Using the Slug Mucosal Irritation assay to investigate the tolerability of tablet excipients on human skin in the context of the use of a nipple shield delivery system

Tolerability of tablet excipients on the nipple

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Purpose

Neonates are particularly challenging to treat. A novel patented drug delivery device containing a rapidly disintegrating tablet held within a modified nipple shield (NSDS) was designed to deliver medication to infants during breastfeeding. However, concerns exist around dermatological nipple tolerability with no pharmaceutical safety assessment guidance to study local tissue tolerance of the nipple and the areola.

This is the first Slug Mucosal Irritation (SMI) study to evaluate irritancy potential of GRAS excipients commonly used to manufacture rapidly disintegrating immediate release solid oral dosage form.

Methods

Zinc sulphate selected as the antidiarrheal model drug that reduces infant mortality, was blended with functional excipients at traditional levels [microcrystalline cellulose, sodium starch glycolate, croscarmellose sodium, magnesium stearate]. Slugs were exposed to blends slurried in human breast milk to assess their stinging, itching or burning potential, using objective values such as mucus production to categorize irritation potency.

Results

Presently an in vivo assay, previously validated for prediction of ocular and nasal irritation, was used as an alternative to vertebrate models to anticipate the potential maternal dermatological tolerability issues to NSDS tablet components. The excipients did not elicit irritancy. However, mild irritancy was observed when zinc sulphate was present in blends.

Conclusion

These promising good tolerability results support the continued investigation of these excipients within NSDS rapidly disintegrating tablet formulations. Topical local tolerance
effects being almost entirely limited to irritation, the slug assay potentially adds to the existing preformulation toolbox, and may sit in between the in vitro and existing in vivo assays.

250 words (limit 250)

KEY WORDS:
Nipple Shield Delivery System, Slug Mucosal Irritation Assay, Skin Tolerability, Tablet Excipients, Pediatric

ABBREVIATIONS
API: Active Pharmaceutical Ingredient
BAC: Benzalkonium Chloride
CP: Contact Period
GRAS: Generally Recognised as Safe
HIV: Human Immunodeficiency Virus
HLTV: Human T-lymphotropic Virus
HBM: Human Breast Milk
MP: Mucus Production
NC: Negative Control
NSDS: Nipple Shield Delivery System
PC: Positive Control
REC: Research Ethics Committee
PBS: Phosphate Buffered Saline
SIB: Stinging, Itching or Burning
SMI: Slug Mucosal Irritation
INTRODUCTION

Identifying the paediatric drug product technology gap

The development of age appropriate medicines which deliver an active pharmaceutical ingredient (API) to children at the required rate and extent is a complex process. Neonates are a particularly challenging sub-population to address for formulation scientists due to issues including dysphagia, taste aversion, and the need for frequent dose modifications. (1)

Liquid formulations have been typically the dosage forms of choice for paediatric drug administration, but are often not practical in developing countries because of high cost, lack of access to refrigeration, contamination issues and limited shelf life. (2 - 4) They may also be unpalatable and contain undesirable or unsafe preservatives and solvents. (5) Solid oral dosage forms for infants are often scaled down from adult doses, and there is currently a debate on the limitations of clinical work performed to demonstrate suitability of the dose to the infant. (6, 7) Dispersible tablets can also be used, but require clean sources of water for reconstitution. They also require administration devices for which the volume of reconstituted suspension is calculated based upon body weight, body surface area or age, depending on the therapeutic index of the drug. However there is currently a WHO recommendation if not push for solid oral dosage forms to maximise drug product stability, particularly in developing countries. (8)

A major contributor to this unacceptable statistic is diarrhoeal disease, the second leading cause of mortality in this age group, being responsible for 760,000 deaths yearly. (9) A significant proportion of the 1.7 billion annual cases of diarrhoeal disease reported globally could be prevented through ready access to clean drinking
water alongside better sanitation and hygiene, both of which also limit the utility of medicines designed to be administered post reconstitution in drinking water.

**A potential solution to the paediatric drug product technology gap in neonates**

As a child friendly administration vehicle, milk has gained legitimacy and research continues to demonstrate its multiple benefits as a potential solubilizing, gastroprotective and taste masking agent. (10, 11)

A novel “Nipple Shield Delivery System” (NSDS) has been proposed as a means to address some of these challenges, with preliminary proof-of-concept in vitro simulation studies and non-clinical user-acceptability studies conducted in the past five years. (12 - 16)

This thin disposable device (Figure 1), in one format, could be adapted from an existing nipple shield breastfeeding aid to contain a fast dispersible or rapidly disintegrating tablet, and placed over the mother's breast just before infant feeding (12). When the human breast milk (HBM) passes through the device it releases an API to the infant via the milk. Based on patient need, a wide-range of APIs could be delivered to infants using the NSDS, such as antibiotics, antivirals, antimalarials, vitamins, nutrients, and probiotics. The APIs, which are stored in a dry form prior to reconstitution in HBM during administration, could therefore have longer shelf lives than other dosage forms.

If further proven to be safe, effective and not interfere with the breastfeeding process (13-16), the NSDS has the potential to compliment dispersible tablet use in environments where infants breastfeed exclusively and remove many of the issues which can cause contamination of reconstituted drugs delivered to infants in resource-limited countries. More than 5.9 million children under 5 years of age died
in 2015. Many of these could have been prevented with access to appropriate forms of simple and affordable medicines and corresponding hygienic administration methods. (17)

Zinc supplements, containing zinc sulphate have been shown to reduce the duration of diarrhoea episodes by 25% and are also associated with a 30% reduction in stool volume (9). Zinc supplements are available in developing countries but require cup, bottle, or spoon delivery of the reconstituted suspension, limiting breastfeeding infant acceptability. Zinc sulphate, for which concentrations in HBM have not been conclusively shown to be impacted by maternal supplementation (18), constitutes therefore a relevant and important API to be delivered directly to neonates in an age appropriate manner.

One safety consideration raised and addressed in this work, is the potential maternal tolerability of a modified NSDS. Specifically assessment of dermatological impacts due to potential irritancy of a concentrated suspension/slurry of APIs or excipients in HBM on the mother’s nipples is required. There is no pharmaceutical safety assessment guidance to study the local tissue tolerance of the nipple and the areola. The primary dermal irritation is the test rabbit screening procedure. The concept of the three Rs (refinement, reduction and replacement of laboratory animals) strongly stimulates the development of alternative testing methods, such as in vitro methods and the use of “lower” organisms as test species (e.g. invertebrates, plants and microorganisms). Presently the SMI assay was identified and explored as an in vivo assessment tool for this novel application. It was initially developed at the Laboratory of Pharmaceutical Technology at the University of Ghent to predict the mucosal irritation potency of pharmaceutical formulations and ingredients. (19, 20) The premise of the test is that slugs that are placed on an irritating substance will produce mucus. Tissue damage
can be induced which results in the release of proteins and enzymes from the mucosal surface. Topical local tolerance effects being almost entirely limited to irritation, several studies have shown that the SMI assay is a useful tool for evaluating the local tolerance of pharmaceutical formulations and ingredients. (20-27) A classification prediction model that distinguishes between irritation (mucus production) and tissue damage (release of proteins and enzymes) has been developed. The SMI study is proposed more acceptable and ethical in terms of the principles of reduction, refinement and replacement, compared to previous tests such as the Draize test (23), an invasive procedure which involves applying relatively large volumes (0.5 mL or 0.5 grams of a test substance to the eye or skin of a restrained, conscious mammal (usually a rabbit) and recording its effects. The SMI assay is a simple yet efficient way, to assess mucosal tissue irritation without using large numbers of vertebrates such as mice, rabbits or non-human primates or using more complex reconstructed human epidermis 3D skin models.

The SMI assay complements existing predictive assays which are used in early pharmaceutical development and even has the potential to be used instead of the Draize test. Indeed, the relevance of the SMI assay to reliably predict nasal irritation, stinging and burning sensations has been demonstrated in a clinical trial using several Over the Counter (OTC) liquid nasal formulations, isotonic, and hypertonic saline. (29) It has also been shown that an increased mucus production with exposure to diluted shampoos was related with an increased incidence of stinging and burning sensations in the human eye irritation test. (30) The objective values obtained by means of the predictive SMI model for the mucus production, stinging, itching, or burning potential of the test blends can be estimated according to four categories (none, mild, moderate
and severe). The limits for degree of discomfort on nasal and ocular mucosal surfaces are summarized in Table 1 (29, 30).

The purpose of the present study was to attempt to predict topical irritation namely the stinging, itching or burning potential of a range of GRAS powder blends on the human nipple, some of which contain the model compound zinc sulphate, used in the treatment of diarrheal disease to support the development of a rapidly disintegrating tablet.

**MATERIALS AND METHODS**

**Methods**

**Slugs**

The parental slugs of *Arion lusitanicus* were collected in local gardens along Ghent and Aalter (Belgium) and bred in an acclimatized room at 18 - 20 °C. Test slugs were housed in plastic containers and fed with lettuce, cucumber, carrots, and commercial dog food. Slugs weighing between 3 and 6 g were isolated from the cultures two days before the start of an experiment. The body wall was inspected carefully for evidence of macroscopic injuries. Only slugs with clear tubercles and with a foot surface that showed no evidence of injuries were used for testing purposes. The slugs were placed in a plastic box lined with paper towel moistened with phosphate buffered saline (PBS) (0.01 M phosphate buffer, 0.0027 M potassium chloride, and 0.137 M sodium chloride) (Sigma Aldrich, St. Louis, MO, USA) at pH 7.4 and were kept at 18 – 20 °C. Daily, the body wall of the slugs was wetted with 300 µl PBS using a micropipette.

**Human breast milk**

Anonymised HBM samples were obtained from approximately 20 healthy donors from the Queen Charlotte’s and Chelsea Hospital Milk Bank (Imperial College Healthcare
NHS Trust). The donors had all consented for their milk to be used for research as it was not able to be used for donation. They were screened for HIV 1 and 2, HTLV I and II, hepatitis B and C, and syphilis, and ethical approval for use was obtained from the University of Cambridge (Cambridge Human Biology Research Ethics Committee (REC), University of Cambridge (REC number HBREC.2012.01). Milk from 10 donors was pooled for the experiment, half of which was centrifuged at 5411 g (5500 RPM) using a Sigma 3–16 PK centrifuge (Sigma–Zentrifugen, Osterode, Germany) for 15 min. A fat layer obtained at the top of the flask was then carefully removed using a curved face spatula, and the remaining milk was pooled into a single flask. This fat layer, fat-free milk layer and the milk not centrifuged were then combined in this study to achieve a standardised fat content of 4.6 % wt. Samples were then placed in 50 mL centrifuge tubes and stored at −80 °C to be used thawed prior making the slurries.

Test slurries

The following materials were sourced for the manufacture of blends A - I: zinc sulphate monohydrate (Sigma, Sigma-Aldrich, Dorset, UK); lactose monohydrate (direct compression grade, DFE Pharma Goch, Germany); sodium starch glycolate (Explotab, Mendell GmbH Volklingen, Germany); sodium croscarmellose (FMC Biopolymer, Girvan, UK); microcrystalline cellulose (MCC) (Avicel PH102, FMC Biopolymer, UK); magnesium stearate (Sigma-Aldrich, Dorset, UK); crospovidone (Polypasdone XL, Ashland, UK) and sodium stearyl fumarate (Alubra PG-100, FMC Biopolymer, UK).

The following 330 mg blends, corresponding to the composition of one NSDS tablet containing 20 mg elemental zinc (6% w/w) based on blend A and I, detailed in Table 2, were prepared by hand filling the individual components in to 6.5 mL glass
scintillation vials followed by blending in a Turbula mixer at 44 rpm for 5 minutes. Blend A was comprised of a mixture of zinc sulphate and lactose (filler or bulking agent) in combination with functional excipients at levels commonly used in the formulation design of a rapidly disintegrating immediate release solid oral dosage form, namely: microcrystalline cellulose (compression aid), sodium starch glycolate (disintegrant), croscarmellose sodium (disintegrant) and magnesium stearate (lubricant). The composition of the blends E, F, G and H were chosen based on variations of the lead platform formulation (A) to comprise lactose and each of the functional excipients in blend A individually. An alternative disintegrant (crospovidone, blend B) and lubricant (sodium stearyl fumarate, blend C) were also evaluated. Blends D and I acted as controls for zinc sulphate, and comprised lactose with all of the functional excipients and zinc sulphate respectively. The blends were reconstituted into an homogenous slurry with the aid of vortex mixing until visually suspended, using 1 mL HBM. This volume was chosen to mimic a scenario where burst release of the API and excipients occurred, thus yielding potentially a worst case concentration for the compounds. The final slurries were designed to represent the most concentrated suspension that would be in contact with the mother’s nipple assuming near-instant disintegration of the tablet during breastfeeding.

Methods

The SMI assay

Before a test was considered valid, the following criteria was met: the negative control (PBS) generated a total mucus production less than 5.5% of initial body weight to be classified as causing no stinging, itching, or burning; the positive control (1 w/v% benzalkonium chloride (BAC) (Sigma Aldrich, St. Louis, MO, USA)) in PBS generated
a total mucus production above 17.5% of initial body weight to be classified as causing severe stinging, itching, and burning.

The stinging, itching, or burning (SIB) potency of the test items and the negative and positive controls were evaluated by placing 3 slugs per treatment group 3 times a day on 100 µl of test slurry in a Petri dish. After each 15-min contact period (CP) the slugs were transferred for 60 min into a fresh Petri dish on paper towel moistened with 1 ml PBS to prevent desiccation. After the third CP the slugs were placed in a Petri dish on a membrane filter (cellulose acetate 0.45 µm, 90 mm diameter, Sartorius AG, Goettingen, Germany) moistened with 2 ml PBS until the next day. The overview of the test procedure is illustrated in figure 2.

The amount of mucus produced during each contact period was measured by weighing the Petri dishes with the test item before and after each 15 minute CP. The mucus production was expressed as the % of the body weight. The slugs were weighed before and after each 15 minute CP and 24 hours after the first CP. The total mucus was calculated for each slug and then the mean per treatment group was calculated.

Based on the endpoints of the SMI assay the stinging, itching, or burning potency of the test item(s), as defined in Table 1, was estimated using a classification prediction model. Mortality was documented for slugs exposed to each of the sample slurries, including the controls and HBM 24 hours after the third CP.

RESULTS AND DISCUSSION

The primary aim of this research was to evaluate the potential of commonly used tablet excipients and zinc sulphate used as an antidiarrheal model compound to cause irritation of the human nipple through extrapolation of the SMI assay. This utilizes the
terrestrial slug *Arion lusitanicus*. The body wall of the slugs is layered and has a mucosal surface. The outer single-layered columnar epithelium contains cells with cilia and micro-villi. Mucus secreting cells cover the subepithelial connective tissue. This micro-anatomy is similar to that of the lactiferous ducts of the mother’s nipple which are lined by a columnar epithelium supported by myoepithelial cells. Hence the SMI was hypothesized to be a potential predictive model for nipple irritancy during drug administration via the NSDS. It was assumed that the results of this study would represent a worst case assessment for irritation, since ocular/nasal surfaces are more sensitive than skin.

The average amount of mucus produced during each 15 minute CP and total MP is presented in Table 3, Figure 3.

According to the classification prediction model of the SMI test, the negative control (PBS) did not induce reactions in the slugs (total MP < 3 - 5.5%). The positive control on the other hand (BAC 1% w/v) induced, as expected, a high mucus production during each contact period (total MP ≥ 17.5%) resulting in a classification corresponding to severe stinging, itching, and burning (SIB) reactions. The acceptance criteria were met and the experiment was considered valid. Graphical summaries of the data are presented in Figures 3 and 4.

The total mucus production values for the reconstituted blends are bracketed by the positive and negative controls. Slugs treated with HBM did not produce an increased amount of mucus compared to the negative control. Similarly the blends that did not contain zinc sulphate did not increase mucus production to a level at which it could be classified as “mildly irritant” (Table 1), with the exception of blend H containing lactose and magnesium stearate, which only just exceeded by 0.1% the limit for “no irritation” as classified using the ocular classification only (Table 1 and Table 2). The
total MP for HBM is only slightly negative, and it is therefore concluded that HBM was tolerated very well, resulting in a classification as not causing discomfort. The negative MP been induced (< -0.7%) may have also resulted in tissue damage, but this was not the case.

Slugs treated with blends B, C, D, E, F, and G produced only a slightly increased amount of mucus during each contact period, compared to the negative control. All placebo blends resulted in a total MP < 3 and 5.5%, corresponding with the classification “no SIB reactions”, suggesting acceptable tolerability. For all placebo blends the first contact induced the highest MP, but was much lower during the second and third CP. The slightly negative mucus production that was observed for the negative control in the third CP was hypothesised to be due to the fact that the slugs produced only a minimal amount of mucus during each contact period and also that only a minimal amount of the test substance remains on the body wall of the slugs.

Slugs treated with blends A and I, both containing the API zinc sulphate at a concentration of approximately 6% w/w, produced a higher amount of mucus during each CP in comparison with the negative control and all other blends tested. For blend A, the total MP also increased for the first two CPs; in the third CP it was lower than during the first CP. For blend I, MP was comparable during the first and second CPs, but increased substantially during the third contact period. This is however within the accepted limits. Similar reactions were observed in other experiments, where there is a certain tolerance for the first two CPs, and then an overreaction in the third CP. Although it can be interesting to look at the results of the three CPs separately, it is the total MP over the three CPs together that is used for the classification, as there can be quite some variability in the slugs’ reactions. Both blends induced a total MP
between 10% and 17.5% and were therefore classified as causing moderate SIB reactions.

With an n of 3 for each of the reconstituted slurries, considered alongside formulation (e.g. suspension homogeneity, which was not evaluated) and in vivo variability, it is not possible to assign statistical similarities or differences between the formulations in terms of mucus production, with a large degree of confidence. However, the data suggests that the zinc sulphate is the major contributing factor to mucosal irritancy, and that the lack of response to the GRAS excipients evaluated promotes a large formulation design space. It is to be noted that the concentrations tested represent a higher concentration than would be expected to be released using the NSDS, since it would likely take more than 1 mL of HBM to disintegrate a tablet during use. (12) Therefore, zinc sulfate may not cause as much irritancy as implied by this worst case study.

No mortality was observed immediately following the three CPs, however 24 hours after the final CP, two out of three slugs had died in the cohort exposed to the positive controls as well as formulations A and I (both containing zinc sulphate). The observed mortality in the slugs exposed to the positive control, was not surprising. Slugs treated with benzalkonium chloride using a similar protocol (30 min exposure to irritant each day for 5 days), often indicate tissue damage after the first contact period. (20, 21) This damage accumulates over time, inducing mortality of the slugs. In the current study, damage also occurred after exposure to benzalkonium chloride, although this was not quantified using microscopy, resulting in the observed mortality.

Further, zinc salts (including sulphate) have been previously demonstrated to be highly toxic to freshwater fish and invertebrates and metal sulphates have been incorporated into proprietary slug and snail control products. (31) Exposure to elemental zinc in food
(albeit at much higher concentrations) has been shown to have negative developmental impact on slugs. (32, 33)

The relationship between API chemical structure and the endpoints mortality and MP, has previously been studied for the antimicrobial agent, benzalkonium chloride, and it was concluded that the activity and toxicity of different analogues depend on the alkyl chain length of the bactericidal molecule. (21) The slug mortality and increase in MP of the slugs exposed to slurries containing zinc sulphate appears to be due to the API and possibly related to its chemical structure; further investigations around API structure / SMI assay activity may help to better validate the model for different applications. While the SMI data generated in this study suggest a possibility of local irritation on the human skin, the dermatological impact may be less than implied with this relatively non-invasive testing method, when nasal and ocular classification scales are used, as these are likely to be more sensitive than human skin.

Further validation of the model is required to build an in vivo / in vivo correlation between the SMI model and irritancy of the breastfeeding nipple during administration of a rapidly disintegrating immediate release tablet via the NSDS. Such a correlation would be a useful pre-formulation tool to assess risk during early formulation development. Additional opportunities to improve the predictiveness of the model for the selection of APIs for use with the NSDS include gaining a better understanding of similarities and differences of the histology of the breastfeeding nipple and the slug mucosa, considering species other than Arion lusitanicus for the assay, and essentially, optimising exposure time of the slugs to the concentrated slurries to correlate to an average breast feed. Since the average HBM mass delivered per feed is estimated to be 50-80g of milk over a 7 to 10 minute period, the study of the tablet
components at lower concentrations, correlating to slower release from the NSDS could be tested for irritancy potential. (34-36)

CONCLUSIONS
This was the first SMI study to be conducted evaluating excipients used in the manufacture of solid oral dosage forms, slurried in HBM. The existing SMI model has indicated a potential for local dermatological irritation when zinc sulphate at a potentially worst case concentration is delivered via the NSDS, with further investigation warranted. Mild irritation is suggested by the change in MP, and body weight of the slugs exposed to slurries of lactose and functional GRAS excipients that are used to induce rapid release of API from solid oral dosage forms. This implies these excipients could be used within functional levels as part of a flexible formulation design space that could facilitate the incorporation of a wide range of non-irritant APIs, with different physicochemical properties, into tablets for use with the NSDS. This could potentially broaden the utility of this novel drug delivery platform for the clinical treatment of neonates.

ACKNOWLEDGEMENTS AND DISCLOSURES
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to the HBM samples. These samples were provided by the Imperial College Healthcare NHS Trust Tissue Bank. Other investigators may have received samples from these same tissues. The research was supported by the National Institute for Health Research (NIHR) Biomedical Research Centre based at Imperial College Healthcare NHS Trust and Imperial College London. The views expressed are those of the authors and not necessarily those of the NHS, the NIHR or the Department of Health.

Stephen Gerrard is an inventor of the nipple shield delivery system (US patent 8357117 B2. See http://justmilk.org).

REFERENCES


Fig. 1. A) Illustration of the Nipple shield delivery system (NSDS) design. B) NSDS prototype (11)
Fig. 2. Overview of the slug mucosal irritation (SMI) assay test procedure

15’ Contact Period
100 µl test item

Transfer slug on wet paper towel in a new Petri dish for 1 h.

Measure Mucus production CP1

n=3

15’ Contact Period
100 µl test item

Transfer slug on wet paper towel in a new Petri dish for 1 h.

Measure Mucus production CP2

n=3

15’ Contact Period
100 µl test item

Measure Mucus production CP3

n=3
**Figure 3.** Total % MP of slurried tablet blends A to I [PBS: negative control; BAC1% positive control; HBM: human breast milk, * contains Zinc]
Fig. 4. Mean slug body weights as a function of time
Table I. Cut-off values of Total Mean Mucus Production (%) for classification of ocular and nasal mucosal discomfort with the slug mucosal irritation (SMI) assay:

(24, 25)

<table>
<thead>
<tr>
<th>Degree of discomfort</th>
<th>Nasal</th>
<th>Ocular</th>
</tr>
</thead>
<tbody>
<tr>
<td>None</td>
<td>&lt; 5.5%</td>
<td>&lt; 3%</td>
</tr>
<tr>
<td>Mild</td>
<td>≥ 5.5 – &lt; 10%</td>
<td>≥ 3 – &lt; 8%</td>
</tr>
<tr>
<td>Moderate</td>
<td>≥ 10 – &lt; 17.5%</td>
<td>≥ 8 – &lt; 15%</td>
</tr>
<tr>
<td>Severe</td>
<td>≥ 17.5%</td>
<td>≥ 15%</td>
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</table>
## Table II. Blends (total 330mg) for reconstitution in HBM (1ml) as slurries for SMI evaluation

<table>
<thead>
<tr>
<th>Ingredient</th>
<th>Blend (A)</th>
<th>(B)</th>
<th>(C)</th>
<th>(D)</th>
<th>(E)</th>
<th>(F)</th>
<th>(G)</th>
<th>(H)</th>
<th>(I)</th>
</tr>
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<tbody>
<tr>
<td>Zinc sulphate</td>
<td>54.9</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Direct compression lactose</td>
<td>222.3</td>
<td>297.0</td>
<td>325.0</td>
<td>277.2</td>
<td>320.1</td>
<td>323.4</td>
<td>297.0</td>
<td>326.7</td>
<td>275.1</td>
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<td>Sodium Starch Glycolate</td>
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<td></td>
<td></td>
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<td></td>
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<td>Crosscarmellose sodium</td>
<td>6.6</td>
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<td>6.6</td>
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<td></td>
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<tr>
<td>Microcrystalline cellulose</td>
<td>33.0</td>
<td>33.0</td>
<td>33.0</td>
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<td></td>
<td></td>
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<tr>
<td>Magnesium stearate</td>
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<td></td>
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<tr>
<td>Crospovidone</td>
<td>33.0</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Sodium stearyl fumarate</td>
<td>5.0</td>
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Table III. Amount of mucus produced during each 15 minute CP and total amount of mucus produced

<table>
<thead>
<tr>
<th>Formulation</th>
<th>MP CP1&lt;sup&gt;1&lt;/sup&gt; (%)</th>
<th>MP CP2&lt;sup&gt;1&lt;/sup&gt; (%)</th>
<th>MP CP3&lt;sup&gt;1&lt;/sup&gt; (%)</th>
<th>Total MP&lt;sup&gt;1&lt;/sup&gt; (%)</th>
<th>SIB</th>
<th>Category&lt;sup&gt;2&lt;/sup&gt;</th>
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<tr>
<td>NC - PBS</td>
<td>0.2 ± 0.1</td>
<td>0.2 ± 0.5</td>
<td>-0.3 ± 0.1</td>
<td>0.2 ± 0.3</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>PC – BAC 1% w/v</td>
<td>5.7 ± 0.6</td>
<td>7.1 ± 2.2</td>
<td>7.7 ± 1.5</td>
<td>20.4 ± 4.3</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>A*</td>
<td>3.9 ± 1.1</td>
<td>6.0 ± 1.4</td>
<td>2.6 ± 1.4</td>
<td>12.5 ± 1.5</td>
<td>Moderate</td>
<td></td>
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<tr>
<td>B</td>
<td>0.8 ± 0.1</td>
<td>0.5 ± 0.4</td>
<td>0.6 ± 0.3</td>
<td>1.9 ± 0.5</td>
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<td></td>
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<tr>
<td>C</td>
<td>1.4 ± 0.4</td>
<td>0.7 ± 0.4</td>
<td>0.6 ± 0.3</td>
<td>2.6 ± 0.3</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>D</td>
<td>0.9 ± 1.5</td>
<td>0.5 ± 0.5</td>
<td>0.4 ± 0.1</td>
<td>1.9 ± 1.9</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>E</td>
<td>1.4 ± 1.2</td>
<td>0.3 ± 1.3</td>
<td>0.8 ± 0.4</td>
<td>2.5 ± 1.4</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>F</td>
<td>1.2 ± 0.0</td>
<td>0.8 ± 0.3</td>
<td>0.5 ± 0.7</td>
<td>2.4 ± 0.8</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>G</td>
<td>1.5 ± 0.6</td>
<td>0.0 ± 0.6</td>
<td>0.9 ± 0.9</td>
<td>2.4 ± 0.8</td>
<td>No</td>
<td></td>
</tr>
<tr>
<td>H</td>
<td>1.6 ± 0.7</td>
<td>0.5 ± 0.3</td>
<td>0.9 ± 0.3</td>
<td>3.1 ± 1.3</td>
<td>No</td>
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</tr>
<tr>
<td>I*</td>
<td>3.3 ± 0.2</td>
<td>3.3 ± 0.7</td>
<td>4.7 ± 0.6</td>
<td>11.3 ± 1.0</td>
<td>Moderate</td>
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<tr>
<td>J – HBM</td>
<td>0.0 ± 1.0</td>
<td>-0.2 ± 0.3</td>
<td>-0.5 ± 0.7</td>
<td>-0.7 ± 1.9</td>
<td>No</td>
<td></td>
</tr>
</tbody>
</table>

<sup>1</sup>Mean ± SD, n = 3;  <sup>2</sup>SIB (see table 1) NC: negative control; PC: positive control; HBM: human breast milk; * contains Zinc