sex on the disposition of acetylsalicylic acid and its metabolites, *Br J Clin Pharmacol*, 19 (1985) 675-684.

[80] D. Mattison, A. Zajicek, Gaps in knowledge in treating pregnant women, *Gend Med*, 3 (2006) 169-182.

[81] F. Mesnil, F. Mentre, C. Dubruc, J.P. Thenot, A. Mallet, Population pharmacokinetic analysis of mizolastine and validation from sparse data on patients using the nonparametric maximum likelihood method, *J Pharmacokinet Biopharm*, 26 (1998) 133-161.

[82] P.E. Johnson, D.B. Milne, G.I. Lykken, Effects of age and sex on copper absorption, biological half-life, and status in humans, *The American journal of clinical nutrition*, 56 (1992) 917-925.

[83] M.E. McOmber, C.-N. Ou, R.J. Shulman, Effects of timing, sex, and age on site-specific gastrointestinal permeability testing in children and adults, *J Pediatr Gastroenterol Nutr*, 50 (2010) 269-275.

[84] M. Karlgren, A. Vildhede, U. Norinder, J.R. Wisniewski, E. Kimoto, Y. Lai, U. Haglund, P. Artursson, Classification of inhibitors of hepatic organic anion transporting polypeptides (OATPs): influence of protein expression on drug-drug interactions, *J Med Chem*, 55 (2012) 4740-4763.

[85] Q. Zhou, Q.X. Chen, Z.R. Ruan, H. Yuan, H.M. Xu, S. Zeng, CYP2C9*3(1075A > C), ABCB1 and SLCO1B1 genetic polymorphisms and gender are determinants of inter-subject variability in pitavastatin pharmacokinetics, *Pharmazie*, 68 (2013) 187-194.

[86] J.A. Hubacek, D. Dlouha, V. Adamkova, V. Lanska, R. Ceska, M. Vrablik, Possible gene-gender interaction between the SLCO1B1 polymorphism and statin treatment efficacy, *Neuro Endocrinol Lett*, 33 Suppl 2 (2012) 22-25.

- [87] E. Link, S. Parish, J. Armitage, L. Bowman, S. Heath, I. Matsuda, The Study of the Effectiveness of Additional Reductions in Cholesterol and Homocysteine (SEARCH) Collaborative Group. SLCO1B1 variants and statin-induced myopathy—a genomewide study, *N Engl J Med*, 359 (2008) 789-799.
- [88] O.V. Smirnova, [Sex differences in drug action: the role of multidrug-resistance proteins (MRPs)], *Fiziol Cheloveka*, 38 (2012) 124-136.
- [89] Y. Mai, L. Dou, Z. Yao, C.M. Madla, F.K.H. Gavins, F. Taherali, H. Yin, M. Orlu, S. Murdan, A.W. Basit, Quantification of P-Glycoprotein in the Gastrointestinal Tract of Humans and Rodents: Methodology, Gut Region, Sex, and Species Matter, *Mol Pharm*, (2021).
- [90] B. Meibohm, I. Beierle, H. Derendorf, How important are gender differences in pharmacokinetics?, *Clinical pharmacokinetics*, 41 (2002) 329-342.
- [91] M. Frohlich, N. Albermann, A. Sauer, I. Walter-Sack, W.E. Haefeli, J. Weiss, In vitro and ex vivo evidence for modulation of P-glycoprotein activity by progestins, *Biochem Pharmacol*, 68 (2004) 2409-2416.
- [92] A. Nakayama, O. Eguchi, M. Hatakeyama, H. SAITOH, M. TAKADA, Defferent Absorption Behaviors among Steroid Hormones Due to Possible Interaction with P-Glycoprotein in the Rat Small Intestine, *Biological and Pharmaceutical Bulletin*, 22 (1999) 535-538.
- [93] S.L. Klein, C.W. Roberts, Sex and gender differences in infection and treatments for infectious diseases, Springer 2015.
- [94] C.D.a.A. Klaassen, L.M., Xenobiotic, Bile Acid, and Cholesterol Transporters: Function and Regulation, *Pharmacol Rev*, 62 (2010) 1-96.
- [95] H. Lu, C. Chen, C. Klaassen, Tissue distribution of concentrative and equilibrative nucleoside transporters in male and female rats and mice, *Drug Metab Dispos*, 32 (2004) 1455-1461.
- [96] J.M. Maher, A.L. Slitt, N.J. Cherrington, X. Cheng, C.D. Klaassen, Tissue distribution and hepatic and renal ontogeny of the multidrug resistance-associated protein (Mrp) family in mice, *Drug Metab Dispos*, 33 (2005) 947-955.
- [97] Y.J. Cui, X. Cheng, Y.M. Weaver, C.D. Klaassen, Tissue distribution, gender-divergent expression, ontogeny, and chemical induction of multidrug resistance transporter genes (Mdr1a, Mdr1b, Mdr2) in mice, *Drug Metab Dispos*, 37 (2009) 203-210.
- [98] Y. Tanaka, A.L. Slitt, T.M. Leazer, J.M. Maher, C.D. Klaassen, Tissue distribution and hormonal regulation of the breast cancer resistance protein (Bcrp/Abcg2) in rats and mice, *Biochem Biophys Res Commun*, 326 (2005) 181-187.

- [99] A.J. Lickteig, X. Cheng, L.M. Augustine, C.D. Klaassen, N.J. Cherrington, Tissue distribution, ontogeny and induction of the transporters Multidrug and toxin extrusion (MATE) 1 and MATE2 mRNA expression levels in mice, *Life Sci*, 83 (2008) 59-64.
- [100] D. Rost, K. Kopplow, S. Gehrke, S. Mueller, H. Friess, C. Ittrich, D. Mayer, A. Stiehl, Gender-specific expression of liver organic anion transporters in rat, *Eur J Clin Invest*, 35 (2005) 635-643.
- [101] G. Merino, A.E. van Herwaarden, E. Wagenaar, J.W. Jonker, A.H. Schinkel, Sex-dependent expression and activity of the ATP-binding cassette transporter breast cancer resistance protein (BCRP/ABCG2) in liver, *Mol Pharmacol*, 67 (2005) 1765-1771.
- [102] X. Cheng, C.D. Klaassen, Tissue distribution, ontogeny, and hormonal regulation of xenobiotic transporters in mouse kidneys, *Drug Metab Dispos*, 37 (2009) 2178-2185.
- [103] I.T. Houghton, C.S.T. Aun, T.E. Oh, Vecuronium - an Anthropometric Comparison, *Anaesthesia*, 47 (1992) 741-746.
- [104] F.S. Xue, S.Y. Tong, X. Liao, J.H. Liu, G. An, L.K. Luo, Dose-response and time course of effect of rocuronium in male and female anesthetized patients, *Anesth Analg*, 85 (1997) 667-671.
- [105] H.R. Ochs, D.J. Greenblatt, M. Divoll, D.R. Abernethy, H. Feyerabend, H.J. Dengler, Diazepam Kinetics in Relation to Age and Sex, *Pharmacology*, 23 (1981) 24-30.
- [106] V. Regitz-Zagrosek, U. Seeland, Sex and gender differences in clinical medicine, *Handb Exp Pharmacol*, (2012) 3-22.
- [107] J.B. Schwartz, The current state of knowledge on age, sex, and their interactions on clinical pharmacology, *Clin Pharmacol Ther*, 82 (2007) 87-96.
- [108] J.B. Schwartz, The influence of sex on pharmacokinetics, *Clin Pharmacokinet*, 42 (2003) 107-121.
- [109] M. Frezza, C. di Padova, G. Pozzato, M. Terpin, E. Baraona, C.S. Lieber, High blood alcohol levels in women: the role of decreased gastric alcohol dehydrogenase activity and first-pass metabolism, *New England Journal of Medicine*, 322 (1990) 95-99.
- [110] A. Parlesak, M.H.-U. Billinger, C. Bode, J.C. Bode, Gastric alcohol dehydrogenase activity in man: influence of gender, age, alcohol consumption and smoking in a Caucasian population, *Alcohol and Alcoholism*, 37 (2002) 388-393.
- [111] D.J. Waxman, M.G. Holloway, Sex differences in the expression of hepatic drug metabolizing enzymes, *Mol Pharmacol*, 76 (2009) 215-228.
- [112] C.M. Hunt, W.R. Westerham, G.M. Stave, Effect of age and gender on the activity of human hepatic CYP3A, *Biochem Pharmacol*, 44 (1992) 275-283.

- [113] R. Schmidt, F. Baumann, H. Hanschmann, F. Geissler, R. Preiss, Gender difference in ifosfamide metabolism by human liver microsomes, *Eur J Drug Metab Pharmacokinet*, 26 (2001) 193-200.
- [114] E. Tanaka, Gender-related differences in pharmacokinetics and their clinical significance, *J Clin Pharm Ther*, 24 (1999) 339-346.
- [115] D.J. Greenblatt, L.L. von Moltke, Gender has a small but statistically significant effect on clearance of CYP3A substrate drugs, *J Clin Pharmacol*, 48 (2008) 1350-1355.
- [116] O.P. Soldin, D.R. Mattison, Sex differences in pharmacokinetics and pharmacodynamics, *Clin Pharmacokinet*, 48 (2009) 143-157.
- [117] J.M. Maglich, C.M. Stoltz, B. Goodwin, D. Hawkins-Brown, J.T. Moore, S.A. Kliewer, Nuclear pregnane x receptor and constitutive androstane receptor regulate overlapping but distinct sets of genes involved in xenobiotic detoxification, *Molecular pharmacology*, 62 (2002) 638-646.
- [118] Y. Tsuchiya, M. Nakajima, S. Kyo, T. Kanaya, M. Inoue, T. Yokoi, Human CYP1B1 is regulated by estradiol via estrogen receptor, *Cancer Res*, 64 (2004) 3119-3125.
- [119] E.J. Seaber, R.W. Peck, D.A. Smith, J. Allanson, N.R. Hefting, J.J. van Lier, F.A. Sollie, J. Wemer, J.H. Jonkman, The absolute bioavailability and effect of food on the pharmacokinetics of zolmitriptan in healthy volunteers, *Br J Clin Pharmacol*, 46 (1998) 433-439.
- [120] E. Seaber, N. On, R.M. Dixon, M. Gibbens, W.J. Leavens, J. Liptrot, G. Chittick, J. Posner, P.E. Rolan, R.W. Peck, The absolute bioavailability and metabolic disposition of the novel antimigraine compound zolmitriptan (311C90), *Brit J Clin Pharmacol*, 43 (1997) 579-587.
- [121] E. Seaber, N. On, S. Phillips, R. Churchus, J. Posner, P. Rolan, The tolerability and pharmacokinetics of the novel antimigraine compound 311C90 in healthy male volunteers, *Brit J Clin Pharmacol*, 41 (1996) 141-147.
- [122] S.E. Gaudry, D.S. Sitar, D.D. Smyth, J.K. McKenzie, F.Y. Aoki, Gender and age as factors in the inhibition of renal clearance of amantadine by quinine and quinidine, *Clin Pharmacol Ther*, 54 (1993) 23-27.
- [123] U.B. Berg, Differences in decline in GFR with age between males and females. Reference data on clearances of inulin and PAH in potential kidney donors, *Nephrol Dial Transpl*, 21 (2006) 2577-2582.

- [124] M. Pirmohamed, S. James, S. Meakin, C. Green, A.K. Scott, T.J. Walley, K. Farrar, B.K. Park, A.M. Breckenridge, Adverse drug reactions as cause of admission to hospital: prospective analysis of 18 820 patients, *BMJ*, 329 (2004) 15-19.
- [125] S. Joseph, T.J. Nicolson, G. Hammons, B. Word, B. Green-Knox, B. Lyn-Cook, Expression of drug transporters in human kidney: impact of sex, age, and ethnicity, *Biol Sex Differ*, 6 (2015) 4.
- [126] S.L. Seliger, C. Davis, C. Stehman-Breen, Gender and the progression of renal disease, *Curr Opin Nephrol Hypertens*, 10 (2001) 219-225.
- [127] L. Butera, D.A. Feinfeld, M. Bhargava, Sex differences in the subunits of glutathione-S-transferase isoenzyme from rat and human kidney, *Enzyme*, 43 (1990) 175-182.
- [128] G. Di Pietro, L.A.V. Magno, F. Rios-Santos, Glutathione S-transferases: an overview in cancer research, *Expert opinion on drug metabolism & toxicology*, 6 (2010) 153-170.
- [129] H. Hoensch, I. Morgenstern, G. Petereit, M. Siepmann, W. Peters, H. Roelofs, W. Kirch, Influence of clinical factors, diet, and drugs on the human upper gastrointestinal glutathione system, *Gut*, 50 (2002) 235-240.
- [130] L. Butera, D.A. Feinfeld, M. Bhargava, Sex differences in the subunits of glutathione-S-transferase isoenzyme from rat and human kidney, *Enzyme*, 43 (1990) 175-182.
- [131] J.T. Slattery, J.M. Wilson, T.F. Kalthorn, S.D. Nelson, Dose-Dependent Pharmacokinetics of Acetaminophen - Evidence of Glutathione Depletion in Humans, *Clinical Pharmacology & Therapeutics*, 41 (1987) 413-418.
- [132] O.P. Soldin, S.H. Chung, D.R. Mattison, Sex Differences in Drug Disposition, *Journal of Biomedicine and Biotechnology*, 2011 (2011) 187103.
- [133] B.R. Migeon, X inactivation, female mosaicism, and sex differences in renal diseases, *J Am Soc Nephrol*, 19 (2008) 2052-2059.
- [134] K.V. Juul, B.M. Klein, R. Sandstrom, L. Erichsen, J.P. Norgaard, Gender difference in antidiuretic response to desmopressin, *Am J Physiol Renal Physiol*, 300 (2011) F1116-1122.
- [135] M.K. Schroeder, K.V. Juul, B. Mahler, J.P. Norgaard, S. Rittig, Desmopressin use in pediatric nocturnal enuresis patients: is there a sex difference in prescription patterns?, *Eur J Pediatr*, 177 (2018) 389-394.
- [136] D.A. Ashiru, R. Patel, A.W. Basit, Polyethylene glycol 400 enhances the bioavailability of a BCS class III drug (ranitidine) in male subjects but not females, *Pharm Res*, 25 (2008) 2327-2333.

- [137] F.S. Xue, S.Y. Tong, X. Liao, J.H. Liu, G. An, L.K. Luo, Dose-response and time course of effect of rocuronium in male and female anesthetized patients, *Anesth Analg*, 85 (1997) 667-671.
- [138] E. Seaber, N. On, R.M. Dixon, M. Gibbens, W.J. Leavens, J. Liptrot, G. Chittick, J. Posner, P.E. Rolan, R.W. Pack, The absolute bioavailability and metabolic disposition of the novel antimigraine compound zolmitriptan (311C90), *Br J Clin Pharmacol*, 43 (1997) 579-587.
- [139] D.J. Greenblatt, R.I. Shader, K. Franke, D.S. Maclaughlin, B.J. Ransil, J. Kochwaser, Kinetics of Intravenous Chlordiazepoxide - Sex-Differences in Drug Distribution, *Clin Pharmacol Ther*, 22 (1977) 893-903.
- [140] D.R. Terrell, S.K. Vesely, J.A. Kremer Hovinga, B. Lammle, J.N. George, Different disparities of gender and race among the thrombotic thrombocytopenic purpura and hemolytic-uremic syndromes, *Am J Hematol*, 85 (2010) 844-847.
- [141] A. Cassidy, J.E. Brown, A. Hawdon, M.S. Faughnan, L.J. King, J. Millward, L. Zimmer-Nechemias, B. Wolfe, K.D. Setchell, Factors affecting the bioavailability of soy isoflavones in humans after ingestion of physiologically relevant levels from different soy foods, *J Nutr*, 136 (2006) 45-51.
- [142] C. Carrasco-Portugal Mdel, F.J. Flores-Murrieta, Gender differences in the oral pharmacokinetics of fluconazole, *Clin Drug Investig*, 27 (2007) 851-855.
- [143] S.F. Cooper, D. Drolet, R. Dugal, Comparative Bioavailability of 2 Oral Formulations of Flurazepam in Human-Subjects, *Biopharm Drug Dispos*, 5 (1984) 127-139.
- [144] C. Anderwald, A. Gastaldelli, A. Tura, M. Krebs, M. Promintzer-Schifferl, A. Kautzky-Willer, M. Stadler, R.A. DeFronzo, G. Pacini, M.G. Bischof, Mechanism and Effects of Glucose Absorption during an Oral Glucose Tolerance Test Among Females and Males, *J Clin Endocr Metab*, 96 (2011) 515-524.
- [145] K. Faerch, G. Pacini, J.J. Nolan, T. Hansen, A. Tura, D. Vistisen, Impact of Glucose Tolerance Status, Sex, and Body Size on Glucose Absorption Patterns During OGTTs, *Diabetes Care*, 36 (2013) 3691-3697.
- [146] N.R.C. Campbell, R.D. Hull, R. Brant, D.B. Hogan, G.F. Pineo, G.E. Raskob, Different effects of heparin in males and females, *Clin Invest Med*, 21 (1998) 71-78.
- [147] S. Almeida, A. Filipe, A. Almeida, H. Wong, N. Caparros, M. Tanguay, Comparative bioavailability of two formulations of levofloxacin and effect of sex on bioequivalence analysis. Data from a randomised, 2 x 2 crossover trial in healthy volunteers, *Arzneimittelforschung*, 55 (2005) 414-419.

- [148] P.A. Routledge, W.W. Stargel, B.B. Kitchell, A. Barchowsky, D.G. Shand, Sex-related differences in the plasma protein binding of lignocaine and diazepam, *Br J Clin Pharmacol*, 11 (1981) 245-250.
- [149] G.K. Ciccone, A. Holdcroft, Drugs and sex differences: a review of drugs relating to anaesthesia, *Br J Anaesth*, 82 (1999) 255-265.
- [150] T. Cabaleiro, M. Roman, D. Ochoa, M. Talegon, R. Prieto-Perez, A. Wojnicz, R. Lopez-Rodriguez, J. Novalbos, F. Abad-Santos, Evaluation of the relationship between sex, polymorphisms in CYP2C8 and CYP2C9, and pharmacokinetics of angiotensin receptor blockers, *Drug Metab Dispos*, 41 (2013) 224-229.
- [151] K.H. Lew, E.A. Ludwig, M.A. Milad, K. Donovan, E. Middleton, Jr., J.J. Ferry, W.J. Jusko, Gender-based effects on methylprednisolone pharmacokinetics and pharmacodynamics, *Clin Pharmacol Ther*, 54 (1993) 402-414.
- [152] A.B. Luzier, A. Killian, J.H. Wilton, M.F. Wilson, A. Forrest, D.J. Kazierad, Gender-related effects on metoprolol pharmacokinetics and pharmacodynamics in healthy volunteers, *Clin Pharmacol Ther*, 66 (1999) 594-601.
- [153] A.J. Carcas, P. Guerra, J. Frias, A. Soto, A. Fernandez-Aijon, C. Montuenga, C. Govantes, Gender differences in the disposition of metronidazole, *Int J Clin Pharm Th*, 39 (2001) 213-218.
- [154] G.C. Sun, M.C. Hsu, Y.Y. Chia, P.Y. Chen, F.Z. Shaw, Effects of age and gender on intravenous midazolam premedication: a randomized double-blind study, *Br J Anaesth*, 101 (2008) 632-639.
- [155] F.O. Hassan, Hand dominance and gender in forearm fractures in children, *Strategies Trauma Limb Reconstr*, 3 (2008) 101-103.
- [156] U. Werner, D. Werner, S. Heinbuchner, B. Graf, H. Ince, S. Kische, P. Thurmann, J. Konig, M.F. Fromm, O. Zolk, Gender Is an Important Determinant of the Disposition of the Loop Diuretic Torasemide, *J Clin Pharmacol*, 50 (2010) 160-168.
- [157] D. Kang, D. Verotta, M.E. Krecic-Shepard, N.B. Modi, S.K. Gupta, J.B. Schwartz, Population analyses of sustained-release verapamil in patients: effects of sex, race, and smoking, *Clin Pharmacol Ther*, 73 (2003) 31-40.
- [158] M.E. Krecic-Shepard, C.R. Barnas, J. Slimko, M.P. Jones, J.B. Schwartz, Gender-specific effects on verapamil pharmacokinetics and pharmacodynamics in humans, *J Clin Pharmacol*, 40 (2000) 219-230.

- [159] W.B. White, M.F. Johnson, H.R. Black, W.J. Elliott, D.A. Sica, Gender and age effects on the ambulatory blood pressure and heart rate responses to antihypertensive therapy, *Am J Hypertens*, 14 (2001) 1239-1247.
- [160] J. Zhang, C.Y. Li, M.J. Xu, T. Wu, J.H. Chu, S.J. Liu, W.Z. Ju, Oral bioavailability and gender-related pharmacokinetics of celastrol following administration of pure celastrol and its related tablets in rats, *J Ethnopharmacol*, 144 (2012) 195-200.
- [161] H.L. Lujan, S.E. Dicarolo, Sex differences to myocardial ischemia and beta-adrenergic receptor blockade in conscious rats, *Am J Physiol Heart Circ Physiol*, 294 (2008) H1523-1529.
- [162] J.K. Los, D.A. Welsh, E.G. Herold, W.J. Bagdon, A. Zacchei, Gender differences in toxicokinetics, liver metabolism, and plasma esterase activity: observations from a chronic (27-week) toxicity study of enalapril/diltiazem combinations in rats., *Drug Metab. Dispos.*, 24 (1996) 28-33.
- [163] X.D. Liu, L. Xie, Y. Zhong, C.X. Li, Gender difference in letrozole pharmacokinetics in rats, *Acta Pharmacol Sin*, 21 (2000) 680-684.
- [164] X.D. Liu, X.L. Wang, L. Xie, G.L. Wang, Different effect of erythromycin on absorption kinetics of nimodipine in male and female rats, *Eur J Drug Metab Ph*, 30 (2005) 69-73.
- [165] X.D. Liu, L. Xie, Y. Liang, L. Li, T. Lu, Gender difference in ranolazine pharmacokinetics in rats, *Eur J Drug Metab Ph*, 28 (2003) 119-123.
- [166] M.J. Xu, G.J. Wang, H.T. Xie, H. Li, Q. Huang, R. Wang, Y.W. Jia, T. Lv, Gender difference regarding schizandrin pharmacokinetics in rats, *Eur J Drug Metab Ph*, 33 (2008) 65-68.
- [167] J.H. Schlattjan, F. Biggemann, J. Greven, Gender differences in renal tubular taurocholate transport, *N-S Arch Pharmacol*, 371 (2005) 449-456.
- [168] H.P. Whitley, W. Lindsey, Sex-based differences in drug activity, *American family physician*, 80 (2009) 1254-1258.
- [169] H. Patel, D. Bell, M. Molokhia, J. Srishanmuganathan, M. Patel, J. Car, A. Majeed, Trends in hospital admissions for adverse drug reactions in England: analysis of national hospital episode statistics 1998–2005, *BMC clinical pharmacology*, 7 (2007) 9.
- [170] M. Pirmohamed, S. James, S. Meakin, C. Green, A.K. Scott, T.J. Walley, K. Farrar, B.K. Park, A.M. Breckenridge, Adverse drug reactions as cause of admission to hospital: prospective analysis of 18 820 patients, *Bmj*, 329 (2004) 15-19.
- [171] O.P. Soldin, D.R. Mattison, Sex Differences in Pharmacokinetics and Pharmacodynamics, *Clinical Pharmacokinetics*, 48 (2009) 143-157.

- [172] L. Thunander Sundbom, K. Binge-fors, Women and men report different behaviours in, and reasons for medication non-adherence: a nationwide Swedish survey, *Pharm Pract (Granada)*, 10 (2012) 207-221.
- [173] F. Aubrun, N. Salvi, P. Coriat, B. Riou, Sex- and Age-related Differences in Morphine Requirements for Postoperative Pain Relief, *Anesthesiology*, 103 (2005) 156-160.
- [174] R.M. Craft, Sex differences in drug-and non-drug-induced analgesia, *Life sciences*, 72 (2003) 2675-2688.
- [175] S. Pieretti, A. Di Giannuario, R. Di Giovannandrea, F. Marzoli, G. Piccaro, P. Minosi, A.M. Aloisi, Gender differences in pain and its relief, *Ann Ist Super Sanita*, 52 (2016) 184-189.
- [176] M.S. Cepeda, J.T. Farrar, M. Baumgarten, R. Boston, D.B. Carr, B.L. Strom, Side effects of opioids during short-term administration: effect of age, gender, and race, *Clin Pharmacol Ther*, 74 (2003) 102-112.
- [177] Kando., Gender as a risk factor for adverse events to medications, *Drugs*, 50 (1995) 1-6.
- [178] I. Beierle, B. Meibohm, H. Derendorf, Gender differences in pharmacokinetics and pharmacodynamics, *Int J Clin Pharmacol Ther*, 37 (1999) 529-547.
- [179] G. Labots, A. Jones, S.J. de Visser, R. Rissmann, J. Burggraaf, Gender differences in clinical registration trials: is there a real problem?, *Br J Clin Pharmacol*, 84 (2018) 700-707.
- [180] V. Jarugula, C.-M. Yeh, D. Howard, C. Bush, D.L. Keefe, W.P. Dole, Influence of Body Weight and Gender on the Pharmacokinetics, Pharmacodynamics, and Antihypertensive Efficacy of Aliskiren, *The Journal of Clinical Pharmacology*, 50 (2010) 1358-1366.
- [181] D. Kang, D. Verotta, J.B. Schwartz, Population analyses of amlodipine in patients living in the community and patients living in nursing homes, *Clin Pharmacol Ther*, 79 (2006) 114-124.
- [182] FDA, NORVASC® (amlodipine besylate), 2011.
- [183] A.R. Hobson, Z. Qureshi, P. Banks, N. Curzen, Gender and responses to aspirin and clopidogrel: insights using short thrombelastography, *Cardiovasc Ther*, 27 (2009) 246-252.
- [184] N.R. Jørgensen, E.L. Grove, P. Schwarz, P. Vestergaard, Clopidogrel and the risk of osteoporotic fractures: a nationwide cohort study, *J Intern Med*, 272 (2012) 385-393.
- [185] T. Ciarambino, G. Corbi, A. Filippelli, M. La Regina, O. Para, F. Tangianu, P. Gnerre, N. Ferrara, M. Giordano, C. Politi, Anticoagulant drugs and gender: what is in the elderly? A minireview, *JOURNAL OF GERONTOLOGY AND GERIATRICS*, 67 (2019) 123-126.

- [186] K.W. McConeghy, A. Bress, D.M. Qato, C. Wing, E.A. Nutescu, Evaluation of dabigatran bleeding adverse reaction reports in the FDA adverse event reporting system during the first year of approval, *Pharmacotherapy*, 34 (2014) 561-569.
- [187] S.S. Rathore, Y. Wang, H.M. Krumholz, Sex-Based Differences in the Effect of Digoxin for the Treatment of Heart Failure, *New England Journal of Medicine*, 347 (2002) 1403-1411.
- [188] K.H. Humphries, M. Izadnegahdar, T. Sedlak, J. Saw, N. Johnston, K. Schenck-Gustafsson, R.U. Shah, V. Regitz-Zagrosek, J. Grewal, V. Vaccarino, J. Wei, C.N. Bairey Merz, Sex differences in cardiovascular disease - Impact on care and outcomes, *Front Neuroendocrinol*, 46 (2017) 46-70.
- [189] K.F. Adams, J.H. Patterson, W.A. Gattis, C.M. O'Connor, C.R. Lee, T.A. Schwartz, M. Gheorghiade, Relationship of Serum Digoxin Concentration to Mortality and Morbidity in Women in the Digitalis Investigation Group Trial: A Retrospective Analysis, *Journal of the American College of Cardiology*, 46 (2005) 497-504.
- [190] H. Roukoz, W. Saliba, Dofetilide: a new class III antiarrhythmic agent, *Expert Rev Cardiovasc Ther*, 5 (2007) 9-19.
- [191] T. Ishikawa, K. Mizuno, N. Nakaya, Y. Ohashi, N. Tajima, T. Kushiro, T. Teramoto, S. Uchiyama, H. Nakamura, The relationship between the effect of pravastatin and risk factors for coronary heart disease in Japanese patients with hypercholesterolemia, *Circ J*, 72 (2008) 1576-1582.
- [192] MRC_Report., Adverse reactions to bendrofluazide and propranolol for the treatment of mild hypertension. Report of Medical Research Council Working Party on Mild to Moderate Hypertension, *Lancet*, 2 (1981) 539-543.
- [193] D. Werner, U. Werner, A. Meybaum, B. Schmidt, S. Umbreen, A. Grosch, H.G. Lestin, B. Graf, O. Zolk, M.F. Fromm, Determinants of steady-state torasemide pharmacokinetics: impact of pharmacogenetic factors, gender and angiotensin II receptor blockers, *Clin Pharmacokinet*, 47 (2008) 323-332.
- [194] G.S. Alotaibi, H. Almodaimegh, M.S. McMurtry, C. Wu, Do women bleed more than men when prescribed novel oral anticoagulants for venous thromboembolism? A sex-based meta-analysis, *Thromb Res*, 132 (2013) 185-189.
- [195] K.H. Humphries, C.R. Kerr, S.J. Connolly, G. Klein, J.A. Boone, M. Green, R. Sheldon, M. Talajic, P. Dorian, D. Newman, New-onset atrial fibrillation: sex differences in presentation, treatment, and outcome, *Circulation*, 103 (2001) 2365-2370.
- [196] C. Belmonte, D. Ochoa, M. Román, T. Cabaleiro, M. Talegón, S.D. Sánchez-Rojas, F. Abad-Santos, Evaluation of the Relationship Between Pharmacokinetics and the Safety of

Aripiprazole and Its Cardiovascular Effects in Healthy Volunteers, *J Clin Psychopharmacol*, 36 (2016) 608-614.

[197] S.L. Garnaat, R.B. Weisberg, L.A. Uebelacker, D.S. Herman, G.L. Bailey, B.J. Anderson, K.M. Sharkey, M.D. Stein, The overlap of sleep disturbance and depression in primary care patients treated with buprenorphine, *Substance abuse*, 38 (2017) 450-454.

[198] M. Macaluso, R. Zackula, I. D'Empaire, B. Baker, K. Liow, S.H. Preskorn, Twenty percent of a representative sample of patients taking bupropion have abnormal, asymptomatic electroencephalographic findings, *J Clin Psychopharmacol*, 30 (2010) 312-317.

[199] J. Davidson, Seizures and bupropion: a review, *J Clin Psychiatry*, 50 (1989) 256-261.

[200] G.J. Macphee, J.R. Mitchell, L. Wiseman, A.R. McLellan, B.K. Park, G.T. McInnes, M.J. Brodie, Effect of sodium valproate on carbamazepine disposition and psychomotor profile in man, *British journal of clinical pharmacology*, 25 (1988) 59-66.

[201] T. Sudhop, J. Bauer, C.E. Elger, K. von Bergmann, Increased high-density lipoprotein cholesterol in patients with epilepsy treated with carbamazepine: a gender-related study, *Epilepsia*, 40 (1999) 480-484.

[202] T.S. Barclay, A.J. Lee, Citalopram-associated SIADH, *Annals of Pharmacotherapy*, 36 (2002) 1558-1563.

[203] S.L. Lau, C. Muir, Y. Assur, R. Beach, B. Tran, R. Bartrop, M. McLean, D. Caetano, Predicting Weight Gain in Patients Treated With Clozapine: The Role of Sex, Body Mass Index, and Smoking, *J Clin Psychopharmacol*, 36 (2016) 120-124.

[204] J. Nielsen, J.M. Meyer, Risk factors for ileus in patients with schizophrenia, *Schizophr Bull*, 38 (2012) 592-598.

[205] S. West, D. Rowbotham, G. Xiong, C. Kenedi, Clozapine induced gastrointestinal hypomotility: A potentially life threatening adverse event. A review of the literature, *Gen Hosp Psychiatry*, 46 (2017) 32-37.

[206] A. Ventriglio, R.J. Baldessarini, G. Vitrani, I. Bonfitto, A.C. Cecere, A. Rinaldi, A. Petito, A. Bellomo, Metabolic Syndrome in Psychotic Disorder Patients Treated With Oral and Long-Acting Injected Antipsychotics, *Front Psychiatry*, 9 (2018) 744.

[207] M. Ahmed, I. Hussain, S.M. O'Brien, B. Dineen, D. Griffin, C. McDonald, Prevalence and associations of the metabolic syndrome among patients prescribed clozapine, *Irish Journal of Medical Science*, 177 (2008) 205-210.

[208] N.H. Covell, E.M. Weissman, S.M. Essock, Weight gain with clozapine compared to first generation antipsychotic medications, *Schizophr Bull*, 30 (2004) 229-240.

- [209] L. Bailey, S. Varma, N. Ahmad, S. Gee, D.M. Taylor, Factors predicting use of laxatives in outpatients stabilized on clozapine, *Therapeutic Advances in Psychopharmacology*, 5 (2015) 256-262.
- [210] S.G. Anderson, M. Livingston, L. Couchman, D.J. Smith, M. Connolly, J. Miller, R.J. Flanagan, A.H. Heald, Sex differences in plasma clozapine and norclozapine concentrations in clinical practice and in relation to body mass index and plasma glucose concentrations: a retrospective survey, *Ann Gen Psychiatry*, 14 (2015) 39.
- [211] N. Tunsirimas, P. Pariwatcharakul, S. Choovanichvong, W. Ratta-apha, Clozapine-induced agranulocytosis and leukopenia: Incidence, associated factors, and rate of hematologic adverse-effects monitoring in psychiatric out-patient services in Thailand, *Asian Journal of Psychiatry*, 41 (2019) 13-16.
- [212] S.A. Hollingworth, K. Winckel, N. Saiepour, A.J. Wheeler, N. Myles, D. Siskind, Clozapine-related neutropenia, myocarditis and cardiomyopathy adverse event reports in Australia 1993–2014, *Psychopharmacology*, 235 (2018) 1915-1921.
- [213] E.S. Palva, Gender-related differences in diazepam effects on performance, *Med Biol*, 63 (1985) 92-95.
- [214] R.L. Doty, J. Treem, I. Tourbier, N. Mirza, A double-blind study of the influences of eszopiclone on dysgeusia and taste function, *Pharmacol Biochem Behav*, 94 (2009) 312-318.
- [215] L. Manthey, C. Leeds, E.J. Giltay, T. van Veen, S.A. Vreeburg, B.W. Penninx, F.G. Zitman, Antidepressant use and salivary cortisol in depressive and anxiety disorders, *Eur Neuropsychopharmacol*, 21 (2011) 691-699.
- [216] S. Bano, S. Akhter, M.I. Afridi, Gender based response to fluoxetine hydrochloride medication in endogenous depression, *J Coll Physicians Surg Pak*, 14 (2004) 161-165.
- [217] C.M. Pariante, Risk factors for development of depression and psychosis. Glucocorticoid receptors and pituitary implications for treatment with antidepressant and glucocorticoids, *Ann N Y Acad Sci*, 1179 (2009) 144-152.
- [218] R.H. Perlis, C.M. Beasley, Jr., J.D. Wines, Jr., R.N. Tamura, C. Cusin, D. Shear, J. Amsterdam, F. Quitkin, R.E. Strong, J.F. Rosenbaum, M. Fava, Treatment-associated suicidal ideation and adverse effects in an open, multicenter trial of fluoxetine for major depressive episodes, *Psychother Psychosom*, 76 (2007) 40-46.
- [219] N. Shaparin, P.W. Slattum, I. Bucior, S. Nalamachu, Relationships Among Adverse Events, Disease Characteristics, and Demographics in Treatment of Postherpetic Neuralgia With Gastroretentive Gabapentin, *Clin J Pain*, 31 (2015) 983-991.

- [220] E. Baca, M. Garcia-Garcia, A. Porrás-Chavarino, Gender differences in treatment response to sertraline versus imipramine in patients with nonmelancholic depressive disorders, *Prog Neuropsychopharmacol Biol Psychiatry*, 28 (2004) 57-65.
- [221] E.J.S. Sonuga-Barke, D. Coghill, J.S. Markowitz, J.M. Swanson, M. Vandenberghe, S.J. Hatch, Sex differences in the response of children with ADHD to once-daily formulations of methylphenidate, *J Am Acad Child Adolesc Psychiatry*, 46 (2007) 701-710.
- [222] R.B. Fillingim, T.J. Ness, T.L. Glover, C.M. Campbell, B.A. Hastie, D.D. Price, R. Staud, Morphine responses and experimental pain: sex differences in side effects and cardiovascular responses but not analgesia, *J Pain*, 6 (2005) 116-124.
- [223] N. Pomara, B. Shao, S.J. Choi, H. Tun, R.F. Suckow, Sex-related differences in nortriptyline-induced side-effects among depressed patients, *Prog Neuropsychopharmacol Biol Psychiatry*, 25 (2001) 1035-1048.
- [224] J.H. Peniston, Q. Xiang, E.M. Gould, Factors affecting acceptability of titrated oxymorphone extended release in chronic low back pain - an individual patient analysis, *Curr Med Res Opin*, 26 (2010) 1861-1871.
- [225] R.K. McHugh, E.E. Devito, D. Dodd, K.M. Carroll, J.S. Potter, S.F. Greenfield, H.S. Connery, R.D. Weiss, Gender differences in a clinical trial for prescription opioid dependence, *J Subst Abuse Treat*, 45 (2013) 38-43.
- [226] M.R. Lofwall, P.A. Nuzzo, S.L. Walsh, Effects of cold pressor pain on the abuse liability of intranasal oxycodone in male and female prescription opioid abusers, *Drug and alcohol dependence*, 123 (2012) 229-238.
- [227] B. Vazquez, H. Yang, B. Williams, S. Zhou, A. Laurenza, Perampanel efficacy and safety by gender: Subanalysis of phase III randomized clinical studies in subjects with partial seizures, *Epilepsia*, 56 (2015) e90-e94.
- [228] C.E. Wright, T.L. Sisson, A.K. Ichhpurani, G.R. Peters, Steady-state pharmacokinetic properties of pramipexole in healthy volunteers, *J Clin Pharmacol*, 37 (1997) 520-525.
- [229] N. Yasui-Furukori, S. Tsuchimine, M. Saito, T. Nakagami, Y. Sato, S. Kaneko, Association between major Multidrug Resistance 1 (MDR1) gene polymorphisms and plasma concentration of prolactin during risperidone treatment in schizophrenic patients, *Prog Neuropsychopharmacol Biol Psychiatry*, 31 (2007) 1230-1234.
- [230] N. Yasui-Furukori, M. Saito, S. Tsuchimine, T. Nakagami, Y. Sato, N. Sugawara, S. Kaneko, Association between dopamine-related polymorphisms and plasma concentrations of prolactin during risperidone treatment in schizophrenic patients, *Prog Neuropsychopharmacol Biol Psychiatry*, 32 (2008) 1491-1495.

- [231] J. Usall, D. Suarez, J.M. Haro, Gender differences in response to antipsychotic treatment in outpatients with schizophrenia, *Psychiatry Res*, 153 (2007) 225-231.
- [232] Susan G. Kornstein, M.D. , Alan F. Schatzberg, M.D. , Michael E. Thase, M.D. , Kimberly A. Yonkers, M.D. , James P. McCullough, Ph.D. , Gabor I. Keitner, M.D. , Alan J. Gelenberg, M.D. , Sonia M. Davis, Dr.P.H. , Wilma M. Harrison, M.D. , and, Martin B. Keller, M.D., Gender Differences in Treatment Response to Sertraline Versus Imipramine in Chronic Depression, *American Journal of Psychiatry*, 157 (2000) 1445-1452.
- [233] K.M. Deligiannidis, A.J. Rothschild, B.A. Barton, A.R. Kroll-Desrosiers, B.S. Meyers, A.J. Flint, E.M. Whyte, B.H. Mulsant, S.-P.S. Group, A gender analysis of the study of pharmacotherapy of psychotic depression (STOP-PD): gender and age as predictors of response and treatment-associated changes in body mass index and metabolic measures, *The Journal of clinical psychiatry*, 74 (2013) 1003-1009.
- [234] S.N. Ebert, X.K. Liu, R.L. Woosley, Female gender as a risk factor for drug-induced cardiac arrhythmias: evaluation of clinical and experimental evidence, *J Womens Health*, 7 (1998) 547-557.
- [235] M.D. Drici, B.C. Knollmann, W.X. Wang, R.L. Woosley, Cardiac actions of erythromycin: influence of female sex, *Jama*, 280 (1998) 1774-1776.
- [236] P. Nasveld, S. Kitchener, M. Edstein, K. Rieckmann, Comparison of tafenoquine (WR238605) and primaquine in the post-exposure (terminal) prophylaxis of vivax malaria in Australian Defence Force personnel, *Trans R Soc Trop Med Hyg*, 96 (2002) 683-684.
- [237] B. Damholt, G. Golor, W. Wierich, P. Pedersen, M. Ekblom, M. Zdravkovic, An open-label, parallel group study investigating the effects of age and gender on the pharmacokinetics of the once-daily glucagon-like peptide-1 analogue liraglutide, *J Clin Pharmacol*, 46 (2006) 635-641.
- [238] I. Lee, H.J. Kaminski, T. McPherson, M. Feese, G. Cutter, Gender differences in prednisone adverse effects, Survey result from the MG registry, 5 (2018) e507.
- [239] Y.K. Loke, S. Singh, C.D. Furberg, Long-term use of thiazolidinediones and fractures in type 2 diabetes: a meta-analysis, *Cmaj*, 180 (2009) 32-39.
- [240] S. Liabeuf, V. Gras, J. Moragny, C. Durand-Maugard, K. Masmoudi, M. Andréjak, Trospium chloride for overactive bladder may induce central nervous system adverse events, *European Geriatric Medicine*, 5 (2014) 220-224.
- [241] J. Cassidy, C. Twelves, D. Cameron, W. Steward, K. O'Byrne, D. Jodrell, L. Banken, T. Goggin, D. Jones, B. Roos, E. Bush, E. Weidekamm, B. Reigner, Bioequivalence of two tablet formulations of capecitabine and exploration of age, gender, body surface area, and creatinine

clearance as factors influencing systemic exposure in cancer patients, *Cancer Chemother Pharmacol*, 44 (1999) 453-460.

[242] J.A. Sloan, R.M. Goldberg, D.J. Sargent, D. Vargas-Chanes, S. Nair, S.S. Cha, P.J. Novotny, M.A. Poon, M.J. O'Connell, C.L. Loprinzi, Women experience greater toxicity with fluorouracil-based chemotherapy for colorectal cancer, *J Clin Oncol*, 20 (2002) 1491-1498.

[243] J.A. Sloan, C.L. Loprinzi, P.J. Novotny, S. Okuno, S. Nair, D.L. Barton, Sex differences in fluorouracil-induced stomatitis, *J Clin Oncol*, 18 (2000) 412-420.

[244] P.O. Tuomainen, A. Ylitalo, M. Niemelä, K. Kervinen, M. Pietilä, J. Sia, K. Nyman, W. Nammas, K.E. Airaksinen, P.P. Karjalainen, Gender-based analysis of the 3-year outcome of bioactive stents versus paclitaxel-eluting stents in patients with acute myocardial infarction: an insight from the TITAX-AMI trial, *J Invasive Cardiol*, 24 (2012) 104-108.

[245] G.W. Mikhail, R.T. Gerber, D.A. Cox, S.G. Ellis, J.M. Lasala, J.A. Ormiston, G.W. Stone, M.A. Turco, A.A. Joshi, D.S. Baim, A. Colombo, Influence of sex on long-term outcomes after percutaneous coronary intervention with the paclitaxel-eluting coronary stent: results of the "TAXUS Woman" analysis, *JACC Cardiovasc Interv*, 3 (2010) 1250-1259.

[246] Z. Zelinkova, E. Bultman, L. Vogelaar, C. Bouziane, E.J. Kuipers, C.J. van der Woude, Sex-dimorphic adverse drug reactions to immune suppressive agents in inflammatory bowel disease, *World journal of gastroenterology*, 18 (2012) 6967-6973.

[247] M. Gandhi, F. Aweeka, R.M. Greenblatt, T.F. Blaschke, Sex Differences in Pharmacokinetics and Pharmacodynamics, *Annual Review of Pharmacology and Toxicology*, 44 (2004) 499-523.

[248] V. Regitz-Zagrosek, *Sex and gender differences in pharmacology*, Springer Science & Business Media 2012.

[249] R.R. Makkar, B.S. Fromm, R.T. Steinman, M.D. Meissner, M.H. Lehmann, Female gender as a risk factor for torsades de pointes associated with cardiovascular drugs, *Jama*, 270 (1993) 2590-2597.

[250] A. Parekh, E.O. Fadiran, K. Uhl, D.C. Throckmorton, Adverse effects in women: implications for drug development and regulatory policies, *Expert review of clinical pharmacology*, 4 (2011) 453-466.

[251] ICH, Guidance on E14 Clinical Evaluation of QT/QTc Interval Prolongation and Proarrhythmic Potential for Non-Antiarrhythmic Drugs, Fed Regist, 2005, pp. 61134-61135.

[252] A. Arya, Gender-related differences in ventricular repolarization: beyond gonadal steroids, *Journal of cardiovascular electrophysiology*, 16 (2005) 525-527.

- [253] H. Vierhapper, P. Nowotny, W. Waldhäusl, Sex-specific differences in cortisol production rates in humans, *Metabolism*, 47 (1998) 974-976.
- [254] T.J. Gan, P.S. Glass, J. Sigl, P. Sebel, F. Payne, C. Rosow, P. Embree, Women emerge from general anesthesia with propofol/alfentanil/nitrous oxide faster than men, *Anesthesiology*, 90 (1999) 1283-1287.
- [255] A.R. Eugene, J. Masiak, A pharmacodynamic modelling and simulation study identifying gender differences of daily olanzapine dose and dopamine D2-receptor occupancy, *Nordic journal of psychiatry*, 71 (2017) 417-424.
- [256] K.L. Bigos, B.G. Pollock, K.C. Coley, D.D. Miller, S.R. Marder, M. Aravagiri, M.A. Kirshner, L.S. Schneider, R.R. Bies, Sex, race, and smoking impact olanzapine exposure, *The Journal of Clinical Pharmacology*, 48 (2008) 157-165.
- [257] S.C. Mitchell, R.L. Smith, R.H. Waring, The menstrual cycle and drug metabolism, *Curr Drug Metab*, 10 (2009) 499-507.
- [258] S.A. Ritz, D.M. Antle, J. Cote, K. Deroy, N. Fraleigh, K. Messing, L. Parent, J. St-Pierre, C. Vaillancourt, D. Mergler, First steps for integrating sex and gender considerations into basic experimental biomedical research, *FASEB J*, 28 (2014) 4-13.
- [259] B. Rael, N. Romero-Parra, V.M. Alfaro-Magallanes, L. Barba-Moreno, R. Cupeiro, X. Janse de Jonge, A.B. Peinado, F.S.G. Iron, Body Composition Over the Menstrual and Oral Contraceptive Cycle in Trained Females, *Int J Sports Physiol Perform*, 16 (2020) 375-381.
- [260] N.S. Stachenfeld, D.L. Keefe, Estrogen effects on osmotic regulation of AVP and fluid balance, *Am J Physiol Endocrinol Metab*, 283 (2002) E711-721.
- [261] V.A. Damoiseaux, J.H. Proost, V.C. Jiawan, B.N. Melgert, Sex differences in the pharmacokinetics of antidepressants: influence of female sex hormones and oral contraceptives, *Clin Pharmacokinet*, 53 (2014) 509-519.
- [262] I. Rodriguez, M.J. Kilborn, X.K. Liu, J.C. Pezzullo, R.L. Woosley, Drug-induced QT prolongation in women during the menstrual cycle, *JAMA*, 285 (2001) 1322-1326.
- [263] M. Hara, P. Danilo, Jr., M.R. Rosen, Effects of gonadal steroids on ventricular repolarization and on the response to E4031, *J Pharmacol Exp Ther*, 285 (1998) 1068-1072.
- [264] G.H. Kamimori, N. Sirisuth, D.J. Greenblatt, N.D. Eddington, The influence of the menstrual cycle on triazolam and indocyanine green pharmacokinetics, *J Clin Pharmacol*, 40 (2000) 739-744.
- [265] T.J. Nicolson, H.R. Mellor, R.R. Roberts, Gender differences in drug toxicity, *Trends Pharmacol Sci*, 31 (2010) 108-114.

- [266] S.E. Andrade, J.H. Gurwitz, R.L. Davis, K.A. Chan, J.A. Finkelstein, K. Fortman, H. McPhillips, M.A. Raebel, D. Roblin, D.H. Smith, M.U. Yood, A.N. Morse, R. Platt, Prescription drug use in pregnancy, *Am J Obstet Gynecol*, 191 (2004) 398-407.
- [267] A.M. Moyer, E.T. Matey, V.M. Miller, Individualized medicine: Sex, hormones, genetics, and adverse drug reactions, *Pharmacol Res Perspect*, 7 (2019) e00541.
- [268] R. McGready, K. Stepniewska, E. Seaton, T. Cho, D. Cho, A. Ginsberg, M.D. Edstein, E. Ashley, S. Looareesuwan, N.J. White, F. Nosten, Pregnancy and use of oral contraceptives reduces the biotransformation of proguanil to cycloguanil, *Eur J Clin Pharmacol*, 59 (2003) 553-557.
- [269] J. Mwinyi, I. Cavaco, R.S. Pedersen, A. Persson, S. Burkhardt, S. Mkrtchian, M. Ingelman-Sundberg, Regulation of CYP2C19 expression by estrogen receptor alpha: implications for estrogen-dependent inhibition of drug metabolism, *Mol Pharmacol*, 78 (2010) 886-894.
- [270] J. Elflein, Contraceptive use among women in England 2019/20, by type and age, Statista, 2020.
- [271] J.R. Oesterheld, K. Cozza, N.B. Sandson, Oral contraceptives, *Psychosomatics*, 49 (2008) 168-175.
- [272] I. Tantcheva-Poór, M. Zaigler, S. Rietbrock, U. Fuhr, Estimation of cytochrome P-450 CYP1A2 activity in 863 healthy Caucasians using a saliva-based caffeine test, *Pharmacogenetics and Genomics*, 9 (1999) 131-144.
- [273] M. Sandberg, I. Johansson, M. Christensen, A. Rane, E. Eliasson, The impact of CYP2C9 genetics and oral contraceptives on cytochrome P450 2C9 phenotype, *Drug metabolism and disposition*, 32 (2004) 484-489.
- [274] M.T. Granfors, J.T. Backman, J. Laitila, P.J. Neuvonen, Oral contraceptives containing ethinyl estradiol and gestodene markedly increase plasma concentrations and effects of tizanidine by inhibiting cytochrome P450 1A2, *Clinical Pharmacology & Therapeutics*, 78 (2005) 400-411.
- [275] J. Matthaai, M.V. Tzvetkov, J. Strube, D. Sehr, C. Sachse-Seeboth, J.v.B. Hjelmberg, S. Möller, U. Halekoh, U. Hofmann, M. Schwab, Heritability of caffeine metabolism: Environmental effects masking genetic effects on CYP1A2 activity but not on NAT2, *Clinical Pharmacology & Therapeutics*, 100 (2016) 606-616.
- [276] S. Palovaara, O. Pelkonen, J. Uusitalo, S. Lundgren, K. Laine, Inhibition of cytochrome P450 2B6 activity by hormone replacement therapy and oral contraceptive as measured by bupropion hydroxylation, *Clinical Pharmacology & Therapeutics*, 74 (2003) 326-333.

- [277] K.M. Knights, C.F. McLean, A.L. Tonkin, J.O. Miners, Lack of effect of gender and oral contraceptive steroids on the pharmacokinetics of (R)-ibuprofen in humans, *Br J Clin Pharmacol*, 40 (1995) 153-156.
- [278] W.H. Utian, The International Menopause menopause-related terminology definitions, *Climacteric*, 2 (1999) 284-286.
- [279] A.M. Moyer, E.T. Matey, V.M. Miller, Individualized medicine: Sex, hormones, genetics, and adverse drug reactions, *Pharmacology Research & Perspectives*, 7 (2019) e00541.
- [280] M.F. Paine, S.S. Ludington, M.-L. Chen, P.W. Stewart, S.-M. Huang, P.B. Watkins, Do men and women differ in proximal small intestinal CYP3A or P-glycoprotein expression?, *Drug metabolism and disposition*, 33 (2005) 426-433.
- [281] R.Z. Harris, S.M. Tsunoda, P. Mroczkowski, H. Wong, L.Z. Benet, The effects of menopause and hormone replacement therapies on prednisolone and erythromycin pharmacokinetics, *Clinical Pharmacology & Therapeutics*, 59 (1996) 429-435.
- [282] C.-U. Pae, L. Mandelli, T.-S. Kim, C. Han, P.S. Masand, D.M. Marks, A.A. Patkar, D.C. Steffens, D. De Ronchi, A. Serretti, Effectiveness of antidepressant treatments in premenopausal versus postmenopausal women: a pilot study on differential effects of sex hormones on antidepressant effects, *Biomedicine & pharmacotherapy*, 63 (2009) 228-235.
- [283] C. Barth, A. Villringer, J. Sacher, Sex hormones affect neurotransmitters and shape the adult female brain during hormonal transition periods, *Front Neurosci*, 9 (2015) 37-37.
- [284] GLAAD, GLAAD Media Reference Guide - Transgender, 2021.
- [285] K. Wylie, G. Knudson, S.I. Khan, M. Bonierbale, S. Watanyusakul, S. Baral, Serving transgender people: clinical care considerations and service delivery models in transgender health, *The Lancet*, 388 (2016) 401-411.
- [286] J. Ronda, A. Nord, R. Arrington-Sanders, R. Naik, C.M. Takemoto, J. Baskin, S. Lanzkron, L.H. Pecker, Challenges in the management of the transgender patient with sickle cell disease, *Am J Hematol*, 93 (2018) E360-E362.
- [287] L.R. Cirrincione, K.J. Huang, Sex and gender differences in clinical pharmacology: Implications for transgender medicine, *Clinical Pharmacology & Therapeutics*, n/a (2021).
- [288] L.J. Gooren, K. Wierckx, E.J. Giltay, Cardiovascular disease in transsexual persons treated with cross-sex hormones: reversal of the traditional sex difference in cardiovascular disease pattern, *Eur J Endocrinol*, 170 (2014) 809-819.
- [289] S. Maraka, N. Singh Ospina, R. Rodriguez-Gutierrez, C.J. Davidge-Pitts, T.B. Nippoldt, L.J. Prokop, M.H. Murad, Sex Steroids and Cardiovascular Outcomes in Transgender

Individuals: A Systematic Review and Meta-Analysis, *J Clin Endocrinol Metab*, 102 (2017) 3914-3923.

[290] D. Getahun, R. Nash, W.D. Flanders, T.C. Baird, T.A. Becerra-Culqui, L. Cromwell, E. Hunkeler, T.L. Lash, A. Millman, V.P. Quinn, Cross-sex hormones and acute cardiovascular events in transgender persons: a cohort study, *Annals of internal medicine*, 169 (2018) 205-213.

[291] N.M. Nota, C.M. Wiepjes, C.J. de Blok, L.J. Gooren, B.P. Kreukels, M. den Heijer, Occurrence of acute cardiovascular events in transgender individuals receiving hormone therapy: results from a large cohort study, *Circulation*, 139 (2019) 1461-1462.

[292] A. Radix, J. Sevelius, M.B. Deutsch, Transgender women, hormonal therapy and HIV treatment: a comprehensive review of the literature and recommendations for best practices, *J Int AIDS Soc*, 19 (2016) 20810.

[293] M. Ibarra, M. Vazquez, P. Fagiolino, Sex Effect on Average Bioequivalence, *Clin Ther*, 39 (2017) 23-33.

[294] FDA, Bioavailability and Bioequivalence Requirements, Code of Federal Regulations Title 21, Food and Drug Administration, 2020.

[295] CDER, Guidance for Industry: Bioavailability and Bioequivalence Studies for Orally Administered Drug Products - General Considerations, US Food and Drug Administration, MS, USA, 2003.

[296] EMA, Guideline on the Investigation of Bioequivalence, in: C.f.M.P.f.H. Use (Ed.), European Medicines Agency, London, United Kingdom, 2010.

[297] M.L. Chen, S.C. Lee, M.J. Ng, D.J. Schuirmann, L.J. Lesko, R.L. Williams, Pharmacokinetic analysis of bioequivalence trials: implications for sex-related issues in clinical pharmacology and biopharmaceutics, *Clin Pharmacol Ther*, 68 (2000) 510-521.

[298] J. Flores Perez, H. Juarez Olguin, C. Flores Perez, G. Perez Guille, A. Guille Perez, A. Camacho Vieyra, A. Toledo Lopez, M. Carrasco Portugal, I. Lares Asseff, Effects of gender and phase of the menstrual cycle on the kinetics of ranitidine in healthy volunteers, *Chronobiol Int*, 20 (2003) 485-494.

[299] D. Reker, S.M. Blum, C. Steiger, K.E. Anger, J.M. Sommer, J. Fanikos, G. Traverso, "Inactive" ingredients in oral medications, *Sci Transl Med*, 11 (2019).

[300] D. Reker, Y. Shi, A.R. Kirtane, K. Hess, G.J. Zhong, E. Crane, C.H. Lin, R. Langer, G. Traverso, Machine Learning Uncovers Food- and Excipient-Drug Interactions, *Cell Rep*, 30 (2020) 3710-3716 e3714.

- [301] A.W. Basit, J.M. Newton, M.D. Short, W.A. Waddington, P.J. Ell, L.F. Lacey, The effect of polyethylene glycol 400 on gastrointestinal transit: implications for the formulation of poorly-water soluble drugs, *Pharm Res*, 18 (2001) 1146-1150.
- [302] J.D. Schulze, W.A. Waddington, P.J. Eli, G.E. Parsons, M.D. Coffin, A.W. Basit, Concentration-dependent effects of polyethylene glycol 400 on gastrointestinal transit and drug absorption, *Pharm Res*, 20 (2003) 1984-1988.
- [303] Y. Mai, D.A.I. Ashiru-Oredope, Z. Yao, L. Dou, C.M. Madla, F. Taherali, S. Murdan, A.W. Basit, Boosting drug bioavailability in men but not women through the action of an excipient, *Int J Pharm*, 587 (2020) 119678.
- [304] A.J. McGregor, J.S. Markowitz, J. Forrester, R.I. Shader, Joining the Effort: The Challenges in Establishing Guidelines for Sex- and Gender-specific Research Design in Clinical Therapeutic Studies, *Clin Ther*, 39 (2017) 1912-1916.
- [305] M. Koziolk, S. Alcaro, P. Augustijns, A.W. Basit, M. Grimm, B. Hens, C.L. Hoad, P. Jedamzik, C.M. Madla, M. Maliepaard, L. Marciani, A. Maruca, N. Parrott, P. Pávek, C.J.H. Porter, C. Reppas, D. van Riet-Nales, J. Rubbens, M. Statelova, N.L. Trevaskis, K. Valentová, M. Vertzoni, D.V. Čepo, M. Corsetti, The mechanisms of pharmacokinetic food-drug interactions – A perspective from the UNGAP group, *European Journal of Pharmaceutical Sciences*, 134 (2019) 31-59.
- [306] E. Prewett, J. Smith, C. Nwokolo, A. Sawyerr, R. Pounder, Twenty-four hour intragastric acidity and plasma gastrin concentration profiles in female and male subjects, *Clinical Science*, 80 (1991) 619-624.
- [307] L. Dou, F.K.H. Gavins, Y. Mai, C.M. Madla, F. Taherali, M. Orlu, S. Murdan, A.W. Basit, Effect of Food and an Animal's Sex on P-Glycoprotein Expression and Luminal Fluids in the Gastrointestinal Tract of Wistar Rats, *Pharmaceutics*, 12 (2020).
- [308] L. Dou, Y. Mai, C.M. Madla, M. Orlu, A.W. Basit, P-glycoprotein expression in the gastrointestinal tract of male and female rats is influenced differently by food, *Eur J Pharm Sci*, 123 (2018) 569-575.
- [309] F. Kees, M. Bucher, F. Schweda, H. Gschaidmeier, L. Faerber, R. Seifert, Neoimmun versus Neoral: a bioequivalence study in healthy volunteers and influence of a fat-rich meal on the bioavailability of Neoimmun, *Naunyn-Schmiedeberg's archives of pharmacology*, 375 (2007) 393-399.
- [310] P. Bhupathy, C.D. Haines, L.A. Leinwand, Influence of sex hormones and phytoestrogens on heart disease in men and women, *Womens Health (Lond)*, 6 (2010) 77-95.

- [311] C. Tannenbaum, D. Day, A. Matera, Age and sex in drug development and testing for adults, *Pharmacol Res*, 121 (2017) 83-93.
- [312] J.R. Docherty, S.C. Stanford, R.A. Panattieri, S.P.H. Alexander, G. Cirino, C.H. George, D. Hoyer, A.A. Izzo, Y. Ji, E. Lilley, C.G. Sobey, P. Stanley, B. Stefanska, G. Stephens, M. Teixeira, A. Ahluwalia, Sex: A change in our guidelines to authors to ensure that this is no longer an ignored experimental variable, *Br J Pharmacol*, 176 (2019) 4081-4086.
- [313] T. von Erlach, S. Saxton, Y. Shi, D. Minahan, D. Reker, F. Javid, Y.-A.L. Lee, C. Schoellhammer, T. Esfandiary, C. Cleveland, L. Booth, J. Lin, H. Levy, S. Blackburn, A. Hayward, R. Langer, G. Traverso, Robotically handled whole-tissue culture system for the screening of oral drug formulations, *Nature Biomedical Engineering*, 4 (2020) 544-559.
- [314] V.A. Welch, O.F. Norheim, J. Jull, R. Cookson, H. Sommerfelt, P. Tugwell, C. Equity, S. Boston Equity, CONSORT-Equity 2017 extension and elaboration for better reporting of health equity in randomised trials, *BMJ*, 359 (2017) j5085.
- [315] V. Welch, M. Petticrew, P. Tugwell, D. Moher, J. O'Neill, E. Waters, H. White, P.R.-E.B. group, PRISMA-Equity 2012 extension: reporting guidelines for systematic reviews with a focus on health equity, *PLoS Med*, 9 (2012) e1001333.
- [316] K. Shah, C.E. McCormack, N.A. Bradbury, Do you know the sex of your cells?, *Am J Physiol Cell Physiol*, 306 (2014) C3-18.
- [317] C. Hartmanshenn, M. Scherholz, I.P. Androulakis, Physiologically-based pharmacokinetic models: approaches for enabling personalized medicine, *Journal of Pharmacokinetics and Pharmacodynamics*, 43 (2016) 481-504.
- [318] P. Arora, G. Gudelsky, P.B. Desai, Gender-based differences in brain and plasma pharmacokinetics of letrozole in sprague-dawley rats: Application of physiologically-based pharmacokinetic modeling to gain quantitative insights, *PLOS ONE*, 16 (2021) e0248579.
- [319] G. Koren, H. Nordeng, S. MacLeod, Gender differences in drug bioequivalence: time to rethink practices, *Clinical Pharmacology & Therapeutics*, 93 (2013) 260-262.
- [320] M. Elbadawi, B. Muñiz Castro, F.K.H. Gavins, J.J. Ong, S. Gaisford, G. Pérez, A.W. Basit, P. Cabalar, A. Goyanes, M3DISEEN: A novel machine learning approach for predicting the 3D printability of medicines, *International Journal of Pharmaceutics*, 590 (2020) 119837.
- [321] M. Elbadawi, S. Gaisford, A.W. Basit, Advanced machine-learning techniques in drug discovery, *Drug Discovery Today*, (2020).
- [322] M. Davies, R.D.O. Jones, K. Grime, R. Jansson-Löfmark, A.J. Fretland, S. Winiwarter, P. Morgan, D.F. McGinnity, Improving the Accuracy of Predicted Human Pharmacokinetics:

Lessons Learned from the AstraZeneca Drug Pipeline Over Two Decades, *Trends in Pharmacological Sciences*, 41 (2020) 390-408.

[323] E. Callaway, 'It will change everything': DeepMind's AI makes gigantic leap in solving protein structures, *Nature*, 588 (2020) 203-204.

[324] D. Cirillo, S. Caturana-Solarz, C. Morey, E. Guney, L. Subirats, S. Mellino, A. Gigante, A. Valencia, M.J. Rementeria, A.S. Chadha, N. Mavridis, Sex and gender differences and biases in artificial intelligence for biomedicine and healthcare, *npj Digital Medicine*, 3 (2020) 81.

[325] C. Huang, E.A. Clayton, L.V. Matyunina, L.D. McDonald, B.B. Benigno, F. Vannberg, J.F. McDonald, Machine learning predicts individual cancer patient responses to therapeutic drugs with high accuracy, *Scientific Reports*, 8 (2018) 16444.

[326] N. Rohani, C. Eslahchi, Drug-Drug Interaction Predicting by Neural Network Using Integrated Similarity, *Scientific Reports*, 9 (2019) 13645.

[327] K. Raja, M. Patrick, J.T. Elder, L.C. Tsoi, Machine learning workflow to enhance predictions of Adverse Drug Reactions (ADRs) through drug-gene interactions: application to drugs for cutaneous diseases, *Scientific Reports*, 7 (2017) 3690.

[328] N. Tomašev, X. Glorot, J.W. Rae, M. Zielinski, H. Askham, A. Saraiva, A. Mottram, C. Meyer, S. Ravuri, I. Protsyuk, A. Connell, C.O. Hughes, A. Karthikesalingam, J. Cornebise, H. Montgomery, G. Rees, C. Laing, C.R. Baker, K. Peterson, R. Reeves, D. Hassabis, D. King, M. Suleyman, T. Back, C. Nielson, J.R. Ledsam, S. Mohamed, A clinically applicable approach to continuous prediction of future acute kidney injury, *Nature*, 572 (2019) 116-119.

[329] N. Desai, A.J. Edwards, T.B. Ernest, C. Tuleu, M. Orlu, 'Big Data' informed drug development: a case for acceptability, *Drug Discovery Today*, (2020).

[330] Whose genomics?, *Nature Human Behaviour*, 3 (2019) 409-410.

[331] A. Coravos, S. Khozin, K.D. Mandl, Developing and adopting safe and effective digital biomarkers to improve patient outcomes, *npj Digital Medicine*, 2 (2019) 14.

[332] J.M. Ramsey, J.D. Cooper, B.W.J.H. Penninx, S. Bahn, Variation in serum biomarkers with sex and female hormonal status: implications for clinical tests, *Scientific Reports*, 6 (2016) 26947.

[333] W.N. Price, Big data and black-box medical algorithms, *Science translational medicine*, 10 (2018).

[334] D. Gemmati, K. Varani, B. Bramanti, R. Piva, G. Bonaccorsi, A. Trentini, M.C. Manfrinato, V. Tisato, A. Carè, T. Bellini, "Bridging the Gap" Everything that Could Have

Been Avoided If We Had Applied Gender Medicine, Pharmacogenetics and Personalized Medicine in the Gender-Omics and Sex-Omics Era, *Int J Mol Sci*, 21 (2019).

[335] M.A. Alhnan, E. Kidia, A.W. Basit, Spray-drying enteric polymers from aqueous solutions: A novel, economic, and environmentally friendly approach to produce pH-responsive microparticles, *European Journal of Pharmaceutics and Biopharmaceutics*, 79 (2011) 432-439.

[336] A.M. Vargason, A.C. Anselmo, S. Mitragotri, The evolution of commercial drug delivery technologies, *Nature Biomedical Engineering*, (2021).

[337] P.M. Coalition, *The Case for Personalized Medicine*, 4th Edition ed.2014.

[338] G.C. S. Sharifi, D. Pozzi, L. Digiacomo, J. Swann, H.E. Daldrup-Link, M.Mahmoudi,, The role of sex as a biological variable in the efficacy and toxicity of therapeutic nanomedicine, *Advanced Drug Delivery Reviews*, (2021).

[339] R. Govender, S. Abrahmsen-Alami, A. Larsson, S. Folestad, Therapy for the individual: Towards patient integration into the manufacturing and provision of pharmaceuticals, *Eur J Pharm Biopharm*, 149 (2020) 58-76.

[340] S.J. Trenfield, A. Awad, C.M. Madla, G.B. Hatton, J. Firth, A. Goyanes, S. Gaisford, A.W. Basit, Shaping the future: recent advances of 3D printing in drug delivery and healthcare, *Expert Opin Drug Deliv*, 16 (2019) 1081-1094.

[341] C.I. Gioumouxouzis, C. Karavasili, D.G. Fatouros, Recent advances in pharmaceutical dosage forms and devices using additive manufacturing technologies, *Drug Discovery Today*, 24 (2019) 636-643.

[342] S.J. Trenfield, A. Awad, A. Goyanes, S. Gaisford, A.W. Basit, 3D Printing Pharmaceuticals: Drug Development to Frontline Care, *Trends Pharmacol Sci*, 39 (2018) 440-451.

[343] A.J. Capel, R.P. Rimington, M.P. Lewis, S.D.R. Christie, 3D printing for chemical, pharmaceutical and biological applications, *Nature Reviews Chemistry*, 2 (2018) 422-436.

[344] J. Norman, R.D. Madurawe, C.M.V. Moore, M.A. Khan, A. Khairuzzaman, A new chapter in pharmaceutical manufacturing: 3D-printed drug products, *Advanced Drug Delivery Reviews*, 108 (2017) 39-50.

[345] A. Melocchi, F. Briatico-Vangosa, M. Uboldi, F. Parietti, M. Turchi, D. von Zeppelin, A. Maroni, L. Zema, A. Gazzaniga, A. Zidan, Quality considerations on the pharmaceutical applications of fused deposition modeling 3D printing, *International Journal of Pharmaceutics*, 592 (2021) 119901.

- [346] A. Awad, F. Fina, A. Goyanes, S. Gaisford, A.W. Basit, Advances in powder bed fusion 3D printing in drug delivery and healthcare, *Advanced Drug Delivery Reviews*, (2021).
- [347] J. Boetker, J.J. Water, J. Aho, L. Arnfast, A. Bohr, J. Rantanen, Modifying release characteristics from 3D printed drug-eluting products, *Eur J Pharm Sci*, 90 (2016) 47-52.
- [348] P. Januskaite, X. Xu, S.R. Ranmal, S. Gaisford, A.W. Basit, C. Tuleu, A. Goyanes, I Spy with My Little Eye: A Paediatric Visual Preferences Survey of 3D Printed Tablets, *Pharmaceutics*, 12 (2020).
- [349] X. Xu, P. Robles-Martinez, C.M. Madla, F. Joubert, A. Goyanes, A.W. Basit, S. Gaisford, Stereolithography (SLA) 3D printing of an antihypertensive polyprintlet: Case study of an unexpected photopolymer-drug reaction, *Additive Manufacturing*, 33 (2020) 101071.
- [350] M.A. Alhnan, T.C. Okwuosa, M. Sadia, K.-W. Wan, W. Ahmed, B. Arafat, Emergence of 3D Printed Dosage Forms: Opportunities and Challenges, *Pharmaceutical Research*, 33 (2016) 1817-1832.
- [351] A. Goyanes, C.M. Madla, A. Umerji, G. Duran Piñeiro, J.M. Giraldez Montero, M.J. Lamas Diaz, M. Gonzalez Barcia, F. Taherali, P. Sánchez-Pintos, M.-L. Couce, S. Gaisford, A.W. Basit, Automated therapy preparation of isoleucine formulations using 3D printing for the treatment of MSUD: First single-centre, prospective, crossover study in patients, *International Journal of Pharmaceutics*, 567 (2019) 118497.
- [352] K. Vithani, A. Goyanes, V. Jannin, A.W. Basit, S. Gaisford, B.J. Boyd, An Overview of 3D Printing Technologies for Soft Materials and Potential Opportunities for Lipid-based Drug Delivery Systems, *Pharmaceutical Research*, 36 (2018) 4.
- [353] A. Awad, S.J. Trenfield, A. Goyanes, S. Gaisford, A.W. Basit, Reshaping drug development using 3D printing, *Drug Discov Today*, 23 (2018) 1547-1555.

