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The H-bond network surrounding the pyranopterins modulates redox cooperativity in the molybdenum-bisPGD cofactor in arsenite oxidase.

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Abbreviations:

\[ \text{Aio: arsenite oxidase; AMPSO: 3-[(1,1-Dimethyl-2-hydroxyethyl)amino]-2-hydroxypropanesulfonic acid; EPR: Electron Paramagnetic Resonance; HEPES: 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; } K_s \text{: stability constant; MES: 2-(N-morpholino)ethanesulfonic acid; Mo: Molybdenum; MOPS: 3-(N-morpholino)propanesulfonic acid; PGD: Pyranopterin Guanosine Dinucleotide.} \]
Abstract

While the molybdenum cofactor in the majority of bisPGD enzymes goes through two consecutive 1-electron redox transitions, previous protein-film voltammetric results indicated the possibility of cooperative (n=2) redox behavior in the bioenergetic enzyme arsenite oxidase (Aio). Combining equilibrium redox titrations, optical and EPR spectroscopies on concentrated samples obtained via heterologous expression, we unambiguously confirm this claim and quantify Aio’s redox cooperativity. The stability constant, $K_s$, of the MoV semi-reduced intermediate is found to be lower than 10$^{-3}$. Site-directed mutagenesis of residues in the vicinity of the Mo-cofactor demonstrates that the degree of redox cooperativity is sensitive to H-bonding interactions between the pyranopterin moieties and amino acid residues. Remarkably, in particular replacing the Gln-726 residue by Gly results in stabilization of (low-temperature) EPR-observable MoV with $K_s = 4$. As evidenced by comparison of room temperature optical and low temperature EPR titrations, the degree of stabilization is temperature-dependent. This highlights the importance of room-temperature redox characterizations for correctly interpreting catalytic properties in this group of enzymes.

Geochemical and phylogenetic data strongly indicate that molybdenum played an essential biocatalytic roles in early life. Molybdenum’s redox versatility and in particular the ability to show cooperative (n=2) redox behavior provide a rationale for its paramount catalytic importance throughout the evolutionary history of life. Implications of the H-bonding network modulating Molybdenum’s redox properties on details of a putative inorganic metabolism at life’s origin are discussed.

Keywords: Arsenite oxidase; Molybdenum enzyme; optical spectroscopy; EPR spectroscopy; redox titrations;
1. Introduction

In prokaryotes, the enzyme arsenite oxidase (Aio), a member of the vast superfamily of the so-called Molybdenum-bisPyranopterin Guanosine Dinucleotide (Mo-bisPGD) enzymes (previously denoted as the DMSO-reductase or CISM-superfamily [1, 2]), injects reducing equivalents derived from the oxidation of arsenite into a variety of chemiosmotic electron transfer chains [3]. In addition to the catalytic Mo-bisPGD cofactor, the enzyme features a cubane-type iron sulfur center harbored by the large catalytic subunit and a Rieske-type [2Fe-2S] cluster ligated within the smaller subunit of the heterodimeric enzyme [4, 5]. The cubane-type cluster in Aio is a [3Fe-4S] cluster whereas this cofactor mostly corresponds to a [4Fe-4S] center in other members of the superfamily. The small subunit with its Rieske-type center is specific to Aio and not found in other families of Mo-bisPGD enzymes. While the catalytic molybdenum centers present in the majority of Mo-bisPGD enzymes commonly shuttle through two distinct redox transitions (Mo^VI/Mo^V and Mo^V/Mo^IV) featuring a paramagnetic Mo^V state observable by Electron Paramagnetic Resonance (EPR)[6-11], no such Mo^V EPR signals were observed in Aio. The report of n=2 behavior in Aio from Alcaligenes (A.) faecalis as observed by protein film voltammetry (PFV) [12] eventually proposed a rationale for the seemingly missing Mo^V EPR signal. Parts of the results from the initial PFV study were subsequently challenged in an independent study applying the same method to an Aio from a different organism, Rhizobium (R.) sp. Str. NT-26 [13].

In this work, we address the question of the Mo-cofactor’s redox behavior in Aio by redox titrations monitoring (a) the Mo^VI state by optical spectroscopy and (b) the (1-electron reduced) Mo^V state by EPR. No EPR redox titrations on Aio have been published so far and no optical titrations have yet been performed on any member of the superfamily. Our results
confirm the results obtained by Hoke et al. [12] and definitively show that the Mo-bisPGD center in Aio undergoes a positively cooperative (n=2) 2-electron transition with two protons strongly coupled to the redox event. In contrast to PFV, our experimental approach furthermore allowed for the determination of an upper limit for the stability constant (K_S) of the semi-reduced Mo^V state. Expressing the redox properties of Aio and of other Mo-bisPGD enzymes in terms of K_S of the Mo^V state permits a quantitative comparison of the Mo-bisPGD cofactors to quinone-based systems in a common formalistic framework.

The existence of both positive (in Aio) and negative redox cooperativity (in several other Mo-bisPGD enzymes) in the Mo-pterin cofactors’ redox titrations raises the question of the parameters steering the center into one or the other redox regime. To assess these parameters, we have produced and characterized site-directed variants of Aio targeting both the immediate ligand-sphere of the metal and the environment of the coordinating pterin. Only a mutation affecting the pterins was found to substantially stabilize an EPR-detectable Mo^V state and thus to shift Aio’s redox behavior from strongly positive towards more negative cooperativity.

2. Experimental Procedures

2.1. Bacterial strains, plasmid and growth conditions

The aioBA genes of Aio were cloned without the aioB Tat leader sequence into pPROEX-HTb (Invitrogen) and expressed in Escherichia (E.) coli DH5α growing aerobically as already described [5].

2.2. Site-directed mutagenesis
The primers used to create point mutations in the *aioA* gene are shown in Table S1. Variants were made using the Agilent Quick Change II XL site-directed mutagenesis kit according to manufacturer’s instructions as has been done previously [5]. Mutations were confirmed by sequencing both strands.

2.3. Proteins purification

The WT and variant Aio proteins from *R. sp. NT-26* and WT Aio from *A. faecalis* were heterologously expressed in *E. coli* and purified according to a protocol adapted from [5]. The 50 mM MES, 150 mM NaCl (pH 5.5) equilibration buffer of the Superdex 200 10/300 gel filtration column (GE Healthcare) was replaced by a 30 mM MES/30 mM Tricine/30 mM HEPES/30 mM AMPSO/300 mM NaCl, pH 6-9 mix buffer. The presence of 300 mM NaCl has been found to improve protein stability. The *E. coli* NarGH was expressed using the plasmid pNarGHHis6J, purified in one step by affinity chromatography as described previously [14] and finally recovered in 50 mM MOPS pH 7.6 buffer at 90 μM.

2.4. Enzyme Assay

Arsenite oxidase enzyme assays were done as described previously [5], using the artificial electron acceptor, 2,6-dichlorophenolindophenol (DCPIP) 200 μM combined with phenazine methosulfate (PMS) in 50 mM MES (pH 6) or using horse cytochrome *c* in 50 mM Tricine (pH 8).

2.5. Optical titrations
Optical equilibrium redox titrations were performed on purified enzyme obtained from a 2-liter culture and diluted in the mix buffer with pH-values adjusted to 6-9. Enzyme concentrations were approximately 40 µM. Each reductive/oxidative titration was performed twice with two separate preparations at each pH value. The titrations were performed at 13°C [15] using a Cary 5E UV/Vis spectrophotometer, under Argon atmosphere in the presence of the following redox mediators at 10 µM: Ferrocene, 1,4-p-benzoquinone, 2,5-dimethyl-p-benzoquinone, 2-hydroxy 1,2-naphthoquinone, 1,4-naphthoquinone. Titrations were carried out using sodium ascorbate for reduction, and potassium ferricyanide for oxidation. Samples were allowed to equilibrate for several minutes. The redox midpoint potential values of the Mo cofactor were determined by evaluating the change in absorbance at 695 nm after normalizing the spectra to zero at 800 nm to correct for baseline changes between individual spectra. Due to the comparatively low extinction coefficients (ε) of the MoV and the MoIV states, this normalization procedure only affects the ε695 of the MoVI state while leaving the Beer-Lambert dependency on MoVI concentration unaltered. The data were fitted to a Nernstian sigmoid with n = 2 or n = 1 transitions.

2.6. Electron Paramagnetic Resonance

EPR spectroscopy was performed on purified enzymes in the mix buffer (pH 6 or 7) with approximately 40 µM enzyme. During the equilibrium redox titration, the redox potential was poised at 10°C as described in Duval [16], in the presence of the following redox mediators at 100 µM: 1,4-p-benzoquinone, 2,5-dimethyl-p-benzoquinone, 2-hydroxy 1,2-naphthoquinone, 1,4-naphthoquinone. Titrations were carried out using ascorbate for reduction, and ferricyanide for oxidation. Samples were allowed to equilibrate for several minutes. EPR spectra were recorded on a Bruker ElexSys X-band spectrometer fitted with an Oxford
Instruments liquid-Helium cryostat and temperature control system. The EPR spectra of Aio (WT: 32 scans; Q726G variant: 396 scans) were measured at differing temperatures (12 K to 50 K), microwave powers (0.51 µW to 1 mW), and modulation amplitudes (0.4 mT to 1.0 mT) to optimize signal amplitudes of the assayed cofactors. The EPR spectrum (1 scan) of NarGH was recorded at 1 mW, 0.4 mT modulation amplitude and at 50K.

2.7. ESI/MS Analysis

All mutations were confirmed by mass spectrometry performed on purified enzymes. Analyses were performed on a MicroTOF-Q (Bruker) with an electrospray ionization source. Samples were desalted and concentrated in 20mM ammonium acetate buffer prior to analyses with Centricon Amicon with a cut off of 30kDa. Samples were diluted with CH$_3$CN/H$_2$O (1/1-v/v), 0.2% formic acid and were continuously infused at a flow rate of 3 µL/min. Mass spectra were recorded in the 50-7000 mass-to-charge (m/z) range. MS experiments were carried out with a capillary voltage set at 4.5 kV and an end-plate offset voltage at 500 V. The gas nebulizer (N$_2$) pressure was set at 0.4 bar and the dry gas flow (N$_2$) at 4 L/min at a temperature of 190 °C. Data were acquired in the positive mode and calibration was performed using a calibrating solution of ESI Tune Mix in CH$_3$CN/H$_2$O (95/5-v/v). The system was controlled with the software package MicrOTOF Control 2.2 and data were processed with DataAnalysis 3.4.

2.8. ICP/MS Analysis

Molybdenum concentrations were determined in all purified enzymes by ICP/MS. Prior to the analysis, samples were mineralized in a mixture containing 2/3 of nitric acid (65 % Purissime)
and 1/3 of hydrochloric acid (37%, Trace Select). Samples were diluted five-fold before ICP/MS analysis. The ICP-MS instrument was an ICAP Q (ThermoElectron), equipped with a collision cell. The calibration curve was obtained by dilution of a certified multi-element solution. Molybdenum concentrations were determined using Plasmalab software, at a mass of interest m/z=95.

3. Results

Two X-ray crystal structures of Aio from two members of the Proteobacteria A. faecalis and R. sp. str. NT-26 [4, 5], have been determined. Strong conservation of structure between both enzymes in particular in the vicinity of the Mo-center was observed [5]. However, when studied with respect to their electrochemical properties, these two enzymes were reported to differ substantially [12, 13]. In the A. faecalis Aio, the Mo-bisPGD center was found to display a strongly positive cooperative 2-electron redox transition with a midpoint potential slightly below +300 mV at pH 6 and a pH-dependence thereof indicating the strong coupling of two protons to the redox event [12] (represented by the dashed red line in Fig. 1). In contrast, the enzyme from R. sp. NT-26 was reported to feature a higher (by almost 100 mV) redox potential and a pH-dependence corresponding to only one proton per two electrons [13] (Fig. 1, dashed blue line). Our first goal therefore was to clarify these divergences.

3.1. Re-examination of divergent electrochemical data on the Mo-bisPGD cofactor in the Aio from A. faecalis and R. sp. NT-26
Rather than by the voltammetric method, redox changes of Mo centers traditionally are followed via the EPR signal of the 1-electron-reduced, paramagnetic Mo\textsuperscript{V} state. However, no Mo\textsuperscript{V} EPR signal has so far been detected in Aio [17], a fact which would find a straightforward rationalization in the cooperative 2-electron redox behavior proposed by Hoke et al. [12] implying a highly destabilized semi-reduced intermediate state. We therefore resorted to optical spectroscopy. The UV/Vis absorption spectra of the molybdenum cofactor in these Mo-bisPGD enzymes, however, are broad and feature low extinction coefficients. Optical redox titrations therefore require high sample concentrations and consequently are rarely performed. To the best of our knowledge, the DMSO reductase Dor from *Rhodobacter sphaeroides* was, prior to this work, the only Mo-bisPGD enzyme intensively studied by optical spectroscopy with the aim to establish the redox properties of the Mo-center [18]. However, even in Dor, optical spectroscopy was not used to directly monitor equilibrium redox titrations of the Mo center.

Fig. 2A shows oxidized-minus-reduced difference spectra measured on the *R. sp. NT*-26 enzyme in a range of ambient potentials. These spectra closely resemble that of the native enzyme from *A. faecalis* [17] (for the full wavelength range spectrum, see Fig. S1A). The Mo-center strongly contributes to the spectrum in the 600 to 800 nm range (as already shown for DMSO reductase [18]), with a broad peak at 695 nm (Fig. 2A) on which we evaluated the Mo cofactor’s $E_m$ values. In this spectral region, the absorbance of the two iron-sulfur centers is negligible. The recorded data closely correspond to an $n=2$ Nernst curve (Fig. 2B, blue trace) but cannot be explained by a single-electron $n=1$ transition (red curve) and an $E_{m,pH6}$ value of +240 ±10 mV was obtained. All titration waves in the pH range from 6 to 9 correspond to such 2-electron transitions (Fig. S1B), although the data obtained at pH 9 admittedly show a higher scatter than at other pH values due to progressive degradation of the sample. In this pH range, the difference spectra of the wild-type (WT) enzyme show no obvious contributions.
from a Mo\textsuperscript{V} state characterized by a prominent feature at 500/550 nm in the enzyme Dor [18].

No significant amount of Mo\textsuperscript{V} can be detected by EPR throughout the addressed pH range (see below and Figure 3). The pH dependence of the observed n=2 transitions (see also Table 1) has a uniform slope of -50 ±10 mV/pH unit over the assayed pH range (Fig. 1 our data points are indicated by blue squares and the deduced regression curve is shown as a continuous blue line), in line with the theoretical value of -56 mV per pH unit expected at 13 °C for a strongly proton-coupled electron transfer and an H\textsuperscript{+}/e\textsuperscript{-} ratio of 1.

As shown in Fig. 1, the E\textsubscript{m}-values and pH dependences thereof in Aio from R. sp. NT-26 (continuous blue line and blue squares, respectively) closely match the results obtained on the native enzyme from A. faecalis (dashed red line, [12]) whereas they differ substantially from those reported for the native enzyme from R. sp. NT-26 [13] (dashed blue line). Since we used the recombinant R. sp. NT-26 enzyme, it was necessary to assess whether the observed differences were a result of the heterologous expression system. We consequently performed the characterization of the expressed A. faecalis enzyme in our high-yield system. The values obtained from the redox titrations (Fig. S1C) are shown in Fig. 1 (orange triangles) and correspond well to the data by Hoke \textit{et al.} [12] measured on the native enzyme (dashed red line). The Mo centers in the WT Aios from A. faecalis and R. sp. NT-26 therefore behave similarly both with respect to redox potential and to pH dependence thereof. The divergent results reported in [13] (as illustrated by the dashed blue line in Fig. 1 lying substantially above all other data and featuring a different slope) could not be reproduced in our experiments. Overall it can be concluded that both systems undergo strongly proton-coupled n=2 redox transitions.

\subsection*{3.2. A quantitative measure of redox cooperativity in 2-electron transitions}
We therefore conclude that the 2-electron redox transition in Aio features strongly positive cooperativity, \textit{i.e.} the first reduction step renders the second one very oxidizing resulting in a simultaneous uptake of two electrons and \textit{vice versa} for the oxidizing direction. Intuitively, one might expect that, by virtue of electrostatic repulsion, the negative charge of the first electron must push the second reduction step towards lower redox potentials. This is indeed what is observed in many cases and what is referred to as “negative redox cooperativity”. Negative redox cooperativity characterizes the behavior of many multi-center redox proteins (e.g. see [19]) corresponding to electrostatic effects of the first redox event on the second one. Rare cases of positive cooperativity have also been reported and have been rationalized by redox-induced conformational changes [20]. The emblematic examples for positive redox cooperativity, however, are the 2-electron transitions of quinones in aqueous solutions. A quantitative description of redox cooperativity in general was developed by Michaelis [21]. A more general presentation of the mathematical description together with numerous examples from organic chemistry was provided by Clark [22]. An introduction to the conceptual framework of cooperative 2-electron redox chemistry is presented in the Supplemental Material. This formalism emphasizes that the regimes of positive and negative cooperativity actually form a continuum with a smooth transition between the two extremes. In the region of negative cooperativity the transition from the fully oxidized to the 1-electron reduced form (E₁) occurs at substantially more positive potentials than that of the subsequent transition to the fully reduced state (E₂) ([21, 22]). The individual 1-electron transitions can therefore be directly observed and their E₁ and E₂ values determined (see Fig. S2A). By contrast, in a redox reaction with strong positive cooperativity E₁ is much lower than E₂, resulting in the simultaneous uptake/loss of two electrons, and the titration wave will within experimental accuracy resemble a single n=2 Nernst curve (Fig. S2D). The latter case is precisely what we observe in the titration curve of the fully oxidized Mo^{VI} state in Aio (Fig.
2B). Fig. S2, however, also illustrates that for $\Delta E = E_1 - E_2$ in the vicinity of 0 (Figs. S2B and S2C), the theoretical titration curves of the fully oxidized state deviate from both the n=1 and the n=2 dependences in principle allowing experimental access to $\Delta E$. The scatter of our experimental data points (Fig. 2B), however, renders this kind of approach insufficient for $\Delta E$ value determinations prompting us to use EPR monitoring of the paramagnetic Mo$^V$ state to obtain at least limiting values for the stability constant $K_S$ of the semi-reduced state and hence $\Delta E = E_1 - E_2$ (which are related by $\log K_S = (E_1 - E_2) * F/RT$). Two distinct approaches allow the deduction of $K_S$ and $\Delta E$ from the titration curve of the semi-reduced Mo$^V$ state. The traditional method proceeds through the determination of the fractional population of this state which is related to $\Delta E$ via the dependence shown in Fig. 4B. This approach is complicated for the case of Mo-bisPGD enzymes by the fact that Mo-cofactor occupancy in these enzymes commonly doesn’t reach 100% and must thus be determined by independent methods. A different way to access the values of $K_S$ and $\Delta E$, discussed by Robertson et al. [23], exploits the width of the bell-shaped titration curve of the semi-reduced state. As shown in Fig. 4A, this width can be converted into $\Delta E$ for values of $\Delta E > -100$ mV. Since the width of the bell curve asymptotically tends towards roughly 68 mV for very negative $\Delta E$s, it becomes virtually independent of $\Delta E$ below about -100 mV. At higher values, however, measuring the width directly permits calculating $\Delta E$ and $K_S$ without having to resort to quantifications of total Mo and Mo$^V$. The latter method proved particularly powerful for the case of the Aio variants as detailed below.

3.3. Placing a limit on the $K_S$ value of the Mo$^V$ state in WT Aio

Equilibrium redox titrations (at pH 6) monitored by X-band EPR spectroscopy have been performed on samples of Aio. A tiny EPR spectrum attributable to a Mo$^V$ state was
observed (Fig. 3, black spectrum) and found to titrate at $E_m = +240 \pm 10 \text{ mV}$ (Fig. 4C, open squares). The observed changes on ambient potential were redox-reversible and therefore represent a genuine redox transition rather than degradation-induced phenomena. The observed titration behavior corresponds to the $\text{Mo}^V/\text{Mo}^IV$ transition of the cofactor. Since no decrease in signal size was observed while titrating the sample to more positive potentials (Fig. 4C, open squares) the $\text{Mo}^VI/\text{Mo}^V$ redox transition to the fully oxidized state must occur at higher potentials than were attainable in our equilibrium titrations using potassium ferricyanide as oxidant. The $n=1$ redox $\text{Mo}^V/\text{Mo}^IV$ transition observed at 240 mV by EPR may appear inconsistent with the optically determined 2-electron transition at 240 mV (measured on the $\text{Mo}^VI$ state) raising doubts whether the EPR- and optically monitored redox transitions correspond to the same electrochemical species. We therefore quantified the $\text{Mo}^V$ signal in the WT by double integration of the $\text{Mo}^V$ EPR spectrum.

Comparing this double integral to that obtained on the $\text{Mo}^V$ state of respiratory nitrate reductase (Nar, Fig. 3, orange curve) and correcting for experimental conditions and $\text{Mo}$ content in Aio (quantified at around 80% by ICP-MS), we find that the maximal $\text{Mo}^V$ signal attained during our EPR titrations of WT (Fig. 4C), corresponds to only 2 % of total $\text{Mo}$ present in the sample. According to the dependence shown in Fig. 4B, the population of the $\text{Mo}^V$ state in the maximum of the bell-curve of Fig. 4C should be close to 100 % of total cofactor of its harboring enzyme. We therefore conclude that a small fraction (2 %) of our sample features a very strongly stabilized intermediate redox state of the $\text{Mo}$-bis$\text{PGD}$ cofactor. Whether this fraction corresponds to a non-physiological state or an alternative configuration of the enzyme cannot be decided at present. Whatever the origin of this minor fraction, the overwhelming majority (98 %) of Moco strongly destabilizes the $\text{Mo}^V$ state. The 2% contribution of the negative cooperativity redox transition as seen in EPR is by far too small to be detectable in our optical titration experiments (Fig. 2B). Since no other signal
attributable to Mo\textsuperscript{V} was detected, the stabilisation of Mo\textsuperscript{V} in the majority of enzymes (98 %) in the redox transition with strong positive cooperativity must be much smaller than the observed 2 % of the minority population with negative cooperativity. Taking 1% as an upper limit yields $\Delta E$ values below -200 mV (Fig. 4B) and $K_S < 4 \times 10^4$. The degree of redox cooperativity in Aio can therefore be quantitatively expressed by these $\Delta E$ and $K_S$ values.

3.4. Molecular determinants tuning redox cooperativity in Mo-bisPGD enzymes

Redox cooperativity in quinones, the arguably most thoroughly studied class of 2-electron redox compounds \cite{22}, is generally considered to be mediated by the charge-compensating effect of protonation/deprotonation reactions \cite{24} and/or hydrogen-bonding interactions \cite{25} (see also our short introduction to the electrochemistry of 2-electron compounds in the Supplemental Material). It therefore is tempting to apply this paradigm also to Mo-bisPGD enzymes when searching for the parameters which steer the cofactor towards one redox regime or the other. We consequently looked for redox-coupled protonation/deprotonation events and/or redox-induced pK-changes as potentially cooperativity-tuning parameters. Fig. 5A shows a structure overlay of the ligand environment of the Mo-atom in Aio from \textit{R. sp. NT-26} to that of the Nar from \textit{E. coli} for which stabilized Mo\textsuperscript{V} states (at pH 8) have been reported \cite{26-29}. Two fundamentally distinct locations in the environment of the Mo-ion feature intriguing structural differences possibly related to charge-compensating effects.

(1) The direct ligand sphere of the Mo atom. While in all representatives of the superfamily, four coordination sites of the Mo-ion are occupied by sulfur atoms provided in pairs by each of the two pyranopterins, the 5th ligand to the Mo-atom is variable. It is an aspartate (Asp222) in Nar but can be serine or cysteine in other Mo-bisPGD enzymes whereas
the 6th coordination site appears to be reserved for the catalytic reaction, *i.e.* is occupied by an oxo-, hydroxo- or sulfur- group (for a recent review, see [30]). In the X-ray structures of Aio, however, the 5th coordination position on the Mo-ion is fully vacant (Fig. 5A). However, these structures have been obtained in the reduced state of the enzyme, which doesn’t rule out differing conformations while the Mo center is oxidized. EXAFS and Raman studies indeed suggested the presence of a distended oxo or a hydroxo group as the 5th ligand in the oxidized state [31], in addition to the canonical oxo-ligand present in several other members of the superfamily. The stoichiometry of 2 protons per 2 electrons in Aio’s redox transition prompted Hoke *et al.* [12] to favor the hypothesis that the additional oxygen ligand is indeed an oxo group and that reduction of the enzyme would entail double protonation of this oxo group, followed by dissociation of the produced water molecule. Such a reaction mechanism provides the essential ingredients for H+-linked destabilization of the intermediate redox state as in the case of quinones.

To test this hypothesis we have generated variants of Aio potentially providing a 5th ligand to the Mo-atom. Sequence alignments of Aio and Nar suggest Ala203 of the *R.* sp. NT-26 Aio as the residue corresponding to the ligating Asp222 of Nar [32]. We have therefore replaced A203 by Ser, Cys and Asp to mimic ligand permutations so far observed in the superfamily. The A203S and A203C variants showed enzymatic and electrochemical properties similar to those of the WT enzyme (see Table 1) whereas the A203D variant had no detectable activity, was highly unstable and showed significantly modified UV/Vis-spectroscopic properties of the Mo cofactor. The signal amplitude of the spectral contribution at 695 nm (Fig. S4) together with metal analysis results (5% Mo content quantified by ICP-MS) demonstrated that the Mo content of this variant was very low. We nevertheless were able to evaluate the redox properties of its residual Mo-center at pH 7 and determined an $E_m$
of + 140 mV (Table 1) and a positive cooperative 2-electron transition. No EPR signal attributable to Mo\textsuperscript{V} was detected in any of the A203 variants (Fig. 3).

The data obtained for these variants thus do not straightforwardly support a link between absence of a protein ligand to the Mo-atom and Aio's unique redox properties. However, the similarity of the Cys and Ser variants to the WT enzyme (we verified all the variants by ESI/MS) raises doubts as to whether the Ser and Cys mutations have actually introduced a 5\textsuperscript{th} ligand to the Mo-atom. The structural overlay of corresponding sequence stretches in Aio and Nar shown in Fig. 5A (grey for Nar and blue for Aio) highlights a substantially different fold in Aio of the whole stretch of amino acids between sequence positions 199 (end of β-sheet) and 209 (beginning of α-helix). This modified conformation moves the amino acid corresponding to the ligand in Nar away from the Mo-atom. It therefore isn’t obvious that the entire sequence stretch actually did restructure upon introduction of the potential Mo-ligand.

Concerning the A203D variant, two scenarios are conceivable. (a) As for the two other variants, the Asp residue remained too far from the Mo-center to become a ligand. The introduced negative charge positioned about 12 Å from the Mo-center induced an \(E_m\) downshift due to electrostatic interaction. (b) The Asp residue became the 5\textsuperscript{th} Mo-ligand but the far-reaching reorganization of the flanking chain resulted in instability of cofactor binding. The introduction of a 5\textsuperscript{th} ligand would then have severely affected spectral properties and \(E_m\), however without detectably shifting the redox transition towards the negative cooperativity regime.

Irrespective of whether a 5\textsuperscript{th} ligand has been introduced or not, it is worth noting that the scenario of a present/absent oxo-group at the 5\textsuperscript{th} ligating position as cooperativity-tuning parameter fails to provide a unifying mechanism for the redox behavior of the entire superfamily. As mentioned, Aio represents an extreme but not the only case of redox positive
cooperativity in this superfamily. Dor also does so but the 5\textsuperscript{th} coordination site of its Mo-atom isn’t vacant but occupied by an O atom from a Ser residue.

(2) The H-bonding network surrounding the pyranopterins: The four pyranopterin-sulfurs coordinating the Mo-atom (Fig. 5) are part of an extended conjugated system and electron density in the Mo-orbitals therefore may be influenced by even remote parts of the pterin moieties. Indeed, a role of the pyranopterins as "non-innocent" ligands has been increasingly discussed over recent years [33, 34]. In particular, the pyranopterins are embedded in an extensive H-bonding network provided by the ambient protein and are thus likely candidates for providing cooperativity-tuning charge compensation effects. We have therefore looked for inter-enzyme differences in the vicinity of the two pyranopterins. While the respective "outer" (i.e. pointing away from the Mo-center) protons on both pterins are H-bonded by backbone-amides in all structures of representatives from the superfamily, intriguing differences can be found with respect to the "inner" hydrogens (Fig. 5B). In the well-studied model system Nar, two prominent His residues have been proposed to engage in multiple H-bond interactions [33]. The so-called “bridging” His1092 provides a H-bond interconnection between the proximal (P) and the distal (D) pyranopterins while a “stabilizing” His1098 is considered to be crucial for fixing the P pyranopterin in its particular conformation [33]. The His1098 residue is indeed conserved in many members of the superfamily while His1092 is frequently replaced by an Arg (e.g. Arg720 in Aio, Fig. 5B) residue showing similar H-bond interactions. In the structural comparison of Aio and Nar shown in Fig. 5 as well as in comprehensive multiple sequence alignments of representatives of the superfamily [35], however, Aio stands out (together with an as yet uncharacterized enzyme from Desulfovibrio gigas and the acetylene hydratase from Pelobacter acetylenicus [36] by the presence of a glutamine residue (Gln726) in the position of the canonical stabilizing His (Fig. 5B). According to both available structures of Aio, the oxygen atom on
the Gln726 side-chain is a strong H-bond acceptor to the proton on the N₅ nitrogen of the P
pyranopterin (Fig. 5A). The almost singular presence of this particular amino acid in a
strategic position prompted us to assess its role in Aio’s redox chemistry through site-directed
mutagenesis. We therefore substituted Gln726 with a glycine residue which is unable to
engage in hydrogen bond interactions from its side-chain.

This variant was found to feature prominent EPR lines in the spectral region
characteristic for Mo⁵ centers as shown in Fig. 3 (red spectrum) in addition to much smaller
signals resembling those of the paramagnetic species already observed in the wild type
(detected at high ambient potentials where the strong EPR signal is absent; see below and
Table S2). The dominant spectrum is distinguishable from both that of the WT enzyme and
that of Nar. Its spectral features do not arise from the [3Fe-4S]cluster which shows no
measureable signal at 50K [17]. The Rieske [2Fe-2S] cluster, which indeed is still visible at
50K, is observed at lower redox potentials without contributions from the other centers and
was subtracted out of the red spectrum shown in Fig. 3. None of the two iron-sulfur centers
present in the enzyme thus contribute to this spectrum. Its saturation behavior was found to
correspond to that of typical Mo-bisPGD centers (data not shown). As detailed below, the
bell-shaped titration curve of this paramagnetic center resembles that of Mo⁵ states in other
members of the superfamily and the E₁ and E₂ values of the two redox transitions as obtained
by EPR are fully consistent with the 2-electron potential measured by our optical approach.

To obtain signal-to-noise ratios allowing for the identification of finer spectral structures, the
spectrum shown in this figure was extensively accumulated (396 times). The spectral features
indicated by asterisks in Fig. 3 most likely correspond to hyperfine lines arising from the
minor⁹⁵Mo- and ⁹⁷Mo-isotopes with nuclear spin I= 5/2. A literature survey suggests that the
spectrum of our variant (see Table S2 for g values) most closely resembles that reported for
Mo⁵ in the enzyme Fdh from Methanobacterium formicicum [6]. The ensemble of these
observations therefore demonstrates that the observed spectrum indeed corresponds to the Mo\textsuperscript{V} state in the variant enzyme.

The quality of the data points during the titration of the Mo\textsuperscript{V} signal in the Q726G variant (Fig. 4C) allows a reliable determination of the full width at half maximum (W\textsubscript{HH}) of this curve yielding a value of 93 mV (Fig. 4C) which translates into a $\Delta E$ of +36 mV and also to 45% stabilization of the Mo\textsuperscript{V} state, according to the dependences illustrated in Fig. 4AB. This indicates that the variant enzyme stabilizes the Mo\textsuperscript{V} state observable by EPR at cryogenic temperatures with $K_S = 4$ ($\log K_S = 0.6$) to the exception of the strongly stabilized fraction also observed in the WT. The Q726G mutation thus substantially stabilizes Mo\textsuperscript{V} as compared to the WT.

According to the theoretical titration curves (Fig. S2B), a Mo redox behavior with $\Delta E +36$ mV as observed by low temperature EPR should also give rise to deviations from $n=2$ behavior detectable in room temperature optical titrations of the Mo\textsuperscript{VI} state. Fig. 2C shows the result of such an optical titration on the Q726G variant (see inset for comparison of the spectrum recorded on Q726G with the one from the WT). While the data points in the variant (Fig. 2C) clearly show a shallower dependence of the signal amplitude on ambient redox potential than in the WT (Fig. 2B), they do not yet approach $n=1$ behavior as predicted from the simulations (Fig. S2B). Fitting the data to the equation given in Supplementary Material as formulated by Clark [22], yields a $\Delta E$ of -44 mV, \textit{i.e.} much higher than the limiting value of -200 mV found in the WT but indisputably lower than the EPR value of +36 mV. Previous results reported for Dor suggest a straightforward rationalization for this discrepancy. Bastian \textit{et al.} [7] have analyzed the redox behavior of the Mo\textsuperscript{V} state both at 298 K and at 168 K and have found appreciable but dissimilar temperature dependences for the two individual 1-electron transitions. While $E_1$ was observed to be constant within experimental precision, $E_2$ increased by about 60 mV when analyzed at cryogenic temperatures. In Aio, both transitions
appear to be temperature-dependent with $E_1$ increasing by about 40 mV and $E_2$ decreasing by the same amount when going to low temperatures. In the framework of the scenario that the Mo-cofactor's redox properties are controlled by the H-bonding network surrounding the pyranopterins as suggested by our mutagenesis results and as discussed in more detail below, differences in the effect of temperature on the individual 1-electron transitions find an explanation in differential modifications of the pK values on involved protonation sites. The pK values of numerous protonable/deprotonable groups are in fact temperature-dependent[37].

4. Discussion

4.1. On the complementarities of the optical and the EPR approaches

The data concerning the Q726G variant detailed above together with previous results on Dor demonstrate that both individual redox transitions can feature temperature-dependent redox potentials. This potentially influences the apparent overall 2-electron midpoint potential, the stability constant $K_S$ of the semi-reduced state or both these parameters. Obviously, catalytic turnover is always determined above 0 °C and electrochemical parameters determined at cryogenic temperatures may therefore be misleading in certain cases when correlated with enzyme activities. This fact adds to the potential of the optically monitored equilibrium redox titration approach. Not only will this approach always produce 2-electron $E_m$-values applicable to the conditions of enzyme assays but it allows, as shown above, to also determine the $E_1$ and $E_2$-values of the individual 1-electron transitions down to potential inversions $\Delta E$ of about -50 mV. Obtaining equivalent information from room
temperature titrations of the EPR detectable Mo$^\mathrm{V}$ state requires substantially higher enzyme concentrations which are not always attainable.

4.2. The H-bond environment of the pyranopterins controls the Mo-cofactor’s redox behavior

The comparison of the results obtained on the WT and the Q726G enzymes demonstrates that the electrochemical parameters of both redox transitions of the Mo-cofactor are strongly influenced by the H-bonding environment of the pyranopterins. Elimination of the H-bond relay provided by Gln726 in Aio affects both $E_1$ and $E_2$ and results in an increased stabilization of the Mo$^\mathrm{V}$ state, by four orders of magnitude. Remarkably, the results published by Wu et al. during the course of our work [38] show that similar phenomena occur in Nar. The substitution of His1098, structurally equivalent to the Gln726 residue in Aio, by an Ala also substantially increases the stability of the Mo$^\mathrm{V}$ state ($K_S$ from 28 for the WT to 1822 for the H1098A variant). Variants of other H-bonding His only led to marginal stabilization or even destabilized the Mo$^\mathrm{V}$ state [38]. The fully congruent results obtained on Nar and on Aio emphasize the preeminent importance of the H-bond interactions provided by the amino acid residues at this specific structural/sequence position in the protein and we predict that mutagenesis work on other members of the superfamily will reveal a corresponding influence of the H-bond environment of the pyranopterins in the control of the Mo-cofactor’s redox behavior.

4.3. The variability of redox cooperativity in the Mo-bisPGD cofactors resembles that of quinones
The relationship between $\Delta E = E_1 - E_2$ and $K_S$ in 2-electron redox compounds is most conveniently visualized by the type of plot shown in Fig. 6, *i.e.* the graphical representation of the relationship $\log K_S = (E_1 - E_2)*F/RT$, introduced by Mitchell as “the redox seesaw” [39]. In many Mo-bisPGD enzymes such as *Rhodobacter sphaeroides* f. sp. *denitrificans* Dimethylsulfoxide reductase Dms, *Rhodovulum sulfidophilum* Dimethylsulfide dehydrogenase Ddh, *E. coli* Nar and *M. formicicum* Formate dehydrogenase Fdh [6, 8, 10, 27], positive $\Delta E$ values in the range of +100 mV (violet arrows in Fig. 6) and even as high as 775 mV for periplasmic nitrate reductase Nap [9] (out of range in Fig. 6) have been determined. The redox properties of these enzymes thus clearly fall within the regime of negative cooperativity. However, not all of them do, for *E. coli* Nar [11] a $\Delta E$ as low as +40 mV has been reported at pH 7.6. As already mentioned, in Dor from *Rhodobacter sphaeroides*, the $E_1$ and $E_2$ values are inverted by almost -60 mV at pH 7 (corresponding to $\log K_S = -1$) [7]. In these cases the individual 1-electron redox transitions therefore show weakly, to substantially positive, redox cooperativity. The case of Aio extends the range of accessible 2-electron electrochemical behavior of the Mo-bisPGD cofactor far into the regime of positive cooperativity with $\log K_S$ below -3.

For comparison, the redox patterns of selected quinones as observed in biological systems (brown arrows) [40-43] are indicated in Fig. 6. This pattern ranges from the strongly negative cooperative behavior of the menaquinone in Nar [40] ($\log K_S \sim +2$) to that of the so-called Q$_o$-site quinone of bc$_1$ complexes [44, 45] ($\log K_S = -14$, out of range in Fig. 6). In the case of quinones, charge-compensating effects of protonation/deprotonation reactions [24] and/or hydrogen-bonding interactions [25] have been put forward to explain modulation of redox cooperativity. The Mo-bisPGD cofactors’ redox properties thus resemble those of quinones.
4.4. Repercussions on the role of Mo in early life

Species distribution analyses and phylogenetic reconstructions indicate a very deep ancestry of the Mo-bisPGD superfamily dating back to early life [35]. We have in the past speculated that the transition metal Mo may, during life’s inorganic infancy, have performed the positive cooperative (n=2) redox reactions crucial to bioenergetics [46, 47] which subsequently have been assumed by small organic molecules such as quinones and flavins [35, 48-50]. However, the finding that the redox cooperativity in Mo-enzymes is induced by the environment of the pyranopterin ligands suggests that respective protonatable groups were likely also present in the Mo-bearing minerals involved in the emergence of the earliest metabolic reactions. This observation favors as promising candidates the mixed and variable valence double layer oxyhydroxides such as hydrotalcite or green rust, the interlayers of which are readily protonated and deprotonated [51]. In this state they can contain various counter-ions including molybdates and thiomolybdates [52, 53]. Soluble mixed MoIV oxide and sulfide complexes could have been supplied to the interlayers from the alkaline hydrothermal fluid and alternately oxidized and reduced therein [54]. Whether there are circumstances in which oxidation and reduction of these complexes could have involved 2-electron redox behavior with positive cooperativity is not known but is ripe for experimentation.

Author Contributions

J.M.S. carried out the molecular biology, S.D. and B.S.C. carried out the biochemical and optical spectroscopy experiments, S.D., B.S.C. and S.G. carried out the EPR experiments.
D.L. carried out the ESI/MS analyses, F.C. carried out the ICP/MS analyses. S.D., B.S.C., S.G. and W.N. analyzed the data. All authors wrote the paper.

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Supplementary data

Supplementary data to this article can be found online at http:…
Legends

Figure 1. Values and pH dependences of the Mo cofactor’s redox potentials. Blue squares and the blue continuous line (representing an H⁺/e⁻ ratio of 1) represent the results obtained in the present work on the R. sp. NT-26 Aio. The blue dotted line indicates the data reported by Bernhardt and Santini [13] on the R. sp. NT-26 Aio, simulated with an H⁺/e⁻ ratio of 0.5. Orange triangles mark the results obtained in the present work on the heterologously expressed A. faecalis enzyme. The red dotted line corresponds to the data reported by Hoke et al. [12] on the native A. faecalis Aio.

Figure 2. Optical titration of the Mo cofactor in wild type Aio and the Q726G variant from R. NT-26.
A: Optical spectra recorded on the wild type enzyme in the region 600-800 nm, recorded at pH 6 during titrations. B: Dependence of signal amplitudes on ambient redox potential as evaluated at 695 nm and fitted with Nernstian sigmoids using $E_m =$ +240 mV and $n=2$ (blue) or $n=1$ (red) behavior. The figure summarizes data obtained in two consecutive cycles of reductive and oxidative titrations. The experiment has been repeated twice independently on different enzyme preparations. C: Evaluation of signal amplitudes at 705 nm recorded during redox titrations of the Q726G variant. Dashed blue and red lines correspond to $n=2$ and $n=1$ behavior, respectively, as in B, while the continuous black line results from a fit of the data points to the theoretical dependence of the Mo VI state towards ambient redox potential. The inset shows the comparison of the optical spectrum recorded on the WT enzyme (blue line) to that recorded on the Q726G variant (black line).

Figure 3. EPR spectra recorded on wild type and variant Aio from R. sp. NT-26 as well as on E. coli Nar. Approximately 45 μM enzyme were used for redox titrations at pH 6 (for WT, A203C and Q726G enzymes) or pH 7 (for the A203D variant). Spectra were recorded on samples poised at +240 mV at pH 6 in the case of WT, A203C and Q726G and at +140 mV at pH 7 in the case of A203D. In the spectrum recorded on the Q726G enzyme, the spectral features indicated by asterisks most likely correspond to hyperfine lines arising from the minor $^{95}\text{Mo}$- and $^{97}\text{Mo}$-isotopes with nuclear spin $I=\frac{5}{2}$. Spectra recorded on Aios are compared to the spectrum recorded on NarGH purified from E. coli and poised at +155 mV at pH 7.6. Numbers 1, 2 and 3 denote $g_{1,2,3}$ values associated with each of the signals. The chosen experimental conditions allow detecting the Mo
cofactor of Nar in the Mo$^V$ state. All spectra were recorded at a microwave frequency of 9.48 GHz, a microwave power of 1 mW, a temperature of 50K and a modulation amplitude of 0.4 mT.

**Figure 4.** Theoretical dependences of the full width at half maximum ($W_{HH}$) of the Mo$^V$-titration curve (A) and of the maximally observable Mo$^V$ signal (B) on the difference in individual 1-electron redox potentials ($\Delta E = E_1 - E_2$). C: Experimentally determined titration curves for the weak Mo$^V$ signal observed in the wild type enzyme (open squares) and the prominent Mo$^V$ spectrum of the Q726G variant (filled diamonds). For a detailed presentation of the equations describing 2-electron redox transitions, see the tutorial included in the Supplemental Material or visit our dedicated website at http://bip.cnrs-mrs.fr/bip09/2electron.html.

**Figure 5.** Structure comparison of Aio and Nar enzymes. A: Comparative juxtaposition of the 3D-structures of Aio from *R.* sp. NT-26 (in blue) and of Nar from *E. coli.* (in grey). Crucial amino acid residues, the two pyranopterins and protonatable positions on the pterins are highlighted. B: Comparison of the proximal and distal pyranopterins and crucial interacting amino acid residues in Nar (grey) and Aio (blue) as seen from "below" the Mo-bisPGD moiety.

**Figure 6.** "Redox-seesaw" representation of the dependence of $\Delta E = (E_1 - E_2)$ on the stability constant $K_S$ of the half-reduced state. Experimentally determined values for semiquinones (in brown) or Mo$^V$ (in violet) intermediates are represented. The red arrow stands for the values determined in this work for the Q726G variant of Aio.

**Table 1.** Properties of wild type and variant Aio enzymes from *R.* NT26 and *A. faecalis.* Except for the Q726G variant, the redox potential value represents the $E_m$ value of the 2-electron redox transitions. In the case of Q726G, $E_1$ and $E_2$ can be distinguished and are indicated.

**References**


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Graphical abstract
Fig. 1
Fig. 2
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Highlights

- The Mo-bisPGD enzyme arsenite oxidase displays strong redox cooperativity
- Optical titrations appear as a powerful method for assessing Mo-redox properties
- The H-bond network surrounding the pyranopterins-ligands modulates cooperativity
- The Mo-bisPGD cofactor resembles quinones with respect to redox properties