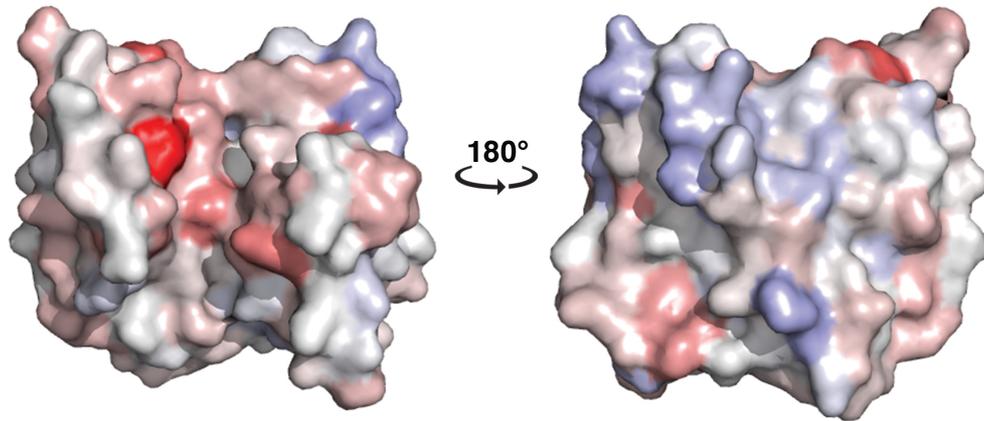


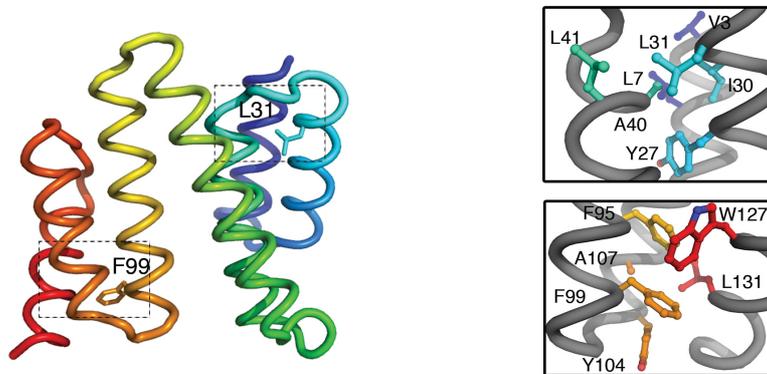
**THE CRYSTAL STRUCTURE OF THE SGT1-SKP1 COMPLEX: THE LINK BETWEEN
HSP90 AND BOTH SCF E3 UBIQUITIN LIGASES AND KINETOCHORES**

Oliver Willhoft, Richard Kerr, Dipali Patel, Wenjuan Zhang, Caezar Al-Jassar, Tina Daviter,
Stefan H. Millson, Konstantinos Thalassinou, Cara K. Vaughan

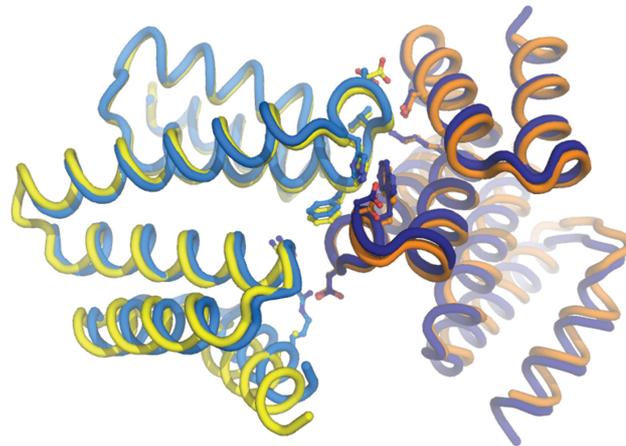
C



D



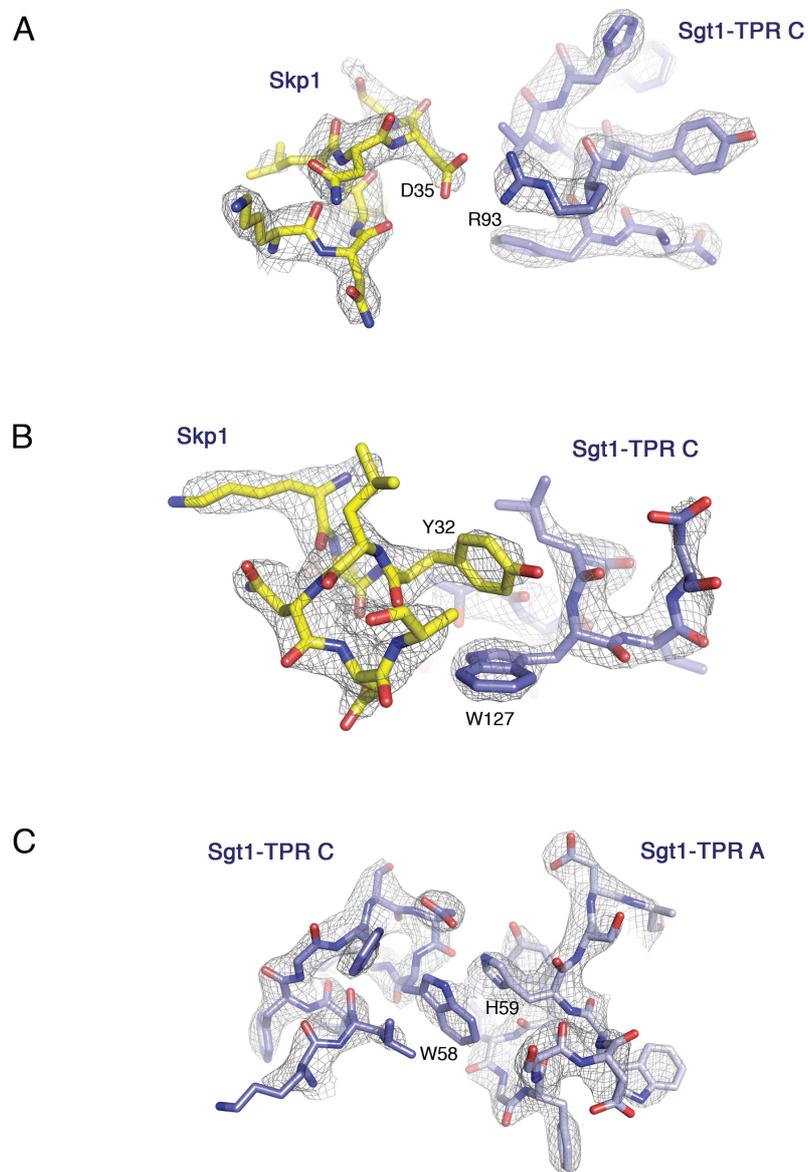
E



(A) RMSD fit for C α atoms of the Sgt1 TPR domain and TPR domains with high sequence identity. The PDB ID for PPP5, Sgt, CPTR3 and Hop TPR1 used for the analysis is shown, and each chain is colour-coded for clarity. (B) Sequence alignment of the TPR domain of homologues of Sgt1 from fungi, higher eukaryotes and plants. White text on a red background

indicates strict identity, red text indicates similarity in a group, blue frame indicates similarity across groups. Residues contributing to the oligomerization interface of the Sgt1 dimer, the Sgt1-Skp1 interaction interface, and both interfaces, are shown as blue, red and green triangles respectively. Secondary structure from the Sgt1 TPR domain is mapped to the primary structure above the alignment {Gouet:2003hk}. **(C)** Surface conservation of the TPR domain generated using a ClustalW alignment of representative eukaryotes and ProtSkin {Ritter:2004gw}. Strongly conserved:red to weakly conserved:blue. **(D)** The Sgt1TPR domain, coloured rainbow from N:blue to C:red. The mutations L31P and F99L are present in the TPR domain of Sgt1-3p. The inset boxes highlight the interactions made by these residues showing both to be within hydrophobic cores. The former mutation introduces a proline into helix 2 and will therefore disrupt the helix and consequently the structural core between helices 1, 2 and 3. The latter is a conservative, non-disruptive mutation. **(E)** The two Sgt1:Sgt1 interfaces in the asymmetric unit are identical. A superposition of the BA dimer (yellow/orange) on the AC (light blue/dark blue) dimer using all atoms is shown. The RMSD of this superposition is 1.46 Å. Residues involved in the interface are shown in ball and stick.

**SUPPLEMENTARY FIGURE S2: ELECTRON DENSITY MAPS AT THE SGT1-SKP1 AND
SGT1-SGT1 INTERFACES**

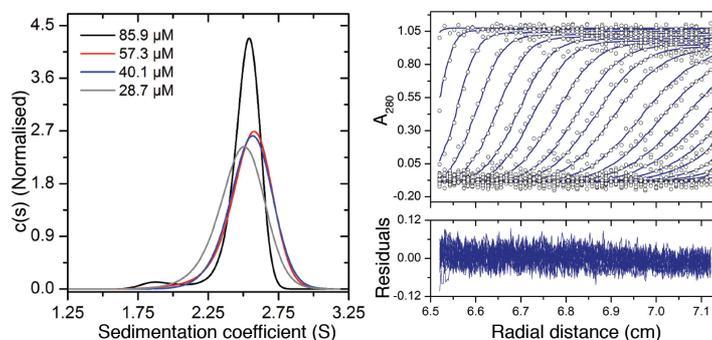


Simulated annealing composite omit maps calculated for the Sgt1-Skp1 interface at (A) Sgt1-R93 and (B) Sgt1-W127A and for the Sgt1 dimerisation interface at (C) H59.

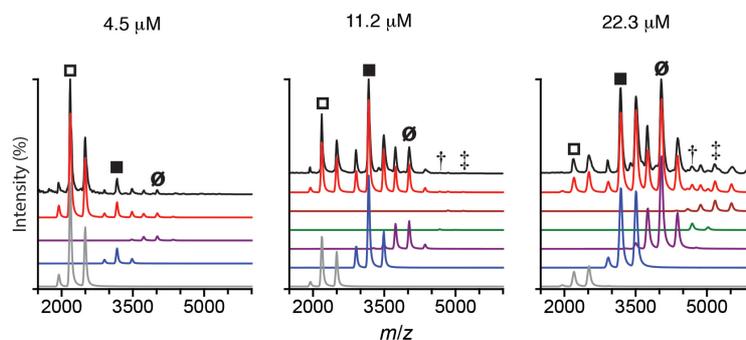
SUPPLEMENTARY FIGURE S3: CHARACTERISATION OF SGT1 TPR DOMAIN

OLIGOMERISATION

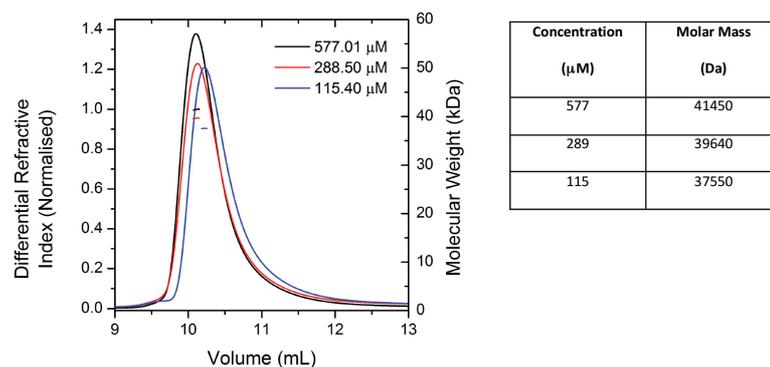
A



B



C



(A) $c(s)$ distributions for SGT1TPR at 28.7 μM , 40.1 μM , 57.3 μM & 85.9 μM monomer and corresponding boundary fits and residuals for the sample generated by SV-AUC.

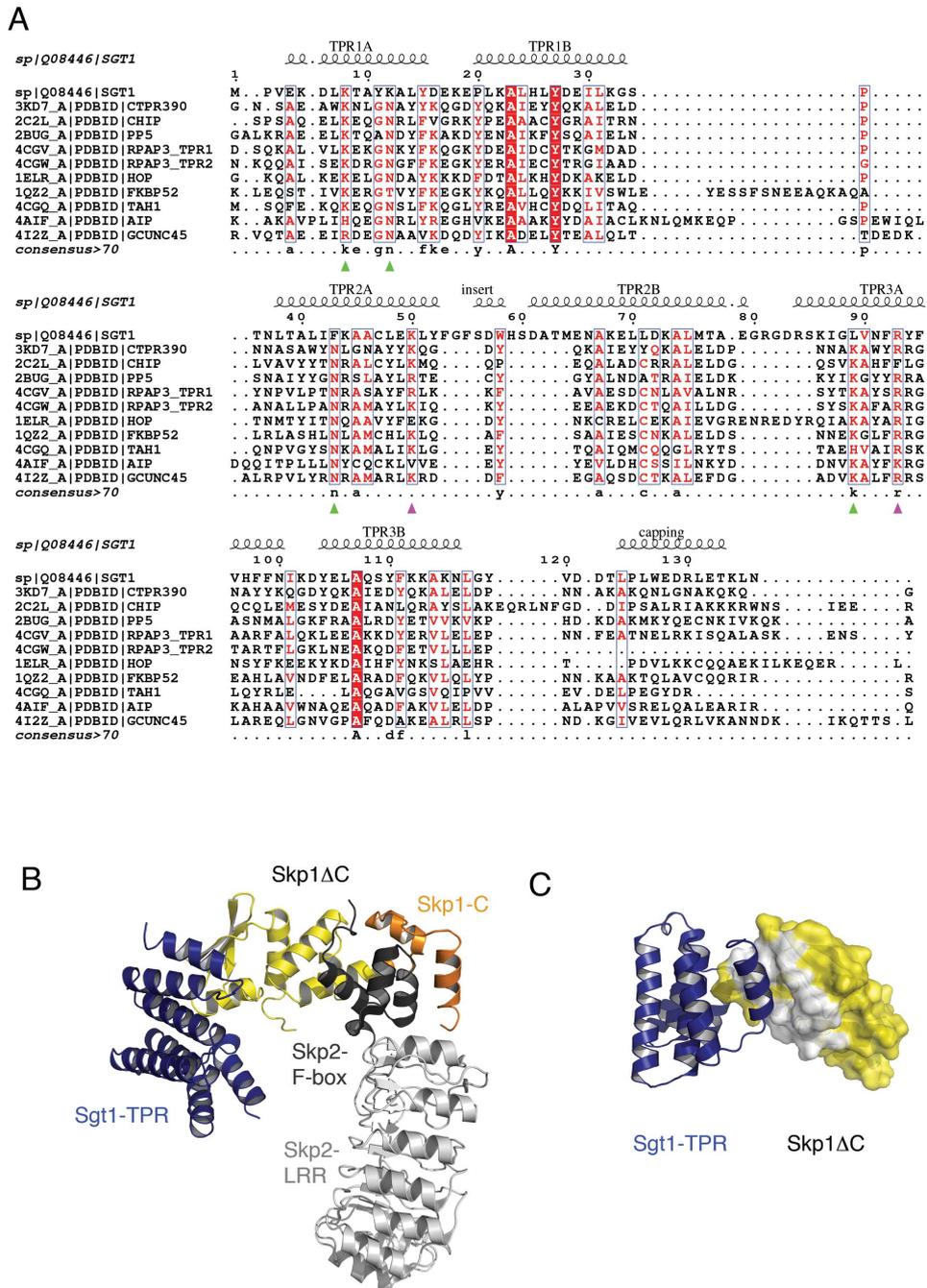
Supplementary Table S1 details the sedimentation coefficients, MW and f/f_0 .

(B) Native nESI-MS spectra of SGT1TPR at 4.5 μM , 11.2 μM and 22.3 μM . Charge state series corresponding to monomer, dimer, and trimer are indicated in grey, blue and purple

respectively. Charge states: wt +13 ○, (wt)₂ +20 ●, (wt)₃ +24 ◆. Spectra were deconvoluted to individual charge state series using Amphitrite {Sivalingam:2013vb} with the raw data shown in black and the sum of simulated spectra in red. Supplementary Table S2 details the percentage contribution of individual species at each concentration.

(C) Chromatograms and estimated molecular weights determined using a Superdex 75 HR10/30 (GE Healthcare) and multi-angle light scattering (SEC-MALS) for Sgt1TPR at 115 μM, 289 μM, 577 μM. An approximately 10-fold increase in sample concentration was used compared to the SV-AUC analysis to account for the dilution effect of size exclusion chromatography.

SUPPLEMENTARY FIGURE S4: SGT1-SKP1 IN THE CONTEXT OF TPR-MEEVD, SKP1-FBXL AND SKP1-CULLIN COMPLEXES



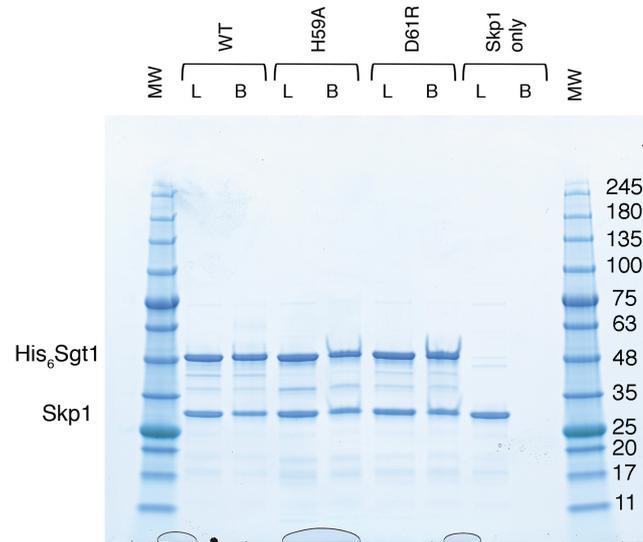
(A) A structure-based multiple sequence alignment using T-Coffee Expresso of the TPR domain of Sgt1 with TPR-cochaperones that interact with Hsp90 via its MEEVD motif with {DiTommaso:2011kx}. Cochaperones with deposited crystal structures with MEEVD interaction were selected for alignment and their PDB is labelled. Residues that contribute

conserved electrostatic interactions with the carboxylate-clamp are highlighted in green and with other residues of the MEEVD motif are highlighted in magenta.

(B) A least squares fit of the Sgt1TPR-Skp1BTB Δ crystal structure (blue and yellow respectively) with the Skp1-Skp2 heterodimer (PDB: 1FQV). The C-terminal region of Skp1 (not present in our construct) is coloured orange to distinguish it from our Skp1BTB Δ construct. The F-box and LRR domains of Skp2 are coloured different shades of grey. **(C)** The Sgt1TPR-Skp1BTB Δ crystal structure showing a surface representation of Skp1 (yellow). Residues of Skp1 that interact with Cul1 are white (from PDB 1LDK).

SUPPLEMENTARY FIGURE S5: ASSOCIATION OF SGT1 DIMERISATION MUTANTS

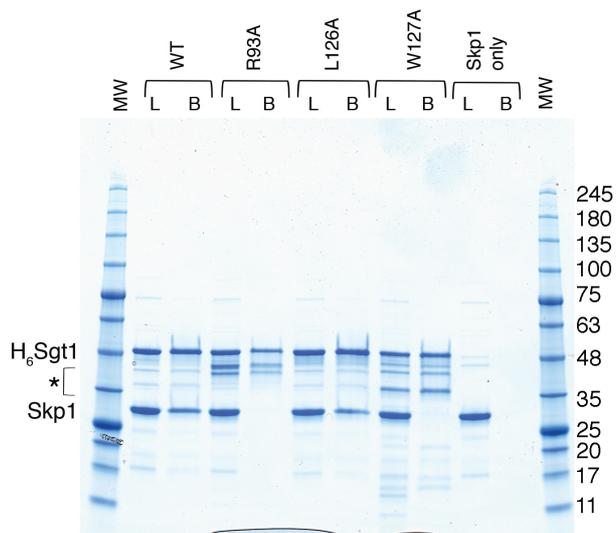
WITH SKP1



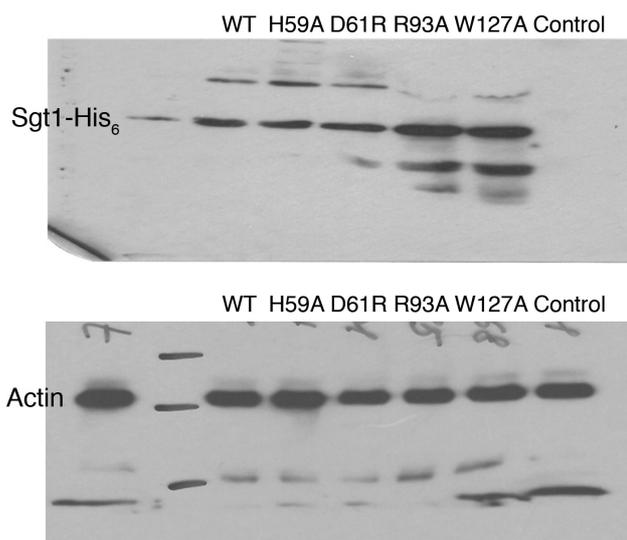
The ability of full-length Skp1 to interact with His₆-tagged full-length wild type (WT) Sgt1 and indicated mutants assessed by pull-down on Ni resin. L = 10% load, B = bound.

FIGURE S6: FULL GELS AND WESTERN BLOTS

A



B



(A) Pulldown of full-length Skp1 using His₆-tagged wild type Sgt1 (WT) and indicated mutants on Ni resin. L = 10% load, B = bound. * = Degradation products of Sgt1 that copurify with the full-length protein. (B) Analysis of Sgt1 levels in JJ345 *S. cerevisiae* cultures in SD-Trp media at 28 °C expressing WT Sgt1 and either WT or mutant Sgt1-His₆. Actin was measured as a loading control.

Supplementary Tables

SUPPLEMENTARY TABLE S1: Sedimentation Velocity Analytical Ultracentrifugation for wild-type, mutant and truncation constructs of Sgt1

Protein	Concentration (μM)	$s_w(20,w)$ (S)	Molar Mass (Da)	Frictional ratio f/f_o	% of total
WT	22.3	4.1	92158	1.78	97.6
	11.2	4.0	84097	1.70	97.6
	4.5	3.9	73891	1.60	96.1
His ₆ -WT	11.2	4.4	84029	1.55	92.0
His ₆ -H59A	11.2	2.5	41023	1.67	95.5
TPR	85.9	2.53	51031	1.797	96.5
	57.3	2.57	47597	1.733	97.6
	40.1	2.56	47644	1.752	96.8
	28.7	2.50	41116	1.583	95

SUPPLEMENTARY TABLE S2: Contribution of individual species to nESI-MS spectra of wild-type, mutant and truncation constructs of Sgt1

Protein	Concentration (μM)	Monomer (%)	Dimer (%)	Trimer (%)
WT	22.3	33.1 ± 5.7	48.3 ± 2.4	18.6 ± 3.4
	11.2	19.7 ± 5.0	61.1 ± 2.1	19.2 ± 5.6
	4.5	21.9 ± 1.1	69.4 ± 7.6	8.6 ± 7.5
His ₆ -H59A	11.2	71.4 ± 4.9	28.6 ± 4.9	-
His ₆ -D61R	11.2	22.1 ± 7.0	72.6 ± 6.0	5.3 ± 3.6
TPR	22.3	10.4 ± 1.8	33.8 ± 4.6	$33.7 \pm 6.5^*$
	11.2	25.7 ± 4.6	45.9 ± 2.1	26.8 ± 4.9
	4.5	80.0 ± 4.9	13.8 ± 2.1	6.3 ± 4.0

Errors are representative of 3 replicates for H59A and 4 replicates for WT and D61R.

* This concentration of Sgt1TPR also has contributions from tetramer ($7.3 \pm 4.1\%$) and pentamer ($10.1 \pm 4.6\%$).

SUPPLEMENTARY TABLE S3: Crystallography statistics

Data collection	SAD
Space Group	P 3₂ 2 1
Wavelength (Å)	0.979494
Unit cell <i>a, b, c</i> (Å)	94.6, 94.6, 123.0
Resolution (Å)	81.9 – 2.8 (2.9 - 2.8)
Total observations	339894
Total unique obs.	15826
R _{merge}	0.089 (0.826)
Completeness (%)	100.0 (100.0)
Anom. Comp. (%)	100.0 (99.9)
Multiplicity	21.5 (21.9)
Anomalous multiplicity	11.5 (11.4)
<i>I</i> / σ <i>I</i>	29.2 (4.9)
Refinement	
R _{work} /R _{Free}	20.10%/23.92%
Number of protein atoms	3707
r.m.s.d. bond lengths (Å)	0.010
r.m.s.d. bond angles (°)	0.980
Ramachandran plot	
Preferred region	98.37%
Allowed	1.63%
Outliers	0%