

abYsis: Integrated Antibody Sequence and Structure — Management, Analysis and Prediction

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Abstract

abYsis is a web-based antibody research system that includes an integrated database of antibody sequence and structure data. The system can be interrogated in numerous ways — from simple text and sequence searches to sophisticated queries that apply 3D structural constraints. The publicly available version includes pre-analysed sequence data from EMBL-ENA and Kabat as well as structure data from the PDB. A researcher's own sequences can also be analysed through the web interface.

A defining characteristic of abYsis is that sequences are automatically numbered with a series of popular schemes such as Kabat and Chothia and then annotated with key information such as CDRs and potential post-translational modifications. A unique aspect of abYsis is a set of residue frequency tables for each position in an antibody, allowing 'unusual residues' (those rarely seen at a particular position) to be highlighted and decisions to be made on which mutations may be acceptable. This is especially useful when comparing antibodies from different species.

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abYsis is useful for any researcher specializing in antibody engineering, especially those developing antibodies as drugs.

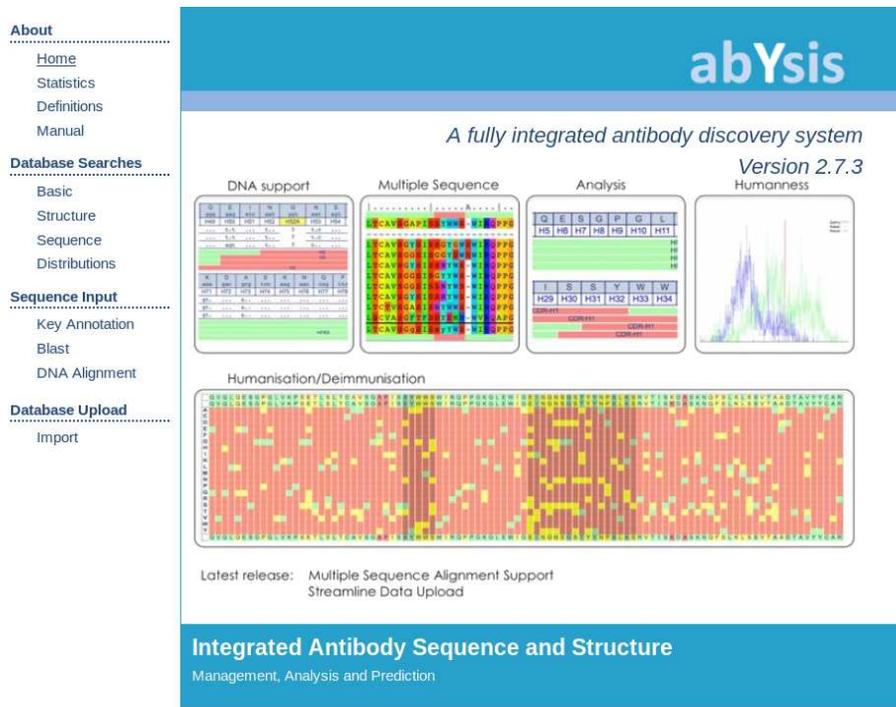
abYsis is available at www.abysis.org.

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1. Research highlights

- abYsis is a web-based antibody research system which integrates sequence data from EMBL-ENA and Kabat with sequence and structure data from the PDB.
- It allows searches based on text, sequence data and structural constraints.
- It automatically numbers sequences and provides detailed annotations to aid interpretation.
- Users can upload and analyze their own sequences in the same way via the web interface.
- It provides a comprehensive set of position and species-specific residue distributions.
- Additional advanced tools are provided to assess antibody characteristics such as humanness.

2. Graphical Abstract



3. Introduction

Antibodies are amongst the most important classes of proteins involved in the adaptive immune system. They act as adapter molecules between the virtually infinite range of possible antigens and the constant effector molecules such as Fc receptors and C1q of the complement system. Through DNA-level splicing of V and J gene segments in the light chain and V, D and J gene segments in the heavy chain, they create a repertoire of around 10^{11} possible sequences[1]. The fact that antibodies are capable of binding to a virtually infinite set of antigens, generally with high specificity and affinity, has led to a huge interest in using them as drugs in the treatment of human disease[2].

Therapeutic antibodies can act through a variety of mechanisms. First, they may act simply to bind to a target in the way that most small-molecule drugs

work. For example, they may bind to receptors acting as agonists or antagonists, or to enzymes acting as inhibitors or as allosteric activators. Since they have a much larger interacting surface than typical small-molecule drugs (which generally bind into pockets on a protein surface), they can also interact with much flatter protein surfaces and can therefore be used to bind to hormones or disrupt protein-protein interactions. Second, antibodies can be used to trigger the immune system in the normal way to kill cells such as tumour cells or virus-infected cells. For example, the antibody, Palivizumab (Synergis), is used to prevent respiratory syncytial virus (RSV) infection in high-risk infants. Third, antibodies can be used as a delivery mechanism, either through antibody-drug conjugates with cytotoxic moieties or through ‘antibody-directed enzyme pro-drug therapy’ (ADEPT), which aims to restrict the action of a cytotoxic drug to cancer sites[3]. Fourth, more recent developments have led to bispecific antibodies capable of binding two antigens, either for convenience of having a single agent, or deliberately cross-linking different molecules[4, 5, 6].

It has been estimated that around a third of all drugs in development are monoclonal antibodies[7, 8] and as of May 2016, according to the Antibody Society, 55 monoclonal antibody products had been approved in the US or Europe with around four new products being approved per year[9]. Therapeutic antibodies have been used for a wide variety of conditions such as cancer[10], transplant rejection[11], rheumatoid arthritis[12], antiviral prophylaxis[13] and Crohn’s disease[14].

Despite the success of antibodies in the clinic, there are a number of problems in developing successful therapeutics. As well as high affinity and specificity, antibodies must be non-immunogenic and have sufficiently high bio-availability. In addition, stability is important to avoid denaturation and aggregation, not only for long shelf-life and persistent bio-availability, but also because denaturation and aggregation can lead to increased immunogenicity[15, 16, 17]. Consequently there is still room for improvement in building better antibodies having higher affinity and lower immunogenicity[18].

Traditionally, producing fully human antibodies has been difficult while an-

antibodies from other species are likely to lead to an immune response (the ‘Human Anti-Mouse Antibody’ (HAMA) response[19, 20], or more generally the ‘Anti-Antibody Response’, AAR[21]). This prevents repetitive administration of the antibody for treatment and may lead to anaphylactic shock. Traditional methods for producing monoclonal antibodies[22] do not work in humans[23, 24]. Techniques such as SLAM[25] allow screening of B cells harvested from peripheral blood, spleen, bone marrow or lymph nodes and, as well as being applicable to non-human species, can be used to identify therapeutic antibody candidates from human peripheral blood. While this can be useful for creating anti-infective agents, the majority of targets are human proteins against which humans do not produce an immune response. While the production of antibodies from phage-display libraries has been very popular[26, 27], the success of these agents in the clinic has so far been limited. Another more recent development is the production of fully human antibodies from transgenic mice[23] where the antibody repertoire is fully human, but this route is extremely expensive. Consequently murine (or other non-human) antibodies are frequently engineered to make them appear ‘more human’. The process of humanization aims to reduce immunogenicity of non-human antibodies by using human constant and variable regions as an ‘acceptor’ into which non-human CDRs from the ‘donor’ antibody with the required specificity are inserted[28, 29]. However, in general naïvely grafted antibodies do not bind well and some framework residues need to be ‘back-mutated’ to the equivalent donor residue in order to restore the binding affinity[30].

Despite the interest in antibodies and the need to engineer them, the range of freely available web-based tools and resources to aid in this process is limited. Probably the best known resource is IMGT[31], a comprehensive collection of data and services available over the web. There are a number of web-based antibody modelling servers, the performance of which was recently compared in the second antibody modeling assessment (AMA-II)[32]. These include PiGS[33], (www.biocomputing.it/pigs), SAbPred[34] (opig.stats.ox.ac.uk/webapps/sabdab-sabpred/WelcomeSAbPred.php),

Total sequences in EMBLIG	127450
Total sequences in IMGT	96970
Total sequences present in both datasets	91898
Total missing from EMBLIG that are present in IMGT	5072
Total missing from IMGT that are present in EMBLIG	35552

Table 1: Comparison of the data in IMGT (release 16Jul2016) and EMBLIG (release 18Apr2016). Sequences in IMGT are filtered to include only antibody variable domain sequences derived from EMBL-ENA. Both datasets were filtered to ensure that translations were at least 50 residues in length.

Rosetta Antibody[35, 36], Kotai Antibody Builder[37] (kotaiab.org) and SmrtMolAntibody[38] (www.macromoltek.com). SAbPred also includes antibody numbering[39] and epitope and paratope prediction. Other resources include SAbDab[40], a database of antibody structures, (opig.stats.ox.ac.uk/webapps/sabdab-sabpred/Welcome.php), Tabhu, a set of tools to aid with antibody humanization[41] (circe.med.uniroma1.it/tabhu/) and Digit[42], a database of antibody variable domain sequences (circe.med.uniroma1.it/digit/).

For many years we have offered a number of web-based antibody tools including KabatMan[43], SACS[44], AbNum[7], expressed-sequence-scored humanness[45], germline-scored humanness[46], V_H/V_L packing-angle prediction[47], assignment of Chothia canonical classes[48] and testing a sequence for unusual residues[43]. These are all available via www.bioinf.org.uk/abs/.

Here we describe our abYsis system, a database of antibody sequence and structure which also integrates many of the analysis tools described above.

4. Results

abYsis contains antibody protein sequences from EMBLIG which contains antibody information extracted from the EMBL-ENA databank (Couch, Porter,

Data Sources	Protein Sequences			Classified			
	Sequences	$\geq 70\text{aa}$	Numbered	Total	Heavy	Light	Paired
EMBLIG	126422	119422	101969	122745	92179	30566	3555
Kabat	19399	14114	12631	19399	11842	7557	2572
PDB	7712	7712	7534	7534	3820	3714	3397
Total	153533	141248	122134	149678	107841	41837	9524

Table 2: Processed content of the abYsis database as of May 2016. In addition, DNA data are available for sequences from EMBLIG and Kabat. The NCBI Germline data and germline information from V-BASE are also included.

Swindells and Martin, in preparation), the Kabat collection[49] (which has not been updated since July 2000), and the Protein Databank (PDB)[50]. Data from other sources such as IMGT can also be loaded, but the vast majority of sequence information in IMGT is contained in EMBLIG or the PDB which we handle separately. In fact, EMBLIG contains many antibody sequences that are not present in IMGT. Comparing the two datasets as described in the Materials and Methods, we found that the two datasets overlap by 28,388 sequences. EMBLIG contains 94.8% of the sequences in IMGT, but 27.9% of sequences in EMBLIG were not present in IMGT; EMBLIG contains $1.3\times$ more sequences than IMGT. See Table 1.

The ability to integrate data from other sources is particularly useful for people who decide to have abYsis in-house as they can include their own sequence data in the resource. Table 2 shows the processed content of the latest release of abYsis (V2.7.3) as of May 2015. Automated numbering of sequences using abnum[7] is key to the annotation of sequences — the current release successfully numbers 122,134 sequences. The system also provides automated pairing of light and heavy chains. Resources such as EMBL-ENA (and therefore EMBLIG) and Kabat do not contain a clear pairing field to link data for light and heavy chains, although a ‘DR/partner’ field is being populated in IMGT and we exploit this when reading IMGT data. Consequently, as described in the Materials and Methods, we attempt to take a cautious approach to pairing

for other data sources. For EMBLIG data, we use the clone information to perform the pairing: 7,110 chains are paired into 3,555 antibodies while antibodies containing a single chain occur 119,312 times because this clone information is only present for $\sim 56\%$ of chains in EMBLIG. The most highly represented species are *Homo sapiens* (88,662), *Mus musculus* (41,254), *Macaca mulatta* (3,561), *Oryctolagus cuniculus* (2,668), *Equus caballus* (1,138), *Oncorhynchus mykiss* (979), *Lama glama* (955), *Camelus dromedarius* (930), *Ginglymostoma cirratum* (893), with synthetic constructs coming in fourth place with 2,968 entries.

The reason that numbering is key to the annotation is that it enables us to identify regions of the sequences (CDRs and frameworks). It provides an implicit sequence alignment (which can be displayed in abYsis using JSAV[51]) and mapping of sequence to structure allowing us to look at the frequency of residue types at each position and to identify key residues that define the conformation of the CDRs[52] or which are important in antibody humanization[30, 53].

abYsis can be used in three main ways: a) by searching the database to find information about sequences (and structures) including detailed annotations, identification of unusual residues, post-translational modification sites, etc., b) by analyzing trends across sequences in the database and c) by entering your own sequences for analysis. These three approaches are described below.

4.1. Searching the database

The main search page allows searches based on identifiers, names, antigens (where described and parsable in the source data), references, authors, organisms, etc., and searches can be restricted to a given dataset (PDB, Kabat, EMBLIG, or other datasets that have been included) or to a given class of chain. In addition, searches can be structure-based by requiring CDRs to match specific canonical classes or specifying residues within a given distance of a particular position; for example, one could find all antibodies with a lysine within 4Å of residue L23. This can be done either in known structures or by using averaged distances calculated across all known structures to extrapolate these constraints

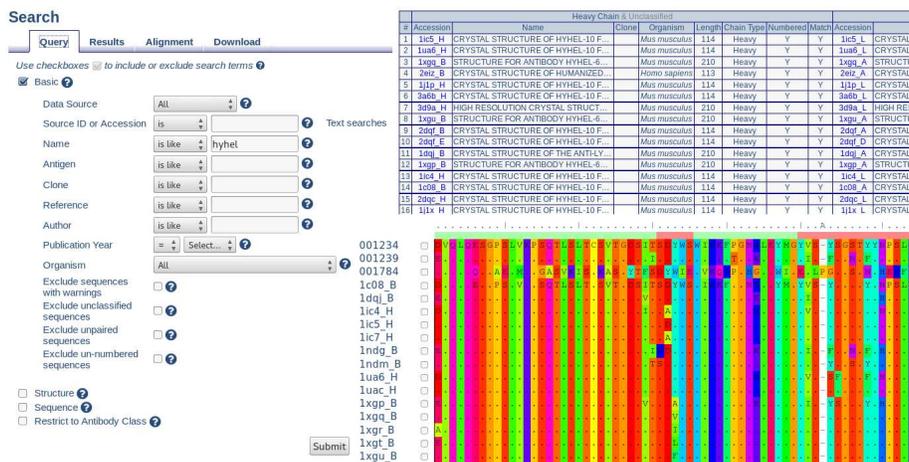


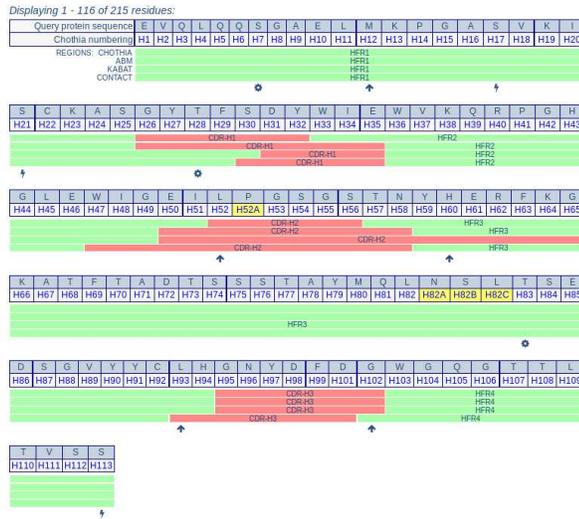
Figure 1: Simple searches of abYsis. On the left is the main search page showing the default simple options. Searching for ‘hyhel’ in the Name field results in a table of hits shown upper right. For clarity, this has been truncated — the full view would show the complete information for the light chains to the right of the heavy chains where paired sequences have been found. Bottom right shows the alignment view of the sequences.

to sequences for which no structure is available. Finally one can also search on the basis of sequence. Searches can specify motifs (either exact or using regular expressions) present throughout the sequence or within particular regions (CDRs, frameworks), region lengths, or residues present or not present at particular positions.

A simple example search is shown in Figure 1 together with two views of the results — as a simple table or as a sequence alignment. The results come from searching for ‘hyhel’ and therefore displaying the HyHEL set of antibodies binding to hen egg white lysozyme described by Smith-Gill and colleagues[54].

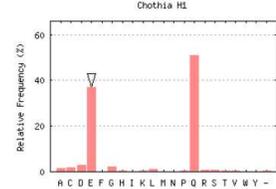
Having selected a particular sequence of interest, much more detailed information is provided in what we term the ‘Key Annotation’ page which provides a set of tabs with different information. The initial summary page provides the sequence with Chothia numbering applied and the Chothia structural loops highlighted. Unusual residues occurring in < 1% of sequences across all species are highlighted as are a range of predicted post-translational modification sites.

a)



Distribution for Chothia position H1

Query sequence has 'E'.
 Select residue numbers for other distributions.



Amino Acid	Non-identical Sets	Relative Frequency	Amino Acid	Non-identical Sets	Relative Frequency
M	17	<1%			
N	14	<1%			
P	108	<1%			
Q	23270	51%			
R	266	<1%			
S	334	<1%			
T	126	<1%			
V	139	<1%			
W	4	<1%			
Y	14	<1%			
-	119	<1%			
Total	45757	100%			

Download a Postscript version of the graph.

b)

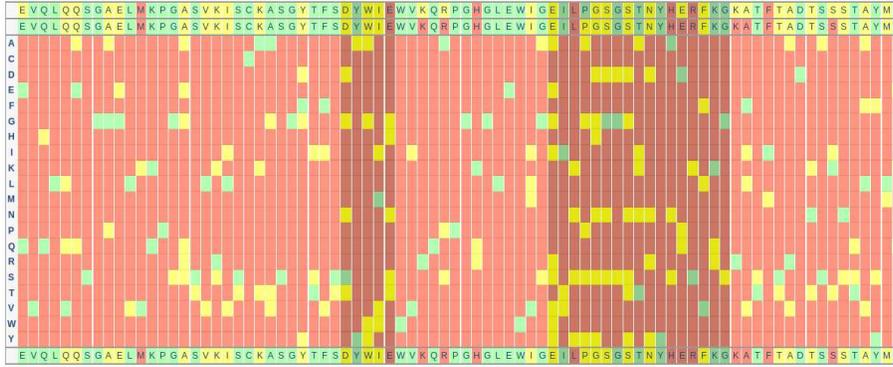


Figure 2: Key annotation results for the heavy chain of antibody HyHEL-5. a) the 'Numbering & Regions' tab — unusual residues are highlighted with an upwards arrow, predicted CK2 phosphorylation sites with a gear wheel and predicted PKC phosphorylation sites with a lightning symbol; b) the 'Humanize' tab.

Information about the sequence (accession code, data source, dates, name, organism, chain type, partner chain and references) is also provided. A button at the bottom of the page allows the sequence to be searched against all the other sequences using BLAST.

The ‘Numbering & Regions’ tab (part of which is shown in Figure 2a) gives more control over numbering schemes allowing Kabat, Chothia, or Martin (Enhanced Chothia)[7] schemes as well as displaying CDRs / structural loops according to definitions provided by Kabat, Chothia, AbM (useful for modelling purposes) and contacts[55]. On the right of the panel is shown a residue distribution graph for the currently selected residue — in this case residue H1. As the graph shows, the most common residues (across all species) are glutamate (E) and glutamine (Q); the amino acid present in this particular sequence is highlighted with an arrow above the relevant bar.

The ‘Unusual Residues’ tab allows the same information to be provided, but allows the comparison to be restricted to a specific species, while the ‘Humanization’ tab gives a much more detailed view of this information. Again clicking on an amino acid will show a graph of its distribution which can be restricted to a particular species. The ‘heatmap’ which is part of the humanization tab and shown in part in Figure 2b, gives a summary of the graphical view of residue frequencies for each position in the sequence. Two thresholds are defined using sliders at the bottom of the page; for each position, amino acid types with a frequency below the lower threshold (i.e. unusual residues) are shown in red, those above the higher threshold (i.e. common) are shown in green with the remaining residues shown in yellow, the CDRs being shown in darker shades. At the top of the heatmap, the sequence is shown with colours based on a first reference sequence and then with colours against a second reference sequence which is also used for the rest of the heatmap. Clicking on a residue in the middle of the heatmap will update the sequence shown at the bottom which can then be exported. Typically one might set mouse as the first reference and human as the second reference thus allowing the identification of unusual human residues.

Other tabs provided include a) assignment of Chothia conformational canoni-

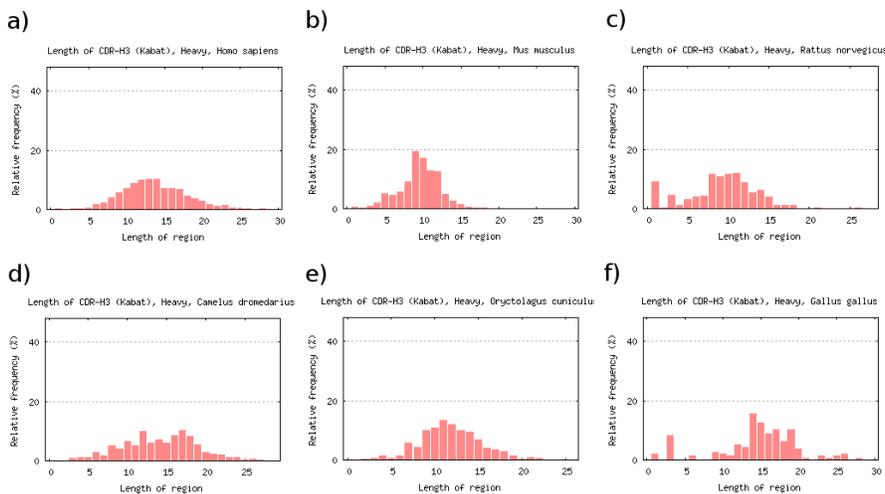


Figure 3: Distributions of CDR-H3 lengths in a) humans, b) mice, c) rats, d) camel, e) rabbit and f) chicken.

cal classes based on sequence; b) a ‘Structure’ tab, only displayed if the selected sequence comes from a PDB file, in which the structure can be viewed and rotated; c) a ‘Germline’ tab which maps a sequence to the closest germline sequences; and d) a ‘Humanness’ tab which implements the method of Abhinandan and Martin for assessing how human-like a sequence is[45].

4.2. Analyzing trends

The distributions search page allows the distributions of residues at particular positions to be analyzed outside the context of a particular sequence. It also allows distributions of the lengths of regions (CDRs and frameworks) to be analyzed. This has been an area of some considerable previous interest. For example, Wu *et al.*[56] analyzed the distribution of the lengths of CDR-H3 loops in humans and other species and concluded that ‘human sequences vary from 2 to 26 amino acids residues, but less extensively in other species’. They updated their analysis in 1993[57] refining the analysis in an attempt to look at defined specificities. Similar detailed analyses of length distributions were performed by Collis *et al.*[58] and Barrios *et al.*[59].

Such analyses become straightforward using abYsis and in Figure 3 we show the length distributions of CDR-H3 in humans, mice, rats, camel, rabbit and chicken. The graph confirms earlier observations that mouse antibodies tend to have shorter CDR-H3 lengths than humans, but also shows that this trend extends to rat antibodies. Despite the recent popularity of rabbit antibodies in generating potential clinical antibodies, the CDR-H3 lengths still tend to be shorter than human antibodies, but camels and chickens tend to have CDR-H3s that are even longer than humans.

4.3. Entering your own sequences

The sequence input pages allow one or more protein and/or DNA sequences to be entered. These are numbered and analyzed in exactly the same way as sequences stored in the database. Consequently all the ‘key annotation’ information described above is also available for these. Sequences can also be searched against the database using BLAST to find the most similar antibodies.

The ‘Humanization’ tab is of particular interest since it allows one to upload a naïvely humanized antibody (i.e. where the CDRs have been grafted from a donor species onto a human acceptor framework) and compare this with unusual residues identified in the mouse framework which may be functionally important. One can also look for unusual residues in the human acceptor which may have an effect on binding or lead to immunogenicity.

The locally-installed version of abYsis also allows sequences to be uploaded and added to the database.

5. Materials and Methods

abYsis is based around a PostgreSQL relational database populated with sequence and structure data. All sequences are numbered using Kabat, Chothia and Martin numbering schemes using abnum[7] and the database is populated using scripts written in Perl. The web interface is implemented using Perl and Mason and the Javascript Sequence Alignment Viewer (JSAV)[51] is used to display sequence alignments. JSmol[60] is used for structure display.

5.1. Comparison of EMBLIG and IMGT

A Perl script was written to compare the content of EMBLIG and IMGT. The IMGT data file contains sequences that are not antibodies and, while mostly coming from EMBL-ENA also comes from sources such as the PDB (that we treat separately). Consequently the IMGT data were filtered to ensure that the sequence a) was present in EMBL-ENA, b) was labelled as IG-Light or IG-Heavy, c) contains a variable domain, d) has a translation provided of at least 50 residues, e) is not a pseudogene. EMBLIG data were also filtered to include only sequences with translations of at least 50 residues.

5.2. Pairing antibody chains

For Kabat data, we pair antibody chains based on the provided ‘name’ field and author information to pair light and heavy chains as described by Martin[43]. For pairing chains within PDB files, we examine the proximity of residues normally found in the interface between light and heavy chains. For EMBLIG data derived from EMBL-ENA, there is no clear name field so we currently use the clone name given in the ‘FT/source/clone=’ field. As well as matching the clone name, pairs of light and heavy chains must also share publication data.

6. Conclusion

abYsis provides a unique integrated web-based environment for exploring antibody sequence and structure, bringing together data from different public sources. In addition, users who install the system locally are able to integrate their own sequence data. A key advantage of abYsis is that we try to pair light and heavy chain sequences. However, we take a cautious approach to this and, in the case of EMBLIG data derived from EMBL-ENA, we use clone information that is only present for ~56% of chains. Consequently, future work will include exploiting information in the ‘FT/CDS/product=’ information which is used to provide a ‘Name’ in abYsis. However this will require cleanup to remove uninformative text (such as ‘immunoglobulin’, ‘gamma’, ‘heavy’, ‘light’ or ‘chain’)

and, for safety, would need to be combined with author and/or publication information. The system provides annotation of sequences stored in the system, or which a user uploads, and allows individual sequences to be compared with those in the underlying database. Integration of sequence and structure data allows analysis of sequence data in a structural context and the system provides tools that can aid with antibody engineering.

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