Reduced levels of S-nitrosothiols in plasma of patients with systemic sclerosis and Raynaud’s phenomenon

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Nitric oxide (PubChem: CID: 145068)
Sulfanilamide (PubChem: CID 5333)
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Dithiothreitol (PubChem: CID 19001)
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Objective: S-Nitrosothiols (RSNOs) are bioactive forms of nitric oxide which are involved in cell signalling and redox regulation of vascular function. Circulating S-nitrosothiols are predominantly in the form of S-nitrosoalbumin. In this study plasma concentrations of S-nitrosothiols were measured in patients with systemic sclerosis (SSc) where NO metabolism is known to be abnormal.

Patients and methods: Venous blood was collected from 16 patients with Raynaud’s phenomenon (RP), 45 with systemic sclerosis (SSc) (34 patients had limited SSc (lcSSc) and 11 diffuse cutaneous disease (dcSSc)). Twenty six healthy subjects were used as controls. Plasma S-nitrosothiol concentrations were measured by chemiluminescence. The measurements were related to the extent of biological age, capillary/skin scores and disease duration.

Results: Plasma RSNO levels in patients with Raynaud’s phenomenon (RP) and in those with SSc was significantly lower compared to the concentrations in control subjects. In SSc, plasma S-nitrosothiols were often below the level of detection (1nM).

Conclusions: Low S-nitrosothiol concentrations were observed in the blood of patients with SSc and patients with RP indicating a profound disturbance of nitric oxide metabolism.

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1. Introduction

Systemic sclerosis (SSc) is a rheumatic and connective tissue disorder that affects the skin and the internal organs (lungs, heart, kidney and gut) leading to tissue fibrosis. The majority of SSc patients exhibit Raynaud’s phenomenon (RP) [1], a vasodilatatory response to cold which induces ischemic attacks and vasospasm of the digits [2]. The pathophysiology of RP includes abnormalities of blood vessels, abnormal neural control mechanisms and platelet activation [3]. Patients with RP do not show clinical features of SSc. However, some cases may evolve to SSc and exhibit abnormal nail fold capillaries and develop autoantibodies that are typically observed in patients with SSc [4]. The clinical subgroups of SSc are classified into limited SSc (lcSSc) and diffuse cutaneous SSc (dcSSc), according to their degree and distribution of skin sclerosis, their immunological profile and extent of microvascular dysfunction [5,6].

Several studies have shown that patients with SSc exhibit many features suggestive of decreased levels of antioxidant vitamins and enzyme activities related to defence against oxidative stress [7]. The fibroblasts show increased release of reactive oxygen species [8] and exhibit stress in their vascular and immune systems and extracellular matrix, which may contribute to progressive fibrosis [9].

The formation of vasodilator nitric oxide (NO) in normal endothelial cells [10] has been the focus of study in the pathogenesis of SSc [11–13]. Decreased endothelial NO formation has been reported in SSc patients [14–16]. However, not all of the evidence points in the same direction [17]. In macrophages, NO produced by the inducible form of the enzyme NO synthase (iNOS) is a cytotoxic agent released as part of the primary defence mechanism against microorganisms. Pulmonary production of NO is increased as lung macrophages express higher amounts of iNOS in SSc patients [18]. Elevated levels of nitrite/nitrate have also been reported in serum of SSc patients [19–21]. The excess NO reacts with superoxide anions to produce peroxynitrite, an oxidant that nitrates tyrosine residues in proteins to form nitrotyrosine. The levels of nitrated proteins in the skin and plasma are increased, especially in patients with diffuse SSc at early stages of the disease [13].

S-nitrosothiols (RSNOs) are more stable forms of bioactive NO in vivo. RSNO formation occurs by several pathways including oxidation of thiols with peroxynitrite [22], or their reaction with N2O3 a product from oxygen and NO [23]. RSNO act as NO donors to specific thiol-regulatory
effector sites including enzymes and signalling proteins by trans-nitrosation [24]. RSNOs bind covalently to SH groups of proteins and low molecular weight thiols in vivo and signal through the modification of cysteine (CYS) residues by covalent addition of an NO moiety, (formally as NO\(^{-}\)), which regulates the function of a broad spectrum of proteins in cells by S-nitrosylation [25]. The reaction of NO with the free thiols of serum albumin, cysteine and, to a lesser extent, glutathione results in the formation of the respective S-nitrosothiols in vivo [26–28]. RSNOs are reduced to NO and nitrite when degraded [29].

We hypothesized that the circulating concentration of RSNOs in SSc would decrease at an early stage of the disease resulting in endothelial damage and impaired NO biosynthesis. These changes may contribute to both the presence of Raynaud’s phenomenon and also the fibrotic changes that ensue in SSc.

2. Materials and methods

2.1. Patients

Patients with RP and systemic sclerosis were enrolled in this study with the approval of the Royal Free Hospital Ethical Practices subcommittee. RP patients included are those with normal capillaroscopy (NC) and negative antinuclear–antibody (ANA) and those with NC or ANA or both, but without evidence of connective tissue disease. Out of 16 RP patients 6 patients had positive ANA and 4 had abnormal capillaroscopy and no patient with ANA and with capillaroscopy. The patients with SSc fulfilled the American College of Rheumatology (formerly the American Rheumatism Association) preliminary diagnostic criteria for SSc [30] and included patients with lcSSc and dcSSc.

The study was carried out with a total of 61 patients selected randomly and who attended the Rheumatology clinic at the Royal Free Hospital exhibiting either Raynaud’s phenomenon (RP) or SSc with subgroups of lcSSc and dcSSc. Both male and female patients were included and none of whom smoked. Twenty six healthy non-smoking subjects (male and female) were randomly selected and used as controls. Informed consent was obtained from all subjects enrolled in the study. Measurement of RSNO concentrations of patients was carried out blind and was compared to the skin thickness assessments for SSc patients measured within three months of the blood samples being taken. RSNO levels were also related to patient age, capillary/skin scores and duration of disease.

2.2. Bloods

Venous blood was taken from patients and healthy controls into pre-chilled tubes containing EDTA (ethylenediaminetetraacetic acid final concentration 2 mM) and NEM (N-ethylmaleimide final concentration 5 mM) and centrifuged for 10 min at 1300 g at 4 °C. NEM prevented RSNO degradation by transnitrosation reactions [31] through alkylation of thiol groups. RSNOs were determined on the day of sample collection.

2.3. Determination of plasma S-nitrosothiols

The assay is based upon the detection of nitric oxide released by copper, iodine–iodide mediated cleavage of RSNOs to form NO, with subsequent quantification by its gas phase chemiluminescence reaction with ozone [32]. 8 ml glacial acetic acid and 2 ml KI (50 mg/ml) was kept in a reaction chamber at 70 °C (custom designed to accommodate larger sample volume or identical in design to a commercially available glass chamber (Sievers Radical Purger™)). One minute prior to injection of the sample (200 μl), CuSO\(_4\) (200 mM) was added and the solution purged with N\(_2\) and used once per sample. The gas stream was passed into Sievers Instruments Model 280 Nitric Oxide Analyser (NOA™ Sievers, Boulder, Colorado, USA). Data collection and analysis were performed using the NO Analysis™ software (Sievers, Boulder, Colorado, USA). To remove the background nitrite in plasma samples, sulfanilamide 0.5% (w/v) in HCl (final concentration 0.2 M) was added. This forms a stable diazoniun salt and prevents NO formation. Antifoam™ (Sievers, Boulder, Colorado, USA) in 1:30 dilution in deionised water was added to the plasma at a ratio of 1:9, immediately before injection of the sample. 100–1000 μl plasma extracts were injected to the reaction chamber, before and after addition of CuSO\(_4\) in order to calculate the level of S-nitrosothiol and exclude N-nitrosamine or iron nitrosyl. The NO\(_2\) measurements were taken by chemiluminescence before the addition of CuSO\(_4\), whereas concentrations of S-nitrosothiols were determined after the addition of CuSO\(_4\) to the reaction chamber with sulfanilamide. RSNO concentrations were calculated by subtraction of the value of NO\(_2\) from the value received after injection in CuSO\(_4\).

2.4. Preparation of standard (SNO-alb)

Standard curves for SNO-albumin (SNO-alb) were obtained over the range of 1nM to 5 μM by addition of known amounts of SNO-alb to PBS with 5 mM NEM. To prepare SNO-albumin human serum albumin (20 mg/ml) was treated with dithiothreitol in phosphate buffer saline (PBS) together with 100 μM diethylenetriaminepenta-acetic acid (DTPA) and gently stirred at 20 °C for 2 h to reduce the Cys-34 thiol group. The sample was dialysed extensively in Visking tubing ((Medicell, London, UK) molecular weight cut off 14,000) at 4 °C against 3 × 3 L PBS, pH 7.4 containing 100 μM DTPA. Any untreated thiol was alkylated with NEM (1 mM) at room temperature and dialysed against PBS/DTPA. The stock solution of SNO-alb was stored at −20 °C. SNO-alb was measured prior to use by the Saville reaction [33].

2.5. Skin scores

Tightness and thickening of skin were measured in the majority of SSc patient using the modified Rodnan skin score method [34]. The total skin surface area was divided into 17 different areas: the skin score evaluated in each by manual palpation. The skin score grading was 0 for uninvolved skin, 1 for mild thickening, 2 for moderate thickening, and 3 for severe thickening (hidebound skin). The total skin score was the sum of the skin scores of the individual areas, the maximum possible score being 51. The skin scores correlate with the extent of dermal fibrosis and the extent of fibrosis and dysfunction of internal organs.

2.6. Nailfold Capillary scores (NC) were measured as described in Ref. [35]

2.6.1. Data analysis

Data were expressed as mean ± SE where \( n \) was the number of individual samples assayed in duplicate or more. Comparisons of multiple means were performed using a one way ANOVA followed by the Student’s Newman–Keuls test. Correlation and regression were performed using the ‘least squares’ method in the Microsoft Excel programme. \( P \) value of <0.05 was considered as statistically significant.

3. Results

3.1. Demographic data of patients and controls

The characteristics of controls and patients are described in Table 1. The age range was similar in each group but the controls had a higher proportion of males.

3.2. Plasma RSNOs in healthy controls: effects of age

The concentrations of RSNOs in healthy controls were relatively low compared to those reported using other methods, and rather below those measured for controls by the originators of this procedure [32]. However the mean age of this cohort is likely to have been higher and reflects a population with a wide age range which seems to influence
RSNO concentration negatively as age increases ($r^2 = 0.22$; correlation; $P = 2.67 \times 10^{-8}$, $n = 26$) but the relationship was not linear.

At ages below 36 in the controls RSNO levels appeared to be higher (6.36 ± 1.1 nM; $n = 11$) and correlated negatively to age ($r^2 = 0.34$; $P = 0.015$); whereas as age increased to 60, RSNO values were lower but not between each other ($r^2 = 0.08$, $P = 0.12$). The low plasma levels of ascorbic acid in SSc may also contribute to a reduction in NO biosynthesis or by reducing thiol availability [46]. Other mechanisms by activation of matrix metalloproteinase-1 [45]. The inhibition of these activities may also contribute to the fall in RSNO levels in patients with SSc and RP, but this appears not to be the only factor, since many patients have RSNOs below the measurable threshold. These very low values were not found in any of the controls. The low levels of RSNOs are consistent with previous studies showing severe disturbances in nitric oxide metabolism in these diseases e.g. increased nitrated proteins and ADMA [13] especially in diffuse SSc.

The reduction of RSNOs in patients with IRP is less consistent, but accords with reports that show disturbances in NO metabolism in these patients [13]. In contrast, in SSc, NO release may be increased transiently because of the induction of NOSII [12].

In these studies we found that RSNO levels were higher in the healthy controls compared to those over the age of thirty six, especially if they were young. The finding is consistent with earlier reports that NO biosynthesis in endothelial cells was reduced with age [42,43] an effect associated with increasing apoptosis [44]. Age may contribute to the low levels of RSNO in patients with SSc and RP, but this appears not to be the only factor, since many patients have RSNOs below the measurable threshold. These very low values were not found in any of the controls. The low levels of RSNOs are consistent with previous studies showing severe disturbances in nitric oxide metabolism in these diseases e.g. increased nitrated proteins and ADMA [13] especially in diffuse SSc.

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The reduction in the level of RSNOs in SSc may track a gradual deterioration of endothelial function post the inflammatory phase of the disease, but low levels were also found in several of the RP patients suggesting that the fall may occur at an early stage in some cases. The decline of RSNOs was also parallel to increasing skin score, which highlights earlier observations where nitric oxide inhibits collagen secretion in human dermal fibroblasts through a cGMP-independent regulatory mechanism by activation of matrix metalloproteinase-1 [45]. The inhibition was less effective in SSc fibroblasts. However these correlations should be regarded with caution as there were many patients who had undetectable levels of RSNO.

The low plasma levels of ascorbic acid in SSc may also contribute to a reduction in NO biosynthesis or by reducing thiol availability [46]. Recent work suggests that S-nitrosothiols and products of S-nitrosylation play key roles in human health and disease.

An understanding of the factors that determine RSNO levels in vivo and the reasons why aberrant depletion of intracellular RSNOs contributes to disease are beginning to emerge [47].
5. Conclusion

The findings in this study strengthen the view that NO metabolism is profoundly disturbed in SSC and that low levels of RSNOs may play a key role in its progress. This may influence not only the universal existence of RP in SSC patients, but also abnormalities in platelet activation, the regulation of collagen activity and defective angiogenesis. The evaluation and study of the concentration of RSNOs in plasma of patients in RP and SSC and correlation with some clinical markers may help to improve safe treatment with NO donor drugs.

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References