

Why disease ecology needs life-history theory: a host perspective

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Complete List of Authors:	Valenzuela-Sánchez, Andrés; Universidad Andres Bello, Centro de Investigación para la Sustentabilidad; ONG Ranita de Darwin, Wilber, Mark; University of California, Santa Barbara, Ecology, Evolution, and Marine Biology Canessa, Stefano; Universiteit Gent Faculteit Diergeneeskunde, Wildlife Health Ghent Bacigalupe, Leonardo Muths, Erin; USGS, Amphibian Research and Monitoring Initiative (ARMI) Schmidt, Benedikt; University of Zürich, Evolutionary Biology; karch, Cunningham, Andrew; Zoological Society of London, Institute of Zoology Ozgul, Arpat; University of Zurich, Department of Evolutionary Biology & Environmental Studies Johnson, Pieter; University of Colorado, Ecology and Evolutionary Biology Cayuela, Hugo; Université Laval, Biology



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2 3 4 5	1	Why disease ecology needs life-history theory: a host perspective
6 7 8	2	Andrés Valenzuela-Sánchez ^{1,2,3*} , Mark Q. Wilber ⁴ , Stefano Canessa ⁵ , Leonardo D.
9 10	3	Bacigalupe ¹ , Erin Muths ⁶ , Benedikt R. Schmidt ^{7,8} , Andrew A. Cunningham ⁹ , Arpat Ozgul ⁷ ,
11 12 12	4	Pieter T. J. Johnson ¹⁰ , Hugo Cayuela ¹¹
13 14 15	5	¹ Instituto de Ciencias Ambientales y Evolutivas, Universidad Austral de Chile, Valdivia, Chile
16 17 18	6	² ONG Ranita de Darwin, Valdivia and Santiago, Chile
19 20 21	7	³ Centro de Investigación para la Sustentabilidad, Universidad Andrés Bello, Santiago, Chile
22 23	8	⁴ Ecology, Evolution, and Marine Biology, University of California, Santa Barbara, Santa Barbara,
24 25 26	9	CA, 93106
27 28 29	10	⁵ Wildlife Health Ghent, Faculty of Veterinary Medicine, Ghent University, Merelbeke, Belgium
30 31	11	⁶ U.S. Geological Survey, 2150 Centre Avenue Bldg C, Fort Collins, Colorado, 80526 USA
32 33 34	12	⁷ Institut für Evolutionsbiologie und Umweltwissenschaften, Universität Zürich, Winterthurerstrasse
35 36 37	13	190, 8057 Zürich, Switzerland
38 39	14	⁸ Info Fauna Karch, UniMail, Bâtiment G, Bellevaux 51, 2000 Neuchâtel, Switzerland
40 41 42	15	⁹ Institute of Zoology, Zoological Society of London, Regent's Park, London NW1 4RY, UK
43 44	16	¹⁰ Department of Ecology and Evolutionary Biology, University of Colorado, Boulder, Colorado,
45 46 47	17	80309, USA
48 49	18	¹¹ IBIS, Department of Biology, University Laval, Pavillon Charles-Eugène-Marchand, Avenue de
50 51 52	19	la Médecine, Quebec City, Canada
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- 31 Author for correspondence: Andrés Valenzuela-Sánchez (e-mail:
- 32 andresvalenzuela.zoo@gmail.com; telephone: + 56 9 50014215; address: Av. Rector
- 33 Eduardo Morales s/n, Edificio Emilio Pugín, Universidad Austral de Chile, Isla Teja,
- 34 Valdivia, Chile)
- 35 Authorship: AV-S conceived the study and all the authors contributed novel ideas and
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42 Abstract

When facing an emerging infectious disease of conservation concern, we often have little information on the nature of the host-parasite interaction to inform management decisions. However, it is becoming increasingly clear that the life-history strategies of host species can be predictive of individual- and population-level responses to infectious disease, even without detailed knowledge on the specifics of the host-parasite interaction. Here, we argue that a deeper integration of life-history theory into disease ecology is timely and necessary to improve our capacity to understand, predict, and mitigate the impact of endemic and emerging infectious diseases in wild populations. Using wild vertebrates as an example, we show that host life-history characteristics influence host responses to parasitism at different levels of organization, from individuals to communities. We also highlight knowledge gaps and future directions for the study of life-history and host responses to parasitism. We conclude by illustrating how this theoretical insight can inform the monitoring and control of infectious diseases in wildlife.

57 Keywords

demography, demographic compensation, outbreak, pace of life, pathogen, slow-fastcontinuum, vertebrates

NOVELTY

We present a novel synthesis on the intersection of life-history and host responses to parasitism, to demonstrate that a deeper integration of life-history theory into disease ecology is a fruitful avenue of research to advance the understanding and mitigation of wildlife infectious diseases. This synthesis highlights that life-history strategies can lead to a variety of host responses to parasitism, modulating host immune responses, the mechanisms of host demographic compensation, the potential for rapid evolution of resistance or tolerance mechanisms, and the efficiency of parasite transmission and disease varasite systems.

 risk in multi-host parasite systems.

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69 INTRODUCTION

Infectious diseases are an important threat to biodiversity (Daszak et al., 2000). This is particularly true for emerging infectious diseases, for which the lack of host-parasite coevolutionary history can lead to extreme levels of parasite virulence and/or host susceptibility, ultimately inducing strong population-level impacts (e.g., Daszak et al., 2000, 2001; Fisher et al., 2012; Scheele et al., 2019a). Nonetheless, empirical evidence further reveals that host population collapse is not the only outcome from a novel host-parasite interaction (Tompkins et al., 2011). Some populations of susceptible hosts can persist despite initial marked population declines (e.g., fish, Rogowski et al., 2020; amphibians, Briggs et al., 2010; marsupials, Wells et al., 2019). Understanding the factors that determine these alternative, sometimes contrasting, population-level impacts of infectious disease has interested disease ecologists for decades and numerous factors about the parasite, the host, and the environment have been identified as important in the dynamics of host-parasite systems (Fig. 1; Anderson and May, 1979; Tompkins et al., 2011).

We argue that a deeper integration of life-history theory (hereafter LHT) into disease ecology is both timely and necessary to improve our capacity to understand, predict, and mitigate the impact of endemic and emerging infectious diseases in wild populations. A related approach that has provided a fruitful avenue of research is the study of how epidemiological parameters, such as parasite transmission rates (De Leo and Dobson, 1996), epidemiological thresholds (Bolzoni et al., 2018), and host competence (Downs et al., 2019), scale allometrically with host body size. As body size is the main factor shaping interspecific variation in life-history traits (Gaillard et al., 2016; Healy et al.,

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92	2019), the allometric scaling of epidemiological parameters with host body size is expected
93	to be, at least partially, associated with host life-history characteristics. Yet, body size is not
94	always an accurate proxy of host life-history traits, especially when high-level taxonomic
95	ranks (e.g., class level or higher) are considered. For example, within mammals, humans
96	and bats show a particularly long lifespan and low fecundity for their relatively small body
97	sizes (Gaillard et al., 2016; Healy et al., 2019). Indeed, after controlling for allometric
98	constraints, considerable interspecific variation in life-history traits remains and other
99	factors, such as life-history trade-offs, phylogeny, and mode of life, are known to play
100	important roles in shaping the diversity of host life histories (Gaillard et al., 2016; Healy et
101	al., 2019). Here, we argue that the position of a host species along the classical slow-fast
102	life-history continuum (see below) can determine their response to parasitic infection (Fig.
103	1). It is worth noting that other host traits, such as population density and the level of
104	sociality (Han et al., 2015, 2020), as well as parasite life-history traits (Barrett et al., 2008;
105	Silk and Hodgson, 2020), also play critical roles in host-parasite dynamics, but those
106	aspects are beyond the scope of this review.

We focus this review on LHT predictions relative to host responses to infectious 107 108 disease at different levels of organization, from individual-level susceptibility to host 109 community assembly (Fig. 1). Although these theoretical predictions are broad in scope, 110 with empirical validations in plant and animal species (e.g., plants, Pagán et al., 2008; invertebrates, Agnew et al., 2008; vertebrates, Johnson et al., 2012), we emphasize 111 examples in wild vertebrate hosts, a group largely underrepresented in previous syntheses 112 on the intersection of life-history and host responses to parasitism (e.g., Michalakis and 113 Hochberg, 1994; Agnew et al., 2000). The review is structured in eight sections. In the first 114

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115	section, we introduce the theory and empirical evidence supporting the existence of a slow-
116	fast continuum of life-history variation in vertebrates. In the second section, which is
117	related to the field of ecoimmunology (see Brock et al., 2014), we briefly discuss how the
118	position of hosts along the slow-fast continuum can help predict the type and strength of
119	host immune defences (for more detailed coverage refer to previous reviews, e.g., Lee
120	(2006), Martin et al. (2006), Tieleman (2018), and Albery and Becker (2020). In the third
121	section, we discuss how life-history constrains the speed of recovery of host populations
122	after short-term disturbances such as disease outbreaks. In the fourth section, we focus on
123	active demographic compensation, a process particularly relevant for the persistence of host
124	populations impacted by emerging infectious diseases. We define the types of active
125	demographic compensation in the context of infectious diseases and discuss how these
126	responses could be modulated by host life histories, introducing a simple theoretical model
127	to illustrate how life-history strategies can be predictive of the magnitude of the negative
128	effects of disease-induced mortality on populations exhibiting density-dependent
129	compensation. In the fifth section, we discuss how host life-history strategies could
130	modulate the rapid evolution of mechanisms of resistance (i.e., the ability of a host to limit
131	or reduce parasite burden) or tolerance (i.e., the ability of a host to limit the negative effects
132	of a given parasite burden). In the sixth section, we briefly review the integration of host
133	life-history, community assembly, and infectious disease. In the seventh section, we discuss
134	how the insights of the previous sections can inform the monitoring and control of
135	infectious diseases in wildlife. In the eighth and concluding section we provide pointers for
136	future directions for the incorporation of LHT in disease ecology.
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SECTION 1: LIFE-HISTORY TRADE-OFFS AND THE SLOW-FAST **CONTINUUM OF LIFE-HISTORY VARIATION**

The pervasiveness of life-history trade-offs (i.e., beneficial change in one life-history trait has a negative impact on another trait) has been central to the development of classical LHT (Stearns, 1989a). The idea of these trade-offs is grounded in the "principle of allocation" of time and energy (Cody, 1966), such that organisms have a limited amount of time and energy to expend, and natural selection acts as a force operating on the allocation of resources to different functions (e.g., growth, reproduction, locomotion, immune function) to maximize fitness (Ricklefs and Wikelski, 2002; Lee, 2006). The most prominent and well-supported life-history trade-offs involve survival and reproduction (Stearns, 1989a; Lebreton, 2006; Healy et al., 2019). The covariation among traits related to survival and reproduction are structured along a major axis of life-history variation termed the slow-fast life-history continuum (Fig. 1): species at the fast end of the continuum are characterized by high fecundity per time unit (e.g., annual fecundity), early age at first reproduction, and short lifespan, while the opposite is expected for species at the slow end (Gaillard et al., 2016).

It has been proposed that the concept of the slow-fast life-history continuum should be restricted to the pattern of covariation in raw (i.e., size-uncorrected) life-history traits sharing the dimension of time (Jeschke and Kokko, 2009; Gaillard et al., 2016). Empirical evidence supports the existence of this "raw" slow-fast continuum in mammals and birds (Herrando-Pérez et al., 2012; Gaillard et al., 2016), while in amphibians and reptiles a comprehensive analysis on the subject is still lacking (Gaillard et al., 2016; but see Fig. 2 in Herrando-Pérez *et al.* (2012) which suggests the existence of the continuum in these taxa).

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In contrast, for fish species, annual fecundity appears to covary positively with pace of life metrics, although for a given position on the slow-fast continuum the interspecific variation in annual fecundity is notoriously high (see Fig. 2 in Herrando-Pérez et al. (2012)). Theory to better understand this counterintuitive "slow-type survival with fast-type reproduction" strategy observed in several fish species is beginning to emerge (see Wright *et al.*, 2020). It is also worth noting that resources can be allocated to facets of reproduction other than fecundity, such as offspring quality and parental investment, a situation that might lead to a lack of covariation between fecundity and pace of life metrics in some ectotherms (Healy et al., 2019). How this deviation from the classical slow-fast continuum modulates the effect of life histories on host responses to infectious disease is still an untapped question.

172 SECTION 2: HOST LIFE-HISTORY AND IMMUNE DEFENCES

The principle of allocation and the ubiquity of parasites suggest that immune defences should covary with the position of a species on the slow-fast life-history continuum, with fast-living species trading investment in immune defence for growth and reproduction (Lee, 2006; Martin et al., 2006). In contrast, the longer lifespan of slow-living species means that they: 1) may encounter more individual infections during their lifetime, increasing the benefits of allocating resources to immune defences; and 2) may encounter a wider range/diversity of infections (e.g., Gutiérrez et al., 2019), creating selective pressures for adaptive (specific, less self-damaging) immunity (Lee, 2006; Woodhams et al., 2016).

181 The differential allocation of energy and resources to immunity between fast-living182 and slow-living species suggests that when exposed to the same parasite and under similar

environmental conditions, individuals from species at different positions along the slow-fast life-history continuum should exhibit different susceptibilities to acquiring infection and developing disease (Joseph et al., 2013). There is evidence of this relationship in two recent experimental studies in amphibians. In the first study, Johnson et al. (2012) experimentally exposed individuals of 13 amphibian species to the trematode *Ribeiroia* ondatrae. They showed that fast-living species were more prone to infection and the development of lesions than slow-living species. In the second study, using a standardized challenge of 20 North American amphibian species, Gervasi et al. (2017) found that individuals from fast-living species were more susceptible to lethal *B. dendrobatidis* infection.

Empirical evidence also reveals that the relative importance of coarse immunity components, which differ in terms of energetic investment (e.g., innate vs adaptive immunity), tend to vary along the slow-fast continuum in vertebrates, with fast-living species favouring components that can be less costly such as innate immunity and behavioural mechanisms of resistance/tolerance (Fig. 1; Lee 2006; Tieleman et al., 2005; Martin et al., 2006; Previtali et al., 2012; Sears et al., 2015; Woodhams et al., 2016; but see Tieleman (2018) for mixed empirical support for a link between host life-history and host immunity in birds).

Although we have highlighted empirical evidence supporting the covariation of immunity with host life-history strategies (i.e., fast-living species tend to invest less in immunity and to favour less costly mechanisms of resistance/tolerance), there is little robust evidence to support the generality of these patterns in vertebrates or other taxa (Albery and Becker, 2020). Given the complexity of the vertebrate immune system (Brock

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et al., 2014) and its high responsiveness to environmental conditions (e.g., food availability, temperature, microbial environment; Sandland and Minchella, 2003: Palacios et al., 2011), providing robust validation to these LHT predictions is not a trivial task. Such validations could represent a major advance in the study of wildlife diseases, allowing improvements in forecasting host susceptibility to novel parasitic infections and assisting the design of disease mitigation strategies (e.g., mass vaccination or habitat management targeting behavioural resistance/tolerance mechanisms, Hettyey et al., 2019). SECTION 3: HOST LIFE-HISTORY CONSTRAINS POPULATION RECOVERY AFTER A DISEASE OUTBREAK The ability of populations to recover from short-term disturbances such as disease outbreaks depends on their demographic resilience (i.e., the inherent ability of a population to prevent a decrease in size after a disturbance; reviewed in Capdevila et al. (2020)). An important prediction in the context of infectious diseases is that, all else being equal, a population of a slow-living species would require a longer time to recover in size after a disease outbreak than a population of a fast-living species (Lebreton, 2006; Capdevila et al., 2020; see Benhaiem et al. (2018) for an example in mammalian hosts). This arises because the maximum population growth rate, which sets the upper limit of the speed of recovery, is expected to decrease towards the slow end of the life-history continuum (Niel and Lebreton, 2005; Lebreton, 2006). Capdevila et al. (2020) introduced an analytical

framework to study demographic resilience and its components that can be used to provide

further empirical support to the above-mentioned prediction. This approach, however, is

based on the analysis of density-independent, time-invariant matrix population models and

229	does not consider changes in vital rates over time (Capdevila et al., 2020). In the following
230	sections, we show that compensatory changes in vital rates over time are important in
231	determining the resilience of host populations to emerging and endemic infectious diseases,
232	especially considering that parasites often operate as long-term sustained perturbations
233	(e.g., endemic infection dynamics).
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235	SECTION 4: HOST LIFE-HISTORY INFLUENCES THE MECHANISM OF
236	ACTIVE DEMOGRAPHIC COMPENSATION
237	Active demographic compensation (defined as the change in one or more demographic
238	rates [e.g., survival, recruitment] to compensate for a reduction in that, or another,
239	demographic rate) determines the capacity of a population to counteract the detrimental
240	effects of infectious diseases. We use "active" to differentiate this concept from Capdevila
241	et al. (2020)'s definition of demographic compensation which focuses on changes in
242	demographic structure rather than changes in the vital rates. We identify two general
243	mechanisms of active demographic compensation in response to infectious disease: 1) a
244	non-specific response that arises from the effect of parasitic infection on host population
245	density (i.e., density-dependent compensation); and 2) an adaptive plastic response of
246	individual hosts to infection (i.e., parasite-induced plasticity of life-history traits).
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248	The role of density-dependent processes in active demographic compensation
249	Early theoretical studies showed that density-dependent compensation could be a key
250	demographic mechanism to offset disease impacts on population growth rate, an idea that
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was supported by empirical evidence in invertebrate hosts (Anderson and May 1981). Essentially, for a parasite to regulate a host population, disease-induced mortality needs to be additive (i.e., any individual that dies from the disease would have survived if the disease was not present) rather than compensatory (i.e., any individual that dies from the disease would have died from other causes if the disease was not present) to other natural sources of density-dependent mortality (Jolles et al., 2006). For example, in overcrowded populations, parasite infection may primarily remove individuals that otherwise would die due to causes linked to overcrowding (e.g., food or space limitations), resulting in negligible differences in net survival rates between infected and uninfected populations (Kistner and Belovsky, 2014). Additionally, a reduction in population size can boost recruitment in a density-dependent fashion, compensating for the reduced survival of infected hosts (e.g., Anderson and May, 1981; Ohlberger et al., 2011; McDonald et al., 2016: Rogowski et al., 2020). In a population of susceptible hosts, density-dependent compensatory responses can lead to effective compensation (i.e., no change in population size) or even overcompensation (i.e., increase in population size; reviewed in Schröder et al. (2014)). To our knowledge, however, there is only a single demonstration of density-dependent overcompensation in this context, involving a protozoan parasite and a larval mosquito host (Washburn et al., 1991).

The demographic buffering hypothesis states that to alleviate negative effects of environmental stochasticity on the long-run population growth rate, vital rates with the largest contribution to the population growth rate (i.e., vital rates exhibiting a high elasticity) should be buffered against environmental variation (reviewed in Hilde *et al.* (2020)). This means that vital rates with a high elasticity should be canalised (i.e., are

274	insensitive to environmental variation). From this hypothesis, McDonald et al. (2016)
275	proposed that vital rates with high elasticity could also be buffered against internal
276	pressures, exhibiting weak dependence on local population density. This knowledge can
277	help predict the type of density-dependent compensatory mechanisms likely to occur in a
278	host population. In the context of host-parasite systems, this hypothesis suggests that, in
279	slow-living species, recruitment should be more sensitive than adult survival to local
280	population density and density-dependent recruitment should be more commonly observed
281	as a mechanism of compensation for infectious diseases (McDonald et al., 2016). This
282	contrasts with fast-living species, where recruitment is expected to be less sensitive to
283	population density than adult survival and, therefore, density-dependent compensatory
284	recruitment would be expected to be less effective and thus less commonly observed.
285	Instead, in fast-living species, as adult survival is expected to be more sensitive to
286	population density, the increased mortality rates due to disease are more likely to be
287	compensatory rather than additive. In a rare test of these LHT predictions, McDonald et al.
288	(2016) found that density-dependent compensatory recruitment contributed to the
289	persistence of a badger (Meles meles) population naturally infected with the bacterium
290	Mycobacterium bovis (Fig. 2a). This response is in accordance with the above-mentioned
291	LHT predictions, as badgers exhibit a slow life-history strategy (McDonald et al., 2016).
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293	Life-history strategies and host population depression due to parasite-induced mortality

We use a susceptible-infected disease model to theoretically explore how life-history
modulates the negative effects of disease on host populations exhibiting density-dependent
compensation. The purpose of this analysis is to illustrate that distinguishing between slow

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and fast-living life-history strategies can help predict the magnitude of the negative effects

of disease-induced mortality on host populations. The models we analyse here are similar to

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9	those used in previous studies of disease-induced depression of host populations and the
0	effects of life-history on disease dynamics (e.g., Anderson and May, 1981; De Leo and
1	Dobson, 1996; Lloyd-Smith et al., 2005; Bolzoni et al., 2008, Han et al., 2015). The key
2	contribution of our analysis is that, in addition to examining how variation in demographic
3	and infection rates between slow- and fast-living species affect disease dynamics (e.g., De
4	Leo and Dobson, 1996; Bolzoni et al., 2008; Han et al., 2015), we also directly compare
5	how structural assumptions regarding the location of density-dependence affect the
6	negative impacts of disease on slow- and fast-living species.
7	Consider a host population with some density of susceptible hosts A_S and infected
8	hosts A_I . Assume that hosts become infected through density-dependent transmission,
9	where increasing host density increases host contact rate (Anderson and May, 1981;
0	McCallum, 2001; see Fig. S2 for frequency-dependent transmission). Also assume that
1	infected hosts suffer disease-induced mortality at some rate α (yr ⁻¹) and infected hosts can
2	recover at some rate γ (yr ⁻¹). We model these processes as

$$4 \qquad \frac{\frac{dA_S}{dt}}{\frac{dA_I}{dt}} = f(A_S + A_I)[A_S + A_I] - g(A_S + A_I)A_S - \beta A_I A_S + \gamma A_I$$

$$(1)$$

where βA_I is the force of infection (yr⁻¹). The function $f(A_S + A_I)$ defines the per capita host reproductive rate as a function of total host population density $A_S + A_I$. Consistent

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318	with the demographic buffering hypothesis, we assume that the per capita reproductive rate
319	of fast-living species is canalized and density-independent such that $f(A_S + A_I) = a$. In
320	contrast, the per capita reproductive rate of slow-living species is predicted to be less
321	canalized and to exhibit density-dependence (e.g., McDonald et al., 2016) and we assume
322	that it takes the form $f(A_S + A_I) = a - s[A_S + A_I]$ (Gurney and Nisbet, 1998), where s is
323	the strength of density-dependence. The function $g(A_S + A_I)$ defines the per capita
324	mortality rate of a host as a function of host density. For fast-living species, the per capita
325	mortality rate can vary with host density and we consider the form $g(A_S + A_I) = \mu + s[A_S$
326	+ A_I] (Anderson and May, 1981). The per capita mortality rates of slow-living species, on
327	the other hand, are predicted to be canalized (e.g., McDonald et al., 2016) such that per
328	capita mortality rate is density-independent, $g(A_S + A_I) = \mu$. In what follows, we refer to
329	the 'fast model' as the model with density-dependence in per capita mortality and the 'slow
330	model' as the model with density-dependence in per capita reproductive rate.

In addition to where density-dependence operates, another distinguishing feature of 331 fast and slow-living species are the magnitudes of their per capita mortality and 332 333 reproductive rates in the absence of density-dependence. Slow-living species tend to be 334 long-lived (low μ) with low reproductive rates (low a), while fast-living species tend to be 335 short-lived (high μ) with high reproductive rates (high a) (Gaillard *et al.*, 2016). This wellknown life-history trade-off is reflected in the model in the host's fundamental recruitment 336 number, which under the assumptions of equation 1 is $R_{0,host} = \frac{a}{\mu} R_{0,host}$ defines the 337 expected number of new hosts produced by a host over its lifetime when density-dependent 338 processes are absent. A host can obtain the same reproductive number by trading-off 339 between a and μ . In this way, fast-living and slow-living species may have the same 340

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2 3 4	341	fundamental recruitment number, but using contrasting strategies (e.g., low a , low μ vs.
5 6	342	high a , high μ). Here we consider a slow-living species that lives on average ten years (
7 8 9	343	$\mu = 0.1 \text{ yr}^{-1}$) and a fast-living species that lives on average half a year ($\mu = 2 \text{ yr}^{-1}$).
10 11 12	344	When a parasite successfully invades it will depress the host population below its
13 14	345	parasite free equilibrium density, which is $A_{\text{parasite free}}^* = \frac{a-\mu}{s}$ for both the slow and fast
15 16 17	346	model. How does the amount of host depression depend on the life-history strategy of the
18 19	347	host? To answer this question, we varied both the disease-induced mortality rate α and the
20 21 22	348	host fundamental recruitment number $R_{0,host}$ for the slow and fast model and compared
22 23 24	349	how much host equilibrium density was depressed in the presence of the parasite relative to
25 26	350	the parasite free equilibrium. Following Anderson and May (1981), we defined population
27 28 20	351	depression as $1 - A_{\text{parasite present}}^* / A_{\text{parasite free}}^*$, where $A_{\text{parasite present}}^*$ is the total equilibrium
29 30 31	352	population density in the presence of the parasite. Zero indicates no population depression
32 33	353	from disease, and a value closer to one indicates a higher population depression.
34 35 36	354	To adequately compare population depression between the two life-history
37 38	355	strategies, we also need to consider R_0 of the parasite. This describes the expected number
39 40 41	356	of infected individuals produced over the lifetime of an average infected host in a fully
42 43	357	susceptible host population. The proportion of a host population infected by a parasite
44 45 46	358	increases with increasing R_0 (Keeling and Rohani, 2008) and will affect the percentage of
40 47 48	359	the host population that experiences disease-induced mortality. Parasite R_0 for the slow
49 50 51	360	model is $R_{0,\text{parasite,slow}} = \frac{A_{\text{parasite free}}^{*}\beta}{\alpha + \gamma + \mu}$ and for the fast model is $R_{0,\text{parasite,fast}} =$
52 53 54	361	$\frac{A_{\text{parasite free}}^{*}\beta}{\alpha + \gamma + \mu + sA_{\text{parasite free}}^{*}}$. When $R_{0,\text{parasite},\cdot} > 1$, the parasite can invade and transmission can be
55 56 57	362	sustained in a host population whose density is at $A_{\text{parasite free}}^*$. As the value of $A_{\text{parasite free}}^*$
58 59		17

2 3 4	363	will vary between the slow and fast model, so will parasite R_0 . To account for this, we
5 6 7	364	ensured that parasite R_0 was the same for the slow and fast model for any parameter set by
7 8 9	365	adjusting the transmission parameter β for the slow model once disease-induced mortality
10 11	366	α and $R_{0,host}$ had been chosen. Biologically, this means that we assumed that parasites of
12 13 14	367	slow-living species generally had a higher transmission rate than those of fast-living
15 16	368	species. This assumption is reasonable given that 1) slow-living species generally have a
17 18 19	369	lower population density and a larger body size than fast-living species (Han et al., 2015),
20 21	370	and 2) transmission rate β scales positively with body size under the assumption of density-
22 23	371	dependent transmission (De Leo and Dobson, 1996; but see Joseph et al., 2013).
24 25 26	372	Our analysis shows that, given the same parasite R_0 and host fundamental
27 28	373	recruitment number $R_{0,host}$, the parasite depressed the population of slow-living species
29 30 31	374	more than fast-living species (Fig 3a,b). For both life-history strategies, population
32 33	375	depression was maximized for intermediate levels of disease-induced mortality α and
34 35 36	376	tended to increase with increasing host fundamental recruitment number (Fig. 3a,b). The
37 38	377	unimodal relationship between disease-induced mortality α and population depression is an
39 40 41	378	inevitable consequence of the fact that population depression has to be zero when $\alpha = 0$
42 43	379	and when α gets high enough that the parasite can no longer persist. Increasing the host
44 45	380	fundamental recruitment number in our model, on the other hand, increases equilibrium
46 47 48	381	host density, which increases transmission efficiency and increases population depression.
49 50	382	However, when the host fundamental recruitment number gets high enough, host births can
51 52	383	eventually compensate for disease-induced mortality and population depression will
54 55 56	384	decrease (Anderson and May, 1981).

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There are two non-exclusive explanations for the comparatively stronger population depression in slow-living species. First, despite ensuring identical parasite R_0 values for the slow and fast model, differences in intrinsic mortality rate or reproductive rate between fast and slow-living species, for example, can directly affect equilibrium parasite prevalence. If the slow model had higher disease prevalence than the fast model, this could explain the increased population depression. In Fig. 3c,d we show that the opposite occurred in most cases, i.e., the equilibrium prevalence was generally higher for the fast model compared to the slow model, given comparable parameters. Second, the stronger depression for slow-living species could be due to either the location of density-dependence (i.e., host survival vs host reproduction) or the differences in mortality rate between the two life-history strategies. If we ignore the biological implausibility and set the mortality rate μ to be the same for the slow and fast model, the stark differences in population depression are largely removed (Fig. S1). This indicates that the differences in population depression between the slow and fast models are driven largely by differences in mortality rate between the two life-history strategies, and not by the location of density-dependence.

We can further understand this result by considering how a proportional change in mortality rate proportionally affects $R_{0,host}$ (i.e., the elasticity of $R_{0,host}$ with respect to μ). Specifically, we can write the elasticity as $\frac{\partial R_{0,\text{host}}}{R_{0,\text{host}}} = -T\partial\mu$, where $T = 1/\mu$ is the average lifespan of the host (Lebreton 2005). When hosts have a short lifespan (i.e., T is small), consistent with a fast life-history strategy, a small change in host death rate μ given by $\partial \mu$ will have a small proportional change on $R_{0,host}$. In contrast, when hosts have a long lifespan (i.e., T is large), consistent with a slow life-history strategy, a small change in host death rate μ will have a large proportional change on $R_{0,host}$. Because equation 1 assumes

2 3 4	408	that the parasite affects host population dynamics by modifying mortality from μ to $\mu + \alpha$	α
- 5 6	409	for infected hosts, the above elasticity analysis suggests that, for a slow- and a fast-living	5
7 8	410	species with the same values of $R_{0,host}$, the proportional impact of disease will be larger f	for
9 10 11	411	the slow-living species (small μ) than for the fast-living one (large μ). This result is	
12 13 14	412	unchanged for frequency-dependent transmission (Fig. S2).	
15 16	413	As a final note, this simple model only considers a host with a single life stage.	
17 18 10	414	When hosts have multiple life stages (e.g., juvenile and adult) that are differentially	
20 21	415	affected by a parasite, the location of density-dependence can interact in more complex	
22 23	416	ways with underlying host and parasite parameters determining the extent of population	
24 25	417	depression in fast and slow-living species. For example, in a slow-living species where	
26 27 28	418	juveniles are substantially less susceptible to infection than adults, disease-induced	
29 30	419	mortality in adults could lead to density-dependent increases in per capita reproductive	
31 32	420	rates and a proportional increase in juvenile population density in the presence of disease).
33 34 25	421	However, in a species with density-dependence in juvenile mortality, this type of	
35 36 37	422	compensation would be harder to obtain. These predicted patterns provide an interesting	
38 39	423	future direction to explore at the intersection of disease ecology and LHT.	
40 41 42 42	424		
43 44 45	425		
46 47	426	The role of life-history plasticity in active demographic compensation	
48 49 50	427	Disease-associated risk can induce plastic changes in life-history traits that can potentiall	ly
51 52	428	result in active demographic compensation. For example, in the Tasmanian devil	
53 54 55 56	429	(Sarcophilus harrisii), females from populations decimated by a transmissible tumour	
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430	decreased their age at first reproduction and produced more offspring, a response that
431	partially offset the long-term impact of the disease and allowed persistence of Tasmanian
432	devil populations (Jones et al., 2008; Lachish et al., 2009; Lazenby et al., 2018). Although
433	a reduced population density due to infectious disease could potentially induce plasticity in
434	life-history traits, several empirical studies in vertebrates indicate that cues pertaining to a
435	parasite (e.g., antigens) or infected conspecifics are enough to trigger plasticity in life-
436	history traits (Wedekind, 2002; Bonneaud et al., 2004; Velando et al., 2006; Hanssen,
437	2006; Pompini et al., 2013; Sköld-Chiriac et al., 2019). We argue that this evidence
438	reinforces the traditional idea of the existence of a density-independent plastic response of
439	hosts to the increased risk of death or reduced fecundity associated with a parasitic
440	infection (Stearns, 1989b).
441	In animals, empirical demonstrations of parasite-induced plasticity in life-history
442	traits were traditionally restricted to invertebrates (e.g., Michalakis and Hochberg, 1994;
443	Agnew et al., 2000), but evidence is accumulating that this occurs across all vertebrate
444	classes as well (Fig. 2b-f; Table S1). Despite being well described at the individual level, it
445	is unclear how parasite-induced plasticity of life-history traits affects host demography and

446 **long-term host-parasite dynamics in vertebrates or other taxa**. Like the results in

447 invertebrates (Agnew *et al.*, 2000), examples from wild vertebrates support theoretical

expectations that the most common type of parasite-induced plasticity in life-history are

449 associated with reproduction. First, LHT predicts that if parasitism reduces an individual's

- 450 residual reproductive value (which is a measure of future reproductive opportunities) due to
 - 451 reduced fecundity, reduced survival, or chronic disease, selective benefits should exist for
- 452 individuals that can divert resources from self-maintenance to increase their current

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453	reproductive effort, in a "terminal investment" strategy to maximize fitness (Minchella,
454	1985; Forbes, 1993; Sorci et al., 1996; Hanssen, 2006; Schwanz, 2008). Second, for
455	individuals that have not reached sexual maturity, reducing the age of first reproduction
456	(i.e., diverting resources from growth to reproduction) should also enhance host fitness
457	since the chances of successful reproduction before either death or sterility are increased
458	(Stearns, 1989b; Hochberg et al., 1992; Michalakis and Hochberg, 1994). Also, in
459	vertebrates with complex life cycles, parasites can induce changes in the timing of life-
460	history transitions and niche shifts (e.g., hatching time in fish and amphibians) that permit
461	hosts to escape stage-specific parasitic infection (Warkentin et al., 2001; Wedekind, 2002;
462	Pompini <i>et al.</i> , 2013).
463	It is worth noting that infected hosts do not always exhibit increased reproductive
464	effort (Duffield et al., 2017). First, the direct negative effects of infectious disease can
465	inhibit reproduction (Richner, 1998). Second, if the prospects of future reproduction are not
466	diminished (e.g., CDV in spotted hyenas; Benhaiem et al., 2018), it would be more

467 efficient to reallocate resources to immune defences rather than to reproduction (but see Perrin et al., 1996). Third, the activation of the immune system is costly, and to sustain an 468 immune response, hosts may need to reallocate resources away from reproduction (Ilmonen 469 470 et al., 2000). Lastly, even if an infectious disease reduces a host's residual reproductive 471 value, investing in immunity could pay off. For instance, using a theoretical evolutionary model of a sexually-transmitted disease producing sterility in females, Johns et al. (2019) 472 473 showed that, even though terminal investment evolved under most scenarios, when immunity was highly cost-effective in delaying sterility, infected females increased 474 immune response at the expense of reproductive effort. 475

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It is unclear how plasticity in host life-history traits covaries with the position of a 476 species on the slow-fast continuum (but see Fig. 1), highlighting the urgent need for 477 theoretical and empirical development of this subject. In a general context, empirical 478 evidence shows that, in contrast to mammals with an intermediate to fast life-history 479 strategy (e.g., Tasmanian devils, wild boars), slow-living mammals are not able to bring 480 forward the onset of reproduction as a mechanism to compensate for an extrinsic cause of 481 increased mortality rate (see Servanty et al. (2011) and references therein). 482

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SECTION 5: HOST LIFE-HISTORY AND EVOLUTIONARY RESPONSES TO 485 **INFECTIOUS DISEASES** 486

Parasites could also drive rapid evolutionary changes in host life-history traits (Stearns et 487 al., 2000; Koella and Restif, 2001; but see Steiner and Tuljapurkar, 2012). Indeed, parasites 488 are ubiquitous in nature and exert selective pressures influencing the evolution of host-life 489 history strategies and maintaining plasticity of host life-history traits (Hochberg et al., 490 1992; Koella and Restif, 2001). This rapid evolution of host life-history traits could lead to 491 evolutionary rescue, i.e., when adaptive evolutionary change halt population decline and 492 prevents extinction (Carlson et al., 2014). Yet, we are unaware of any empirical 493 demonstration of rapid evolutionary change of life-history traits (e.g., fecundity, age at first 494 reproduction) in response to parasitism in vertebrate hosts (but see below for examples of 495 rapid evolution of tolerance/resistance mechanisms). The distinction between rapid 496 evolution of life-history traits and active demographic compensation in response to 497

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2 3 4	498	infectious disease is of practical relevance because evolutionary responses are expected to
5 6	499	be more hard-wired and slower to reverse than density-dependent and plastic responses,
7 8	500	and could alter population dynamics and resilience to other stressors (e.g., extreme climatic
9 10 11	501	events) even after the parasite has disappeared from the host population.
12 13 14	502	Additionally, rapid adaptive evolutionary changes of resistance or tolerance
15 16	503	mechanisms can have important effects on host-parasite dynamics (Duffy and Sivars-
17 18	504	Becker, 2007), allowing evolutionary rescue in susceptible hosts (e.g., Gignoux-Wolfsohn
19 20 21	505	et al., 2019). There are several examples of rapid evolution of resistance or tolerance
22 23	506	mechanisms in wild vertebrates, including amphibians (Savage and Zamudio, 2016), birds
24 25	507	(Bonneaud et al., 2011), and mammals (Epstein et al., 2016; Gignoux-Wolfsohn et al.,
26 27	508	2019). This rapid evolution is more likely to arise if the genetic variants involved in the
28 29 20	509	response to infectious disease are from pre-existing genetic variation rather than from the
30 31 32	510	recruitment of <i>de novo</i> mutations (Barrett and Schluter, 2008; Bonneaud <i>et al.</i> , 2011;
33 34	511	Hedrick, 2013). For example, populations of little brown bats (Myotis lucifugus)
35 36	512	experienced a dramatic and rapid decline due to white-nose syndrome (one monitored
37 38 39	513	colony declined by 98% between 2009-2015) but then started to recover slowly. Gignoux-
40 41	514	Wolfsohn et al. (2019) reported that this recovery was associated with rapid evolution that
42 43	515	occurred as a soft selection at multiple loci in genes linked to hibernation behaviour. These
44 45 46	516	authors concluded that this occurred from standing genetic variation because the short
47 48	517	timescale of fungal infection, mortality, and recovery processes makes selection of novel
49 50 51	518	mutations very unlikely (Gignoux-Wolfsohn et al., 2019).
52 53	519	Our current knowledge about the genetic architecture of resistance and tolerance
54 55 56	520	and the central role of standing genetic variation in the evolvability of host responses to

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521	parasitic infection leads to two important predictions that remain to be empirically tested in
522	vertebrate hosts. First, at the intraspecific level, resistance and tolerance are less likely to
523	evolve rapidly in small, isolated populations where small effective population size (N_e)
524	increases the risk of resistance/tolerance allele loss due to genetic drift and decreases
525	selection effectiveness in response to infectious disease (Eimes et al., 2011). This
526	prediction is supported by information from taxa other than vertebrates. For instance, in
527	plants, individuals from highly connected populations can exhibit higher levels of disease
528	resistance making those populations less susceptible to parasite-driven extinction (Jousimo
529	et al., 2014; Carlsson-Granér and Thrall, 2015). Second, at the interspecific level,
530	covariation between life-history and species' genetic characteristics likely determines the
531	speed of the evolution of parasite resistance and tolerance. Small-sized species with fast life
532	histories usually have higher genetic diversity than large, slow-living species (Wooten and
533	Smith, 1985; McCusker and Bentzen, 2010; Eo et al., 2011). In addition, because of their
534	short generation times, species at the fast end of the life-history continuum evolve at a
535	faster rate than those at the slow end (Bromham, 2011) and have higher non-synonymous to
536	synonymous substitution rate ratios (reflecting selection efficiency due to large N_e) (Figuet
537	et al., 2016). These two predictions suggest that fast-living species should benefit from a
538	higher capacity for rapid evolution than slow-living species in response to infectious
539	disease emergence. In contrast, Bruns et al. (2015) provided theoretical evidence that long-
540	lived hosts can evolve resistance more rapidly than short-lived hosts when the likelihood of
541	exposure to parasites and, therefore, the strength of selection for resistance, increases with
542	longevity. Testing these theoretical expectations is a major challenge but would allow us to

better predict the susceptibility of species and populations to the emergence of infectiousdiseases.

In addition to evolution, rapid changes in host life-history traits or resistance/tolerance mechanisms can be attributed to parasite-induced epigenetic variation (Gómez-Díaz et al., 2012). To date, the role of epigenetic mechanisms on host responses to parasitism remains poorly understood. Available evidence shows that parasite-induced changes in DNA methylation (an epigenetic mechanism) can occur within the sequence of protein-coding genes involved in host immunity and can also affect genes regulating a broad range of molecular and intracellular processes (Zhang et al., 2016; Sagonas et al., 2020). Methylation is a strong predictor of lifespan and aging (Lowe *et al.*, 2018; Anastasiadi and Piferrer, 2020) and partially regulates fertility in vertebrates (e.g., Woods et al., 2018). Therefore, parasite-induced methylation changes could produce aberrant DNA expression, which might alter individual phenome, eventually influencing compensatory responses. Depending on the extent of the germline reprogramming, epigenetic marks driven by parasite infection could be retained and could be transmitted from one generation to the next, allowing the transgenerational inheritance of resistance/tolerance mechanisms or life-history traits in some organisms (Greer *et al.*, 2011; Bošković and Rando, 2018).

562 SECTION 6: FROM POPULATIONS TO COMMUNITIES: INTEGRATING HOST 563 LIFE-HISTORY, COMMUNITY ASSEMBLY, AND INFECTIOUS DISEASE

Life-history traits can be further considered at the scale of ecological communities. The position of a species along the slow-fast life-history continuum can covary with both their Page 27 of 52

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- 3 4	566	epidemiological potential (i.e., host competence, which is defined as the capacity of a host
5 6	567	to maintain and transmit a parasite) and their position within communities (i.e., assembly
7 8	568	order). Thus, LHT offers an intriguing opportunity to more mechanistically link
9 10 11	569	epidemiological and ecological frameworks in the study of disease, particularly for multi-
12 13 14	570	host parasites.
15 16	571	One arena in which this topic has begun to receive more attention is in the study of
17 18 10	572	how changes in community diversity influence parasite transmission and disease risk.
20 21	573	Debate over whether biodiversity losses should consistently lead to higher disease risk
22 23	574	(e.g., the dilution effect) has prompted efforts to understand transmission within complex
24 25	575	multi-host communities from a more mechanistic perspective (e.g., Ostfeld and Keesing,
26 27 28	576	2012). When life-history traits covary with aspects such colonization ability, competitive
29 30	577	dominance, or extinction risk, species composition may be predictable along gradients of
31 32	578	species richness, disturbance regime, productivity, or community age. In amphibian
33 34 35	579	communities, for instance, Johnson et al. (2013) reported up to a 78% decrease in
36 37	580	trematode parasite transmission with an increase in amphibian host diversity. This result
38 39	581	was due to the non-random assembly of host communities: fast-living species with high
40 41 42	582	colonization abilities tended to be the most competent hosts for the trematode. Because
42 43 44	583	these species predominated in low-richness communities, parasite transmission tended to
45 46	584	decline with community diversity. As these species were increasingly accompanied or
47 48	585	replaced by lower-competence hosts at higher levels of species richness, the overall
49 50 51	586	infection competence of the community decreased. If communities were instead assembled
52 53	587	at random with respect to species composition (in laboratory experiments), there was no
54 55 56 57	588	such relationship between species richness and parasite transmission (Johnson et al., 2019;

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3 4	589	but see Becker et al., 2014). This highlights the importance of host life-history
5 6	590	characteristics in affecting both interspecific variation in host infection competence as well
7 8	591	as patterns of realistic assembly in ecological communities, which together could be used to
9 10 11	592	more broadly consider landscape-level transmission dynamics.
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18 19 20	595	SECTION 7: FROM THEORY TO PRACTICE: HOST LIFE-HISTORY AND
21 22 23	596	MITIGATION OF INFECTIOUS DISEASES IN WILDLIFE
24 25	597	We argue that a better understanding of the relationship between host life-history and
26 27	598	disease dynamics can improve the accuracy of disease risk analysis and inform mitigation
28 29 30	599	efforts at different stages of parasite invasion (sensu Langwig et al., 2015).
31 32 33	600	Disease risk analysis focuses on characterizing the potential disease hazards to an
34 35	601	animal, a population, or a species prior to their occurrence (Sainsbury and Vaughan-
36 37	602	Higgins, 2012; Jakob-Hoff et al., 2014). Risk largely depends on the adaptive capacity of
38 39 40	603	the host population/species, which we define as its capacity to cope with, or respond to, an
41 42	604	infectious disease. Because life-history permeates all components of host adaptive capacity
43 44	605	(Jakob-Hoff et al., 2014), life-history traits could be used to identify species at greater risk
45 46 47	606	and prioritize surveillance efforts (Grogan et al., 2014).
48 49	607	During the epizootic phase of parasite invasion, a rapid assessment of life-history
50 51 52	608	traits can help to identify those host species at greater risk and guide resource allocation
53 54	609	accordingly. For example, an initial, coarse assessment might prioritize naïve slow-living
55 56	610	species, populations of which are more likely to be impacted more severely by infectious
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611	disease (Fig. 3a,b). Also, given the greater capacity for an adaptive immune response, LHT
612	suggests that vaccine development would be more effective for slow-living species. LHT
613	insights can help refine initial assessments. For example, mitigation decisions in fast or
614	slow-living species might change depending on whether juveniles, adults or both life stages
615	are affected. Parasite-driven adult mortality will have a greater impact on the population
616	dynamics of slow-living than fast-living species, and high rates of parasite-driven juvenile
617	mortality can limit the efficacy of compensatory recruitment in slow-living species
618	(Valenzuela-Sánchez et al., 2017).
619	After the epizootic phase, fast-living host species might be managed by facilitating
620	host-parasite co-existence by reducing non-disease stressors to indirectly reduce additive
621	mortality (Scheele et al., 2019b). Conversely, slow-living species with slower recovery
622	might be managed more directly, by improving recruitment through habitat manipulation
623	(e.g., Haydon et al., 2002) or by population supplementation through the release of captive-
624	bred or translocated individuals (e.g., Gerber et al., 2019; Mendelson et al., 2019).
625	Importantly, while conservation plans intuitively seek to protect species at greater
626	risk of extinction, in a disease context the protection of one species will often require
627	managing additional species (Dobson, 2004). Life-history theory could help to predict the
628	potential and relative importance of other species to act as a disease reservoir (Han et al.,
629	2020), enabling the causative parasite to persist (Gog et al., 2002; Haydon et al., 2002;
630	Plourde et al., 2017). Empirical evidence shows that fast-living species commonly have a
631	higher host competence (Johnson et al., 2012; Joseph et al., 2013; Plourde et al., 2017;
632	Albery and Becker, 2020). This likely arises due to the lower investment of fast-living
633	species on immune defences, the adaptation of parasites to locally abundant hosts, or both

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634	(Joseph et al., 2013; Albery and Becker, 2020). Therefore, locally abundant fast-living
635	species could be targeted to protect a more vulnerable species at risk, pre-emptively or
636	reactively (Canessa et al., 2019a; Martel et al., 2020). A recent study presented theoretical
637	evidence that challenges the idea that fast-living species will invariably have a higher host
638	competence. Using age-structured, susceptible-infected models, Silk and Hodgson (2020)
639	showed that the demographic host competence (i.e., the ability of host populations to
640	sustain endemic prevalence) of slow-living species can be similar or even higher (especially
641	in the case of density-dependent parasite transmission) than that of fast-living species.
642	Disentangling how immune and nonimmune mechanisms (e.g., demography, behaviour,
643	density-dependence) of host competence interact at the population level seems to be a
644	critical step to better understand the relationship between host competence and life history
645	in multi-host parasite systems.

We foresee two major barriers to the use of LHT to inform wildlife disease 646 647 mitigation. First, relatively few proven, feasible options exist for disease control in wild 648 vertebrates (e.g., Garner et al., 2016). General trade bans for disease prevention (Shea et 649 al., 2014) or culling for outbreak control (Carter et al., 2009; Mysterud et al., 2018) are more likely to be focused on the potential of species to act as vectors of parasite entry than 650 651 on long-term disease dynamics (Pavlin et al., 2009). Actions deployed during the invasion 652 phase are likely to be broad-scope measures applied to a wide range of potential hosts and vectors within a landscape or ecological setting (e.g., culling; Gortazar et al., 2014). The 653 654 second barrier is the scarcity of life-history data for many threatened species (Conde et al., 2019). Understanding long-term demographic processes requires long-term data. Managers 655 can respond to such uncertainty by delaying actions until such knowledge is accumulated 656

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(but risking parasite spread during this period) (Grantham et al., 2009; Wintle et al., 2010), or by making decisions under uncertainty and assessing their effectiveness adaptively (e.g., Shea et al., 2014). Such assessments should be faster and more reliable for fast-living species with shorter generation times and larger cohorts, and hence larger sample sizes. Despite these limitations, disease risk assessments and mitigation plans generally are conducted with a limited knowledge of the system and depend on expert opinion and extrapolation (Canessa et al., 2019b). Therefore, we encourage scientists and practitioners to incorporate knowledge about broad LHT-disease relationships into expert assessments to narrow the decision space (Wintle et al., 2010), even in species where life-history data Per. might be limited.

SECTION 8: CONCLUSIONS AND FUTURE DIRECTIONS

Host life-history characteristics strongly influence host responses to parasitism at different levels of organization, from individuals to communities. While we highlight several empirical examples supporting LHT predictions about host responses to infectious disease in vertebrates, most theoretical expectations lack robust empirical validation. Addressing this challenge is critical for the advancement of theory and practice in infectious disease ecology. We have highlighted several mechanisms that allow host populations to compensate for an increased mortality or reduced fertility due to infectious disease, and how life-history can constrain these responses. While our capacity to disentangle these mechanisms in wild populations has been limited to date, new opportunities are arising to deal with this problem. These include the integration of experimental and observational

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)	approaches (e.g., Washburn et al., 1991; Rogowski et al., 2020) including through new
)	analytical tools, such as integrated population models, that incorporate multiple data types
L	and processes occurring at different levels of organization (e.g., McDonald et al., 2016;
2	Wilber et al., 2016). Although we focused this review on interspecific differences in host
3	life histories, the life-history traits of a species are not strictly static: within and among
1	population variation in life-history traits can depend on biotic or abiotic environmental
5	conditions. How the intraspecific variation in life-history traits influence host responses to
5	parasitism remains poorly understood, but it probably accounts for some of the
7	interpopulation variation in disease impacts that we observe in nature (e.g., Stephenson and
3	Cable, 2015). Accordingly, efforts to quantify trait distributions within communities which
)	capture both intraspecific and interspecific variation in key life-history traits are essential to
)	better understand the importance of host life-history on complex multi-host parasite
L	systems. LHT is a rich source of information that has not been fully applied to meeting the
2	challenges of wildlife disease mitigation. We suggest that applying information gleaned
3	from broad LHT-disease relationships (considering extrapolation in species where life-
1	history data might be limited or non-existent) can contribute significantly to disease risk
5	assessment and the identification of innovative mitigation strategies to address disease
5	threats to wildlife.

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15 16	707	
17 18 19 20	708	REFERENCES
21	700	
22	709	Agreen D. Kaalla I.C. and Michalakia V. (2000). Hast life history responses to
23	/10	Agnew, P., Koena, J.C. and Michalakis, Y. (2000). Host file history responses to
24	711	parasitism. Microbes and Infection 2, 891–896.
25	712	Albery G.F. and Becker, D.I. (2020) Fast-lived hosts and zoonotic risk. <i>Trands in</i>
26	712	Paragitalomy doi: 10.1016/i pt 2020/1 ast fived flosts and 200fford flsk. Trends in
27	/15	<i>T drusuology</i> , dol. 10.1010/j.pt.2020.10.012
28	714	Anastasiadi, D., and Piferrer, F. (2020). A clockwork fish: Age prediction using DNA
29	715	methylation-based biomarkers in the European seabass Molecular Ecology Resources
30	716	20 387 307
31	/10	20, 387-397.
32	717	Anderson R M and May R M (1979) Population biology of infectious diseases. Part I
33	718	Nature 280 361_367
34	/10	Nuture 200, 501-507.
35	719	Anderson, R.M. and May, R.M. (1981). The population dynamics of microparasites and
36	720	their invertebrate hosts <i>Philosophical Transactions of the Royal Society B</i> 291 451–
3/	721	
38	/21	524:
39	722	Barrett, L. G., Thrall, P. H., Burdon, J. J., and Linde, C. C. (2008). Life history determines
40	773	genetic structure and evolutionary potential of host-parasite interactions Trands in
41 42	725	genetic structure and evolutionary potential of nost parasite interactions. Trenas in
4Z 12	724	ecology and evolution 23, 678–685.
45 44	725	Barrett R D H and Schluter D (2008) Adaptation from standing genetic variation Trands
45	725	in Ecology and Evolution 22, 29, 44
46	720	in Ecology and Evolution 23, 38–44.
47	727	Becker C.G. Rodriguez D. Toledo L.F. Longo A.V. Lambertini C. Corrêa D.T.
48	728	Leite DS Haddad CEB and Zamudio K R (2014) Partitioning the net effect of host
49	720	diversity on an emerging emphibien nother on <i>Brocoodings of the Boyal Society B</i> :
50	729	Distant Science 281, 20141706
51	/30	Biological Sciences 281, 20141/96.
52	731	Benhaiem S. Marescot I., East M.L. Kramer-Schadt S. Gimenez O. Lebreton I-D
53	727	and Hofer, H. (2018). Slow recovery from a disease enidemic in the spotted hypera
54	732	land Hotel, H. (2018). Slow recovery nom a disease epidemic in the spotted hyena, a
55	/33	Reystone social carnivore. Communications Biology 1, 201.
56		
57		
58		33
59		
60		

1		
2 3 4 5	734 735	Bolzoni, L., De Leo, G. A., Gatto, M., and Dobson, A. P. (2008). Body-size scaling in an SEI model of wildlife diseases. <i>Theoretical population biology</i> 73, 374–382.
6 7 8 9	736 737 738	Bonneaud, C., Balenger, S.L., Russell, A.F., Zhang, J., Hill, G.E. and Edwards, S.V. (2011) Rapid evolution of disease resistance is accompanied by functional changes in gene expression in a wild bird. <i>Proceedings of the National Academy of Sciences</i> 108, 7866.
10 11 12 13	739 740 741	Bonneaud, C., Mazuc, J., Chastel, O., Westerdahl, H. and Sorci, G. (2004). Terminal investment induced by immune challenge and fitness traits associated with major histocompatibility complex in the house sparrow. <i>Evolution</i> 58, 2823–2830.
14 15 16	742 743	Bošković, A., and Rando, O. J. (2018). Transgenerational epigenetic inheritance. <i>Annual review of genetics</i> 52, 21–41.
17 18 19 20	744 745 746	Briggs, C.J., Knapp, R.A. and Vredenburg, V.T. (2010). Enzootic and epizootic dynamics of the chytrid fungal pathogen of amphibians. <i>Proceedings of the National Academy of Sciences</i> 107, 9695.
21 22 23 24 25	747 748 749	Brock, P. M., Murdock, C. C., and Martin, L. B. (2014). The history of ecoimmunology and its integration with disease ecology. <i>Integrative and Comparative Biology</i> 54, 353– 362.
26 27 28 29	750 751 752	Bromham, L. (2011). The genome as a life-history character: why rate of molecular evolution varies between mammal species. <i>Philosophical Transactions of the Royal Society B: Biological Sciences</i> 366, 2503–2513.
30 31 32	753 754	Bruns, E., Hood, M.E. and Antonovics, J. (2015). Rate of resistance evolution and polymorphism in long- and short-lived hosts. <i>Evolution</i> 69, 551–560.
33 34 35 36	755 756 757	Canessa, S., Bozzuto, C., Pasmans, F. and Martel, A. (2019a). Quantifying the burden of managing wildlife diseases in multiple host species. <i>Conservation Biology</i> 33, 1131–1140.
37 38 39 40 41 42	758 759 760 761 762	 Canessa, S., Spitzen-van der Sluijs, A., Stark, T., Allen, B.E., Bishop, P.J., Bletz, M., Briggs, C.J., Daversa, D.R., Gray, M.J., Griffiths, R.A., Harris, R.N., Harrison, X.A., Hoverman, J.T., Jervis, P., Muths, E., <i>et al.</i> (2019b). Conservation decisions under pressure: Lessons from an exercise in rapid response to wildlife disease. <i>Conservation Science and Practice</i> 2, e141.
43 44 45	763 764	Capdevila, P., Stott, I., Beger, M., and Salguero-Gómez, R. (2020). Towards a comparative framework of demographic resilience. <i>Trends in Ecology and Evolution</i> 35, 776–786.
46 47 48 40	765 766	Carlson, S.M., Cunningham, C.J. and Westley, P.A.H. (2014). Evolutionary rescue in a changing world. <i>Trends in Ecology and Evolution 29</i> , 521–530.
49 50 51 52	767 768	Carlsson-Granér, U. and Thrall, P.H. (2015) Host resistance and pathogen infectivity in host populations with varying connectivity. <i>Evolution</i> 69, 926–938.
53 54 55 56	769 770	Carter, S.P., Roy, S.S., Cowan, D.P., Massei, G., Smith, G.C., Ji, W., Rossi, S., Woodroffe, R., Wilson, G.J. and Delahay, R.J. (2009). Options for the control of disease 2: targeting
57 58 59 60		34

Ecology Letters

2		
3 4 5	771 772	hosts. In <i>Management of Disease in Wild Mammals</i> (eds R.J. Delahay, G.C. Smith and M.R. Hutchings), pp. 121–146. Springer Japan, Tokyo.
6	773	Cody, M.L. (1966). A general theory of clutch size. Evolution 20, 174-184.
7 8	774	Conde D A Staerk J Colchero F da Silva R Schölev J Baden H M Jouvet L Fa
9	775	LE Sved H Jongeians E Meiri S Gaillard J-M Chamberlain S Wilcken J
10	776	Iones $O R$ et al. (2019) Data gaps and opportunities for comparative and conservation
11 12	777	biology. Proceedings of the National Academy of Sciences 116, 9658–9664.
13	778	Daszak, P., Cunningham, A.A. and Hyatt, A.D. (2000). Emerging infectious diseases of
14	779	wildlife Threats to biodiversity and human health. Science 287, 443.
15		
10	780	Daszak, P., Cunningham, A.A. and Hyatt, A.D. (2001). Anthropogenic environmental
18	781	change and the emergence of infectious diseases in wildlife. <i>Acta Tropica</i> 78, 103–116.
19	782	De Leo, G. A., and Dobson, A. P. (1996). Allometry and simple epidemic models for
20	783	microparasites. Nature 379, 720–722.
21	704	Debson A (2004) Deputation dynamics of nothegons with multiple host spacing. The
23 24	784 785	American Naturalist 164, S64–S78.
25	786	Downs, C. J., Schoenle, L. A., Han, B. A., Harrison, J. F., and Martin, L. B. (2019). Scaling
26	787	of host competence Trends in parasitology 35 182–192
27	, 0,	or nost competence. Trends in parasitology 55, 162-192.
28	788	Duffield, K.R., Bowers, E.K., Sakaluk, S.K. and Sadd, B.M. (2017). A dynamic threshold
30	789	model for terminal investment. Behavioral ecology and sociobiology 71, 185.
31	790	Duffy, M.A. and Sivars-Becker, L. (2007). Rapid evolution and ecological host-parasite
32 33	791	dynamics: rapid evolution and disease dynamics. <i>Ecology Letters</i> 10, 44–53.
34		
35	/92	Eimes, J.A., Bollmer, J.L., Whittingham, L.A., Johnson, J.A., Van Oosterhout, C. and
36	793	Dunn, P.O. (2011). Rapid loss of MHC class II variation in a bottlenecked population is
37	794	explained by drift and loss of copy number variation. <i>Journal of Evolutionary Biology</i>
38	795	24, 1847–1856.
39	706	Eq. S.H. Doyle, I.M. and DeWoody, I.A. (2011). Genetic diversity in birds is associated
40	790	with body mass and babitat tyme. <i>Journal of Zoology</i> 282, 220, 226
41	/9/	with body mass and nabital type. <i>Journal of Zoology</i> 285, 220–226.
42 13	798	Epstein, B., Jones, M., Hamede, R., Hendricks, S., McCallum, H., Murchison, E.P.,
43 44	799	Schönfeld, B., Wiench, C., Hohenlohe, P. and Storfer, A. (2016), Rapid evolutionary
45	800	response to a transmissible cancer in Tasmanian devils <i>Nature Communications</i> 7
46	801	12684
47	001	12007.
48	802	Figuet, E., Nabholz, B., Bonneau, M., Mas Carrio, E., Nadachowska-Brzyska, K., Ellegren,
49	803	H. and Galtier, N. (2016). Life history traits, protein evolution, and the nearly neutral
50	804	theory in amniotes. Molecular Biology and Evolution 33, 1517–1527.
51		
52	805	Fisher, M.C., Henk, Daniel.A., Briggs, C.J., Brownstein, J.S., Madoff, L.C., McCraw, S.L.
53	806	and Gurr, S.J. (2012). Emerging fungal threats to animal, plant and ecosystem health.
54 55	807	<i>Nature</i> 484, 186–194.
56	000	Forbes M.P.I. (1003) Parasitism and host reproductive affort Oiles 67 111 150
57	000	For 0.000 , where (1333) , ratasiusin and nost reproductive effort. Othos $07, 444-430$.
58		35
59		
60		

3 4 5 6 7	809 810 811 812	Gaillard, J., Lemaître, J., Berger, V., Bonenfant, C., Devillard, S., Douhard, M., Gamelon, M., Plard, F. and Lebreton, JD. (2016). Life histories, axes of variation in. In <i>Encyclopedia of Evolutionary Biology</i> pp. 312–323. Kliman, R.M. Academic Press, Oxford.	
8 9 10 11	813 814 815	Garner, T.W.J., Schmidt, B.R., Martel, A., Pasmans, F., Muths, E., Cunningham, A.A., Weldon, C., Fisher, M.C. and Bosch, J. (2016). Mitigating amphibian chytridiomycosis in nature. <i>Philosophical Transactions of the Royal Society B</i> 371, 20160207.	
12 13 14 15	816 817 818	Gerber, B.D., Converse, S.J., Muths, E., Crockett, H.J., Mosher, B.A. and Bailey, L.L. (2018). Identifying species conservation strategies to reduce disease-associated declines <i>Conservation Letters</i> 11, e12393.	3.
16 17 18 19 20 21	819 820 821 822	Gervasi, S.S., Stephens, P.R., Hua, J., Searle, C.L., Xie, G.Y., Urbina, J., Olson, D.H., Bancroft, B.A., Weis, V., Hammond, J.I., Relyea, R.A. and Blaustein, A.R. (2017). Linking ecology and epidemiology to understand predictors of multi-host responses to an emerging pathogen, the amphibian chytrid fungus. <i>PLoS ONE</i> 12, e0167882.	
22 23 24 25	823 824 825	Gignoux-Wolfsohn, S.A., Pinsky, M.L., Kerwin, K., Herzog, C., Hall, M., Bennett, A.B., Fefferman, N.H. and Maslo, B. (2019). Genomic signatures of evolutionary rescue in bats surviving white-nose syndrome. <i>bioRxiv</i> , 470294.	
25 26 27 28 29	826 827 828	Gog, J., Woodroffe, R. and Swinton, J. (2002). Disease in endangered metapopulations: th importance of alternative hosts. <i>Proceedings of the Royal Society of London. Series B: Biological Sciences</i> 269, 671–676.	e
30 31 32 33	829 830 831	Gómez-Díaz, E., Jordà, M., Peinado, M. A., and Rivero, A. (2012). Epigenetics of host- pathogen interactions: the road ahead and the road behind. <i>PLoS Pathoghens</i> 8, e1003007.	
34 35 36 37	832 833 834	Gortazar, C., Diez-Delgado, I., Barasona, J.A., Vicente, J., De La Fuente, J. and Boadella, M. (2015). The wild side of disease control at the wildlife-livestock-human interface: A Review. <i>Frontiers in Veterinary Science</i> 1, 27.	L
38 39 40 41	835 836 837	Grantham, H.S., Wilson, K.A., Moilanen, A., Rebelo, T. and Possingham, H.P. (2009). Delaying conservation actions for improved knowledge: how long should we wait? <i>Ecology Letters</i> 12, 293–301.	
42 43 44 45 46	838 839 840	Greer, E. L., Maures, T. J., Ucar, D., Hauswirth, A. G., Mancini, E., Lim, J. P., and Brunet, A. (2011). Transgenerational epigenetic inheritance of longevity in <i>Caenorhabditis elegans</i> . <i>Nature</i> 479, 365–371.	
47 48 49 50	841 842 843	Grogan, L.F., Berger, L., Rose, K., Grillo, V., Cashins, S.D. and Skerratt, L.F. (2014). Surveillance for emerging biodiversity diseases of wildlife. <i>PLOS Pathogens</i> 10, e1004015.	
51 52 53 54 55	844 845	Gurney, W. and Nisbet, R.M. (1998). Ecological Dynamics. Oxford University Press, Oxford, United Kingdom.	
56 57 58 59		3	36

Ecology Letters

2		
3	846	Gutiérrez, J. S., Piersma, T., and Thieltges, D. W. (2019). Micro-and macroparasite species
4 5	847	richness in birds: The role of host life history and ecology. Journal of Animal Ecology
5	848	88, 1226–1239.
7		
8	849	Han, B. A., O'Regan, S. M., Schmidt, J. P., and Drake, J. M. (2020). Integrating data
9	850	mining and transmission theory in the ecology of infectious diseases. <i>Ecology Letters</i>
10	851	23, 1178–1188.
11	050	Han D. A. Dark A. W. Jollog, A. E. and Altizon S. (2015). Infactious disease
12	852	Han, B. A., Park, A. W., Jones, A. E., and Anizer, S. (2013). Infectious disease
14	853	transmission and benavioural allometry in wild mammals. <i>Journal of Animal Ecology</i>
15	854	84, 637–646.
16	855	Hanssen S A (2006) Cost of immune challenge and terminal investment in a long-lived
17	856	hird <i>Ecology</i> 87 2440–2446
18	050	ond. <i>Leology</i> 67, 2116 2116.
19	857	Haydon, D.T., Cleaveland, S., Taylor, L.H. and Laurenson, M.K. (2002). Identifying
20	858	reservoirs of infection: a conceptual and practical challenge. <i>Emerging infectious</i>
22	859	diseases 8, 1468–1473.
23	860	Healy K Ezard T H Jones O R Salguero-Gómez R and Ruckley V M (2010)
24	800	A nimal life history is shared by the page of life and the distribution of age specific
25	801	Annual the history is shaped by the pace of the and the distribution of age-specific
26 27	862	mortanty and reproduction. <i>Nature ecology and evolution</i> 3, 1217–1224.
27 28	863	Hedrick, P.W. (2013). Adaptive introgression in animals: examples and comparison to new
29	864	mutation and standing variation as sources of adaptive variation. <i>Molecular Ecology</i> 22,
30	865	4606–4618.
31		
32	866	Herrando-Pérez, S., Delean, S., Brook, B. W., and Bradshaw, C. J. (2012). Strength of
33 34	867	density feedback in census data increases from slow to fast life histories. <i>Ecology and</i>
35	868	<i>evolution</i> 2, 1922–1934.
36	860	Hettyey & Uiszegi I Herczeg D Holly D Vörös I Schmidt B R and Bosch I
37	805 970	(2020) Mitigating disease impacts in amphibian populations: capitalizing on the thermal
38	070	(2020). Witigating disease impacts in ampinoian populations, capitalizing on the meridian
39	8/1	opumum mismatch between a pathogen and its nost. Frontiers in Ecology and Evolution
40 41	872	7, 254.
42	873	Hilde, C. H., Gamelon, M., Sæther, B. E., Gaillard, J. M., Yoccoz, N. G., and Pélabon, C.
43	874	(2020). The demographic buffering hypothesis: evidence and challenges. <i>Trends in</i>
44	875	Ecology and Evolution 35, 523–538
45	0/0	
46 47	876	Hochberg, M.E., Michalakis, Y. and De Meeus, T. (1992). Parasitism as a constraint on the
47 48	877	rate of life-history evolution. Journal of Evolutionary Biology 5, 491–504.
49	070	Ilmonon D. Torko T. and Hassalquist D. (2000) Experimentally activated immuna
50	070 970	defense in female nied flyestehers results in reduced breeding success. <i>Proceedings of</i>
51	019	the Royal Society R 267, 665, 670
52	000	ine Royai Society D 201, 005–010.
53 54	881	Jakob-Hoff, R.M., MacDiarmid, S.C., Less, C., Miller, P.S., Travis, D. and Kock, R.
55	882	(2014). Manual of procedures for wildlife disease risk analysis. World Organisation for
56		
57		
58		37
59 60		
00		

1 2		
2 3 4 5	883 884	Animal Health. Published in association with the International Union for Conservation of Nature and the Species Survival Commission, Paris.
6 7 8	885 886	Jeschke, J. M., and Kokko, H. (2009). The roles of body size and phylogeny in fast and slow life histories. <i>Evolutionary Ecology 23</i> , 867–878.
9 10 11 12	887 888 889	Johns, S., Henshaw, J.M., Jennions, M.D. and Head, M.L. (2019). Males can evolve lower resistance to sexually transmitted infections to infect their mates and thereby increase their own fitness. <i>Evolutionary Ecology</i> 33, 149–172.
13 14 15 16 17	890 891 892 893	Johnson, P.T.J., Calhoun, D.M., Riepe, T., McDevitt-Galles, T. and Koprivnikar, J. (2019). Community disassembly and disease: realistic—but not randomized—biodiversity losses enhance parasite transmission. <i>Proceedings of the Royal Society B: Biological Sciences</i> 286, 20190260.
18 19 20 21	894 895 896	Johnson, P.T.J., Preston, D.L., Hoverman, J.T. and Richgels, K.L.D. (2013). Biodiversity decreases disease through predictable changes in host community competence. <i>Nature</i> 494, 230–233.
22 23 24 25 26	897 898 899	Johnson, P.T.J., Rohr, J.R., Hoverman, J.T., Kellermanns, E., Bowerman, J. and Lunde, K.B. (2012). Living fast and dying of infection: host life history drives interspecific variation in infection and disease risk. <i>Ecology Letters</i> 15, 235–242.
27 28 29 30	900 901 902	Jolles, A.E., Etienne, R.S. and Olff, H. (2006). Independent and competing disease risks: implications for host populations in variable environments. <i>The American Naturalist</i> 167, 745–757.
31 32 33 34 35	903 904 905 906	Jones, M.E., Cockburn, A., Hamede, R., Hawkins, C., Hesterman, H., Lachish, S., Mann, D., McCallum, H. and Pemberton, D. (2008). Life-history change in disease-ravaged Tasmanian devil populations. <i>Proceedings of the National Academy of Sciences</i> 105, 10023.
36 37 38 39	907 908 909	Joseph, M.B., Mihaljevic, J.R., Orlofske, S.A. and Paull, S.H. (2013). Does life history mediate changing disease risk when communities disassemble? <i>Ecology Letters</i> 16, 1405–1412.
40 41 42 43	910 911 912	Jousimo, J., Tack, A.M.K., Ovaskainen, O., Mononen, T., Susi, H., Tollenaere, C. and Laine, A-L. (2014). Ecological and evolutionary effects of fragmentation on infectious disease dynamics. <i>Science</i> 344, 1289.
44 45 46	913 914	Keeling, M. J., and Rohani, P. (2008). <i>Modeling infectious diseases in humans and animals</i> . Princeton University Press.
47 48 49	915 916	Keelling, M. and Rohani, P. (2008). Modeling Infectious Diseases in Humans and Animals. Princeton University Press, Princeton, New Jersey.
50 51 52 53	917 918 919	Kistner, E.J. and Belovsky, G.E. (2014). Host dynamics determine responses to disease: additive vs. compensatory mortality in a grasshopper–pathogen system. <i>Ecology</i> 95, 2579–2588.
54 55 56	920 921	Koella, J. C., and Restif, O. (2001). Coevolution of parasite virulence and host life history. <i>Ecology Letters 4</i> , 207–214.
57 58 59 60		38

2		
3 4 5 6	922 923 924	Lachish, S., McCallum, H. and Jones, M. (2009). Demography, disease and the devil: life- history changes in a disease-affected population of Tasmanian devils (<i>Sarcophilus</i> <i>harrisii</i>). Journal of Animal Ecology 78, 427–436.
7 8 9 10 11 12 13 14 15 16 17 18 19	925 926 927 928 929	 Langwig, K.E., Voyles, J., Wilber, M.Q., Frick, W.F., Murray, K.A., Bolker, B.M., Collins, J.P., Cheng, T.L., Fisher, M.C., Hoyt, J.R., Lindner, D.L., McCallum, H.I., Puschendorf, R., Rosenblum, E.B., Toothman, M., <i>et al.</i> (2015). Context-dependent conservation responses to emerging wildlife diseases. <i>Frontiers in Ecology and the Environment</i> 13, 195–202.
	930 931 932 933 934	Lazenby, B.T., Tobler, M.W., Brown, W.E., Hawkins, C.E., Hocking, G.J., Hume, F., Huxtable, S., Iles, P., Jones, M.E., Lawrence, C., Thalmann, S., Wise, P., Williams, H., Fox, S. and Pemberton, D. (2018). Density trends and demographic signals uncover the long-term impact of transmissible cancer in Tasmanian devils. <i>Journal of Applied Ecology</i> 55, 1368–1379.
20 21 22	935 936	Lebreton, JD. (2005). Dynamical and statistical modelss for exploited populations. Australian and New Zealand Journal of Statistics, 47, 49–63.
23 24 25	937 938	Lebreton, JD. (2006). Dynamical and statistical models of vertebrate population dynamics. <i>Comptes Rendus Biologies</i> 329, 804–812.
26 27 28	939 940	Lee, K.A. (2006). Linking immune defenses and life history at the levels of the individual and the species. <i>Integrative and Comparative Biology</i> 46, 1000–1015.
29 30 31 32	941 942 943	Lloyd-Smith, J. O., Cross, P. C., Briggs, C. J., Daugherty, M., Getz, W. M., Latto, J., and Swei, A. (2005). Should we expect population thresholds for wildlife disease?. <i>Trends in ecology and evolution 20</i> , 511–519.
33 34 35 36	944 945 946	Lowe, R., Barton, C., Jenkins, C. A., Ernst, C., Forman, O., Fernandez-Twinn, D. S., and Walter, L. (2018). Ageing-associated DNA methylation dynamics are a molecular readout of lifespan variation among mammalian species. <i>Genome Biology</i> 19, 1–8.
37 38 39 40 41	947 948 949	Martel, A., Vila-Escale, M., Fernández-Giberteau, D., Martinez-Silvestre, A., Canessa, S., Van Praet, S., and Picart, M. (2020). Integral chain management of wildlife diseases. <i>Conservation Letters</i> 13, e12707.
42 43 44	950 951	Martin II, L.B., Hasselquist, D. and Wikelski, M. (2006). Investment in immune defense is linked to pace of life in house sparrows. <i>Oecologia</i> 147, 565–575.
45 46 47	952 953	McCallum, H., Barlow, N. and Hone, J. (2001). How should parasite transmission be modelled? Trends in Ecology and Evolution, 16, 295–300.
48 49 50	954 955	McCusker, M.R. and Bentzen, P. (2010). Positive relationships between genetic diversity and abundance in fishes. <i>Molecular Ecology</i> 19, 4852–4862.
51 52 53 54 55	956 957 958	McDonald, J.L., Bailey, T., Delahay, R.J., McDonald, R.A., Smith, G.C. and Hodgson, D.J. (2016). Demographic buffering and compensatory recruitment promotes the persistence of disease in a wildlife population. <i>Ecology Letters</i> 19, 443–449.
56 57 58 59		39

2 3 4 5 6	959 960 961	Mendelson, J.R., Whitfield, S.M. and Sredl, M.J. (2019). A recovery engine strategy for amphibian conservation in the context of disease. <i>Biological Conservation</i> 236, 188–191.
0 7 8 9	962 963	Michalakis, Y. and Hochberg, M.E. (1994). Parasitic effects on host life-history traits: a review of recent studies. <i>Parasite</i> 1, 291–294.
10 11 12 13 14 15	964 965	Minchella, D.J. (1985). Host life-history variation in response to parasitism. <i>Parasitology</i> 90, 205–216.
	966 967	Mysterud, A. and Rolandsen, C.M. (2018). A reindeer cull to prevent chronic wasting disease in Europe. <i>Nature Ecology and Evolution</i> 2, 1343–1345.
16 17 18	968 969	Niel, C. and Lebreton, JD. (2005). Using demographic invariants to detect overharvested bird populations from incomplete data. <i>Conservation Biology</i> 19, 826–835.
19 20 21 22	970 971 972	Ohlberger, J., Langangen, Ø., Edeline, E., Claessen, D., Winfield, I. J., Stenseth, N. C., and Vøllestad, L. A. (2011). Stage-specific biomass overcompensation by juveniles in response to increased adult mortality in a wild fish population. <i>Ecology</i> 92, 2175–2182.
23 24 25	973 974	Ostfeld, R.S. and Keesing, F. (2012). Effects of host diversity on infectious disease. <i>Annual Review of Ecology, Evolution, and Systematics</i> 43, 157–182.
26 27 28	975 976	Pagán, I., Alonso-Blanco, C., García-Arenal, F. (2008). Host responses in life-history traits and tolerance to virus infection in <i>Arabidopsis thaliana</i> . <i>PLoS Pathogens</i> 4, e1000124.
29 30 31 32 33 34	977 978 979 980	Palacios, M. G., Sparkman, A. M., and Bronikowski, A. M. (2011). Developmental plasticity of immune defence in two life-history ecotypes of the garter snake, Thamnophis elegans–a common-environment experiment. <i>Journal of Animal Ecology</i> 80, 431–437.
35 36 37	981 982	Pavlin, B.I., Schloegel, L.M. and Daszak, P. (2009). Risk of importing zoonotic diseases through wildlife trade, United States. <i>Emerging infectious diseases</i> 15, 1721–1726.
38 39 40	983 984	Perrin, N., Christe, P. and Richner, H. (1996). On host life-history response to parasitism. <i>Oikos</i> 75, 317–320.
41 42 43 44 45 46 47	985 986 987 988 989 989	 Plourde, B.T., Burgess, T.L., Eskew, E.A., Roth, T.M., Stephenson, N. and Foley, J.E. (2017) Are disease reservoirs special? Taxonomic and life history characteristics. <i>PLOS ONE</i> 12, e0180716. Pompini, M., Clark, E.S. and Wedekind, C. (2013). Pathogen-induced hatching and population-specific life-history response to waterborne cues in brown trout (<i>Salmo trutta</i>). <i>Behavioral Ecology and Sociobiology</i> 67, 649–656.
48 49 50 51	991 992 993	 Previtali, M.A., Ostfeld, R.S., Keesing, F., Jolles, A.E., Hanselmann, R. and Martin, L.B. (2012). Relationship between pace of life and immune responses in wild rodents. <i>Oikos</i> 121, 1483–1492.
52 53 54 55 56 57	994 995	Ricklefs, R.E. and Wikelski, M. (2002). The physiology/life-history nexus. <i>Trends in Ecology and Evolution</i> 17, 462–468.
58		40

Page 41 of 52

Ecology Letters

1 2		
2 3 4 5	996 997	Richner, H. (1998). Host-parasite interactions and life-history evolution. <i>Zoology</i> 101, 333–344.
6 7 8 9 10	998 999 1000 1001	Rogowski, E. L., Van Alst, A. D., Travis, J., Reznick, D. N., Coulson, T., and Bassar, R. D. (2020). Novel parasite invasion leads to rapid demographic compensation and recovery in an experimental population of guppies. <i>Proceedings of the National Academy of Sciences 117</i> , 22580–22589.
11 12 13 14 15	1002 1003 1004	Sagonas, K., Meyer, B. S., Kaufmann, J., Lenz, T. L., Häsler, R., and Eizaguirre, C. (2020). Experimental parasite infection causes genome-wide changes in DNA methylation. <i>Molecular Biology and Evolution</i> 37, 2287–2299.
16 17 18	1005 1006	Sainsbury, A.W. and Vaughan-Higgins, R.J. (2012). Analyzing disease risks associated with translocations. <i>Conservation Biology</i> 26, 442–452.
19 20 21 22 23	1007 1008 1009	 Sandland, G. J., and Minchella, D. J. (2004). Life-history plasticity in hosts (Lymnaea elodes) exposed to differing resources and parasitism. <i>Canadian journal of zoology 82</i>, 1672–1677. Sauraga, A. F. and Zamudia, K. P. (2016). A deptive telegence to a pathogenia fungue drives.
24 25 26	1010 1011 1012	major histocompatibility complex evolution in natural amphibian populations. <i>Proceedings. Biological sciences</i> 283, 20153115–20153115.
27 28 29 30 31	1013 1014 1015 1016	Scheele, B.C., Pasmans, F., Skerratt, L.F., Berger, L., Martel, A., Beukema, W., Acevedo, A.A., Burrowes, P.A., Carvalho, T., Catenazzi, A., De la Riva, I., Fisher, M.C., Flechas, S.V., Foster, C.N., Frías-Álvarez, P., <i>et al.</i> (2019a). Amphibian fungal panzootic causes catastrophic and ongoing loss of biodiversity. <i>Science</i> 363, 1459.
32 33 34 35 36	1017 1018 1019	Scheele, B.C., Foster, C.N., Hunter, D.A., Lindenmayer, D.B., Schmidt, B.R. and Heard, G.W. (2019b). Living with the enemy: Facilitating amphibian coexistence with disease. <i>Biological Conservation</i> 236, 52–59.
37 38 39	1020 1021	Schröder, A., van Leeuwen, A., and Cameron, T. C. (2014). When less is more: positive population-level effects of mortality. <i>Trends in Ecology and Evolution 29</i> , 614–624.
40 41 42	1022 1023	Schwanz, L.E. (2008). Chronic parasitic infection alters reproductive output in deer mice. Behavioral Ecology and Sociobiology 62, 1351–1358.
43 44 45 46	1024 1025 1026	Sears, B.F., Snyder, P.W. and Rohr, J.R. (2015). Host life history and host-parasite syntopy predict behavioural resistance and tolerance of parasites. <i>Journal of Animal Ecology</i> 84, 625–636.
47 48 49 50	1027 1028 1029	Servanty, S., Gaillard, JM., Ronchi, F., Focardi, S., Baubet, É. and Gimenez, O. (2011). Influence of harvesting pressure on demographic tactics: implications for wildlife management. <i>Journal of Applied Ecology</i> 48, 835–843.
51 52 53 54 55 56	1030 1031 1032	Shea, K., Tildesley, M.J., Runge, M.C., Fonnesbeck, C.J. and Ferrari, M.J. (2014). Adaptive management and the value of information: learning via intervention in epidemiology. <i>PLOS Biology</i> 12, e1001970.
57 58 59 60		41

1 2		
3 4 5 6	1033 1034 1035	Silk, M.J., and Hodgson, D.J. (2020) Life history and population regulation shape demographic competence and influence the maintenance of endemic disease. <i>Nature</i> <i>Ecology and Evolution</i> , doi: 10.1038/s41559-020-01333-8
7 8 9 10	1036 1037 1038	Sköld-Chiriac, S., Nilsson, JÅ. and Hasselquist, D. (2019). Immune challenge induces terminal investment at an early breeding stage in female zebra finches. <i>Behavioral</i> <i>Ecology</i> 30, 166–171.
11 12 13	1039 1040	Sorci, G., Clobert, J. and Michalakis, Y. (1996). Cost of reproduction and cost of parasitism in the common lizard, <i>Lacerta vivipara</i> . <i>Oikos</i> 76, 121–130.
14 15	1041	Stearns, S.C. (1989a). Trade-offs in life-history evolution. Functional Ecology 3, 259–268.
16 17 18	1042 1043	Stearns, S.C. (1989b). The evolutionary significance of phenotypic plasticity. <i>BioScience</i> 39, 436–445.
19 20 21	1044 1045	Stearns, S. C. (2000). Life history evolution: successes, limitations, and prospects. <i>Naturwissenschaften</i> 87, 476–486.
22 23 24 25	1046 1047 1048	Steiner, U. K., and Tuljapurkar, S. (2012). Neutral theory for life histories and individual variability in fitness components. <i>Proceedings of the National Academy of Sciences 109</i> , 4684–4689.
26 27 28 29 30	1049 1050 1051	Stephenson, J. F., van Oosterhout, C., and Cable, J. (2015). Pace of life, predators and parasites: predator-induced life-history evolution in Trinidadian guppies predicts decrease in parasite tolerance. <i>Biology letters</i> 11, 20150806.
31 32	1052 1053	Tieleman, B.I. (2018). Understanding immune function as a pace of life trait requires environmental context. <i>Behavioral Ecology and Sociobiology</i> 72, 55.
33 34 35 36 37	1054 1055 1056	Tieleman, B.I., Williams, J.B., Ricklefs, R.E. and Klasing, K.C. (2005). Constitutive innate immunity is a component of the pace-of-life syndrome in tropical birds. <i>Proceedings of the Royal Society B</i> 272, 1715–1720.
38 39 40 41	1057 1058 1059	Tompkins, D.M., Dunn, A.M., Smith, M.J. and Telfer, S. (2011). Wildlife diseases: from individuals to ecosystems: ecology of wildlife diseases. <i>Journal of Animal Ecology</i> 80, 19–38.
42 43 44 45 46	1060 1061 1062 1063	Valenzuela-Sánchez, A., Schmidt, B.R., Uribe-Rivera, D.E., Costas, F., Cunningham, A.A. and Soto-Azat, C. (2017). Cryptic disease-induced mortality may cause host extinction in an apparently stable host–parasite system. <i>Proceedings of the Royal Society B</i> 284, 20171176.
47 48 49 50	1064 1065 1066	Velando, A., Drummond, H. and Torres, R. (2006). Senescent birds redouble reproductive effort when ill: confirmation of the terminal investment hypothesis. <i>Proceedings of the Royal Society B</i> 273, 1443–1448.
51 52 53 54	1067 1068	Warkentin, K.M., Currie, C.R. and Rehner, S.A. (2001). Egg-killing fungus induces early hatching of red-eyed treefrog eggs. <i>Ecology</i> 82, 2860–2869.
55 56 57 58 59 60		42

Ecology Letters

2		
3 4 5	1069 1070	Washburn, J. O., and Mercer, D. R., and Anderson, J. R. (1991). Regulatory role of parasites: impact on host population shifts with resource availability. <i>Science 253</i> , 185–188
6	10/1	188.
7 8 9	1072 1073	Wedekind, C. (2002). Induced hatching to avoid infectious egg disease in whitefish. <i>Current Biology</i> 12, 69–71.
10 11 12 13	1074 1075 1076	Wells, K., Hamede, R.K., Jones, M.E., Hohenlohe, P.A., Storfer, A. and McCallum, H.I. (2019). Individual and temporal variation in pathogen load predicts long-term impacts of an emerging infectious disease. <i>Ecology</i> 100, e02613.
14 15 16 17	1077 1078 1079	Wilber, M. Q., Langwig, K. E., Kilpatrick, A. M., McCallum, H. I., and Briggs, C. J. (2016). Integral projection models for host–parasite systems with an application to amphibian chytrid fungus. <i>Methods in ecology and evolution</i> 7, 1182–1194.
18 19 20	1080 1081	Wintle, B.A., Runge, M.C. and Bekessy, S.A. (2010). Allocating monitoring effort in the face of unknown unknowns. <i>Ecology Letters</i> 13, 1325–1337.
21 22 23 24 25 26	1082 1083 1084 1085	 Woodhams, D.C., Bell, S.C., Bigler, L., Caprioli, R.M., Chaurand, P., Lam, B.A., Reinert, L.K., Stalder, U., Vazquez, V.M., Schliep, K., Hertz, A. and Rollins-Smith, L.A. (2016). Life history linked to immune investment in developing amphibians. <i>Conservation Physiology</i> 4, cow025.
26 27 28 29 30	1086 1087 1088	 Woods III, L. C., Li, Y., Ding, Y., Liu, J., Reading, B. J., Fuller, S. A., and Song, J. (2018). DNA methylation profiles correlated to striped bass sperm fertility. <i>BMC genomics</i> 19, 244.
31 32 33 34 35 36 27	1089 1090 1091 1092 1093	 Wooten, M.C. and Smith, M.H. (1985). Large mammals are genetically less variable? <i>Evolution</i> 39, 210–212. Wright, J., Solbu, E. B., and Engen, S. (2020). Contrasting patterns of density-dependent selection at different life stages can create more than one fast–slow axis of life-history variation. <i>Ecology and evolution</i> 10, 3068–3078.
38 39 40 41 42	1094 1095 1096	Zhang, X., Justice, A. C., Hu, Y., Wang, Z., Zhao, H., Wang, G., and Xu, K. (2016). Epigenome-wide differential DNA methylation between HIV-infected and uninfected individuals. <i>Epigenetics</i> 11, 750–760.
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Figure 1

















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1 2 3 4	1097	Figure legends
5 6 7	1098	Figure 1 Host life-history characteristics affect the host-parasite interaction at different
, 8 9	1099	levels of organization, from individual-level susceptibility, to population-level responses to
10 11	1100	host community assembly. Life-history theory predicts different outcomes of parasitic
12 13 14	1101	infection across these organizational levels, which are related to the position of the host
15 16	1102	species (or populations) along the slow-fast continuum of life-history variation. Note that
17 18	1103	several of these predictions lack robust empirical validations, and some have not been
19 20 21	1104	tested at all (see main text). For example, we speculate that host species with intermediate
22 23	1105	life-history strategies should exhibit higher plasticity of life-history traits. This is because
24 25	1106	species on the fast end of the life-history continuum are expected to have low plasticity in
26 27 28	1107	fecundity parameters due to the high canalization of recruitment-related traits. Species on
29 30	1108	the slow end, in contrast, are expected to have a low canalization of recruitment-related
31 32	1109	traits. However, physical constraints imposed by their reproductive systems (e.g., large size
33 34 35	1110	of the offspring compared to the size of the uterus in slow-living mammals or large eggs
36 37	1111	compared to the size of the ovaries and oviducts in slow-living amphibians) and a
38 39	1112	potentially costly immune response against infection could limit the potential for active
40 41 42	1113	demographic compensation through increased reproductive effort. Importantly, fully
42 43 44	1114	understanding the effects of infectious disease relies on understanding a range of factors
45 46	1115	(e.g. parasite transmission, environmental effects) that depend on the system evaluated.
47 48 49 50	1116	
50 51 52	1117	Figure 2 Empirical examples of density-dependent compensation (a) and plasticity in life-
53 54	1118	history traits (b-f) in response to infectious disease in vertebrates. In (a) the correlation of
55 56 57	1119	adult survival and recruitment with disease prevalence and host population density in the
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1120	badger-Mycobacterium bovis system is shown. The dark grey and light grey lines in the top
1121	panels of this figure represent the survival of males and females, respectively. Density-
1122	dependent compensatory recruitment allowed the long-term persistence of this badger
1123	population with endemic <i>M. bovis</i> infection (adapted from McDonald <i>et al.</i> [2016]). (b)
1124	Troglodytes aedon females experimentally exposed to Salmonella enterica LPS increased
1125	the amount of yolk per unit of egg mass and food provisioning to nestlings (photo: Mylthon
1126	Jiménez-Castillo). (c) Sarcophilus harrisii females from populations with the transmissible
1127	Devil facial tumour disease have an earlier onset of reproduction and more pouch young
1128	(photo: Rodrigo Hamede). (d) Litoria verreauxii alpine individuals experimentally infected
1129	with Batrachochytrium dendrobatidis have more spermatic cell bundles and a larger
1130	proportion of spermatozoa bundles (males), and larger ovaries and oviducts (females)
1131	(photo: Matt West). (e) Zootoca vivipara females naturally infected with Hematozoa
1132	exhibited a larger relative clutch mass and higher maternal investment per young (photo:
1133	Matthieu Berroneau). (f) Coregonus sp. eggs exposed to water-borne cues from
1134	Pseudomonas fluorescens or conspecific infected eggs hatched earlier (photo: Paul Vecsei).
1135	References and study details (b-f) can be found in the Appendix S2 in Supporting
1136	Information.

1137 **Figure 3** Life-history strategies and host population depression due to parasite-induced 1138 mortality. (a,b) We used equation 1 to compute the equilibrium total host density when the 1139 parasite was absent and when the parasite was present and depressed the host population. 1140 We calculated population depression as $1 - A_{\text{parasite present}}^*/A_{\text{parasite free}}^*$, where zero 1141 indicates no population depression. We varied the host fundamental recruitment number 1142 $R_{0,\text{host}} = \frac{a}{\mu}$ between 2 and 6. To do this, we held intrinsic mortality μ fixed at 0.1 yr⁻¹ for

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the slow-living species and at 2 yr $^{-1}$ for the fast-living species and chose the reproductive rate $a \text{ yr}^{-1}$ to ensure the host fundamental recruitment numbers were the same between hosts. We varied disease-induced mortality rate α between 0 and 6 yr⁻¹. The transmission parameter β was fixed at 2 yr⁻¹ for fast-living species and varied for slow-living species such that $R_{0,\text{parasite,fast}} = R_{0,\text{parasite,slow}}$. The other parameters were s = 1 and $\gamma = 0.4$ yr⁻¹. We also varied the strength of density-dependence s between 0.1 and 1 and γ between 0.1 and 4 and the qualitative results were unaffected (not shown). The color indicates the magnitude of population depression, gray lines and numbers give specific contours of population depression, and the black line indicates where $R_{0,\text{parasite},.} = 1$, to the left of which the parasite could not invade the host population. (c,d) Same as (a,b), except parasite prevalence is plotted for the two life-history strategies. See the main text for details on parameter names.



Figure 1

215x180mm (300 x 300 DPI)



Figure 2

283x168mm (150 x 150 DPI)





Figure 3

842x718mm (96 x 96 DPI)