Abstract: Indocyanine green (ICG) fluorescence imaging is a surgical tool with increasing applications in colorectal surgery. This tool has received acceptance in various surgical disciplines as a potential method to enhance surgical field visualization, improve lymph node retrieval, and decrease anastomotic leaks. In colorectal surgery specifically, small studies have shown intraoperative fluorescence imaging is a safe and feasible method to assess anastomotic perfusion, and its use may impact anastomotic leak rates. Controlled trials are underway to validate these conclusions. In the meantime, new indications for ICG continue to develop, including innovative options for detecting and guiding management of colorectal metastasis to the liver. These advances could offer great value for surgeons and patients, by improving the accuracy and outcomes of oncologic resections. The purpose of this article is to review the literature on the current state of immunofluorescence in colon and rectal surgery and emerging applications.
Indocyanine Green Fluorescence Imaging in Colorectal Surgery: Reviewing the Current Literature, Applications, and Future Direction

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ABSTRACT

Indocyanine green (ICG) fluorescence imaging is a surgical tool with increasing applications in colorectal surgery. This tool has received acceptance in various surgical disciplines as a potential method to enhance surgical field visualization, improve lymph node retrieval, and decrease anastomotic leaks. In colorectal surgery specifically, small studies have shown intraoperative fluorescence imaging is a safe and feasible method to assess anastomotic perfusion, and its use may impact anastomotic leak rates. Controlled trials are underway to validate these conclusions. In the meantime, new indications for ICG continue to develop, including innovative options for detecting and guiding management of colorectal metastasis to the liver. These advances could offer great value for surgeons and patients, by improving the accuracy and outcomes of oncologic resections. The purpose of this article is to review the literature on the current state of immunofluorescence in colon and rectal surgery and emerging applications.

KEY WORDS

Immunofluorescence, indocyanine green (ICG); fluorescence angiography; fluorescence imaging; colorectal surgery; colorectal cancer
Introduction to Fluorescence Imaging

Fluorescence imaging (FI) is emerging as a major contributor to intraoperative decision making during surgical procedures. With FI, the tissue of interest is illuminated with light at an excitation wavelength (750-800 nm) while observing it emit fluorescence at longer emission wavelengths (over 800 nm) (1). Indocyanine green (ICG) is the fluorophore most commonly used in fluorescence imaging. ICG is a water-soluble, tricarbocyanine dye that binds to blood lipoproteins after intravenous injection and remains confined in the intravascular compartment until elimination. It is selectively taken up at the first pass by hepatocytes and excreted unchanged into the bile. This fluorophore has tissue penetration up to 5mm, a plasma half-life of 3–5 min, with biliary excretion in 15–20 min, making it ideal for repeated applications (2). ICG has been used in other clinical applications, such as determining cardiac output, hepatic function, liver blood flow, and ophthalmic angiography for several decades (1, 3, 4). Fluorescence imaging with the application of ICG is an area of new development, and the uses continue to grow in colorectal surgery. The long-standing safety and efficacy of ICG greatly facilitates its introduction to new applications in this field (1). To date, the most publicized application in colorectal surgery has been as fluorescence angiography for perfusion analysis of the colorectal anastomosis (5). However, the indications continue to expand, and these advances could offer great value for surgeons and patients, by improving the visualization, accuracy and outcomes of colorectal resections. The purpose of this article is to review the literature on the current state of immunofluorescence in colon and rectal surgery and emerging applications.
Search strategy and selection criteria

For this narrative review, a team of surgeons that are subject matter experts in fluorescence angiography for gastrointestinal and hepatic surgery met to determine the clinically relevant areas and applications of fluorescence imaging with ICG in colorectal surgery. The consensus was: review of the available equipment, fluorescence angiography for anastomotic assessment, ureter visualization, endoscopic tattooing of colorectal lesions, lymphatic and sentinel lymph node mapping, and colorectal liver metastasis. Within each defined area, searches using the PubMed and Medline electronic databases were performed from database inception to May 2017 for original articles on the use if ICG in the setting of clinical studies in colorectal surgery. The following search terms were used: “fluorescence imaging”, “fluorescence angiography”, “Indocyanine green”, and “ICG” with “colorectal”, “colorectal surgery”, “perfusion”, “lymph node”, “sentinel lymph node”, “colorectal cancer”, “anastomotic leak”, “ureter” “hepatocellular cancer”, and “liver metastasis”, and “fluorophores” AND “imaging” AND “surgery”. Reference lists and specific authors who the team recognized as other subject matter experts in the field were also hand searched. Articles were included if published in English and full text was available. Conference proceedings and videos were not included. Abstracts were initially independently reviewed by 1 author (DK) for relevance to the defined sections and novelty. Then, full text for the selected articles was reviewed by all authors, minus the section on metastatic liver lesions, which was performed by TI. The content was written in a narrative format for full breadth of the details on the technical aspects, current state, and future application of the field.
Fluorescence Angiography: The Technical Details

There are a few options for fluorescent angiography systems for laparoscopic and open surgery, including the Stryker 1588 AIM Platform (Portage, Michigan, USA), PINPOINT™ (Novadaq, Mississauga, Ontario, Canada), the D-Light NIR/ICG™ (Karl Storz, Tuttlingen, Germany), IC-View® (Pulsion Medical Systems, Munich, Germany), PDE-neo System™ (Hamamatsu Photonics K.K., Hamamatsu, Japan), the SPY Elite™ Kit (LifeCell Corporation, Bridgewater, New Jersey, USA), and da Vinci robotic system-Firefly™ (Intuitive Surgical Inc., Sunnyvale, California, USA). These systems function as a conventional laparoscope in white light mode, but can be activated into NIR mode, where the ICG is visualized as white fluorescence on a black background. The Novadaq system offers an additional PINPOINT dual display mode, where the green fluorescence is superimposed over the white-light image, providing a highlighted view of the tissue of interest (6). Surgeons should understand that the signal detectability in fluorescence imaging differs greatly among laparoscopic imaging systems and, in general, is inferior to open imaging systems (7). Studies have demonstrated the feasibility and safety of intraoperative fluorescence angiography using these systems, with no adverse effects related to the infusion of ICG (2, 5, 8–18).

Fluorescence Angiography to Assess Anastomotic Perfusion

Anastomotic leaks are a dreaded complication in gastrointestinal surgery. Despite technical advances in colorectal surgery, the rate of anastomotic leaks has been steady, reported in 3%-20% of colorectal cases (19, 20). The etiology of anastomotic leaks is
multifactorial, and their impact is widespread- adding a significant clinical and economic burden to the patient and healthcare system, as well as a predisposition to local cancer recurrence (20–26). The current diagnostic tests- including intraoperative endoscopic assessment, air leak testing, assessment of anastomotic donuts, measurement of local tissue oxygenation, CT scan, and water-soluble contrast enema - often fail to establish an anastomotic leak at stage early enough to allow timely intervention and minimize morbidity and mortality (24). And while a proximal diverting stoma can minimize the consequences of a leak, it does not reduce the risk (27). Thus, further study and new technology are warranted to address the issue of anastomotic leaks.

Perfusion is vital for healing, and inadequate blood flow can result in failure of anastomotic healing and leak (28). Adequate perfusion of the anastomosis is commonly confirmed by assessing palpable pulses in the mesentery, lack of bowel discoloration, and pulsatile bleeding from its cut ends; however, this method of assessment can be subjective and difficult to quantify.

A valuable tool to visualize perfusion of the bowel anastomosis is fluorescence angiography. Fluorescence angiography can help confirm anastomotic perfusion by visualizing the bowel perfusion intraoperatively in real-time. For detection of blood flow, 2–3 mL of ICG solution (2.5 mg/mL) is injected intravenously during the operation by the anesthetist. After intravenous injection, ICG is visualized as green when excited by light in the near-infrared (NIR) spectrum with a NIR camera in 30-
60 seconds (29). This assessment allows the surgeon to confirm or revise a proposed resection margin after ligation of the vascular pedicle and before the anastomosis is created (5, 8). This interrogation of perfusion before the anastomosis is especially relevant in non-anatomic resections, where aberrant or altered vascular anatomy can impair perfusion to the remaining colon (FIGURE 1). In addition, the surgeon can verify perfusion of the completed anastomosis with an endoscopic mucosal angiography evaluation of the join (9).

There is a growing body of literature supporting intraoperative assessment of perfusion impacting intraoperative management and patient outcomes, including anastomotic leak rate in colorectal surgery. Many studies evaluating immunofluorescence in colorectal anastomosis are retrospective case series in both laparoscopic and robotic colorectal resections (2, 5, 8, 10–18). These studies describe the technique as simple, taking approximately 5 minutes to perform whether laparoscopic or robotic (2, 18), and providing useful intraoperative information about the vascular perfusion during minimally invasive colorectal surgery, with promise to reduce anastomotic leak rates. Boni et al reported results of 107 laparoscopic colorectal resections (40 right colectomies, 10 splenic flexure segmental resections, 35 left colectomies, and 22 anterior resections), where fluorescence demonstrated an insufficiently perfused bowel margin in 4/107 patients, which was revised before anastomosis; none of these patients had a clinical leak (10). Gröne et al reported outcomes for consecutive rectal cancer patients who underwent laparoscopic anterior and lower anterior resection with fluorescence angiography over a 5-months period, finding perfusion imaging influenced surgical decision making in 28%
of the patients; there was 1 anastomotic leak (6%, coloanal anastomosis) in all patients during that period (11). Hellan et al. reported the outcome of perfusion assessment in 40 patients having robotic left-sided colon or rectal resection, where angiography led to a change in the proposed transection line in 40% of cases; anastomotic leak occurred in 2 patients whose site of transection had been revised (days 15 and 40) (18). Jafari et al evaluated 16 robotic low anterior resections with ICG perfusion, where the use of fluorescence angiography resulted in revision of the proximal transection point before formation of the anastomosis in 19% of patients; only 1 anastomotic leak occurred in the fluorescence angiography group compared to 4 in a matched control group (8). Ris et al reported outcomes in 30 consecutive elective minimally invasive colorectal resections (24 left-sided and 6 right-sided resections), where in addition to achieving no postoperative anastomotic leaks, the authors reported that visualizing the perfusion also encouraged avoidance of defunctioning stomas in three patients with low anastomoses (2).

With increasing utilization of the tool, the quality and volume of the evidence for fluorescence angiography in anastomotic assessment continues to grow. Boni et al completed a case-matched study, comparing 42 fluorescent angiography patients to 38 historic controls undergoing laparoscopic low anterior resection. From fluorescent angiography-demonstrated hypoperfusion, the planned anastomotic level was revised in 4.7%. There were 2 anastomotic leaks in the control group, and none in the fluorescent angiography group (30). While the sample size is too small to power conclusions, they felt it could lead to a reduction in the anastomotic leakage after total mesorectal excisions. Kudszus et al performed a case-matched study in 402 colorectal cancer
patients, with a subgroup analysis by elective/emergent case status and age, for the rate of anastomotic revision for leakage. They found an overall reduction in the absolute revision rate of the anastomosis of 4% in with fluorescence angiography, with significantly reduced rate of revision in the subgroup analysis of patients undergoing elective resections (3.1% vs. 7.7%, p = 0.04, risk of revision reduced by 60%) and in patients older than 70 years (4.3% vs. 11.9%; p = 0.04, risk of revision reduced by 64%), supporting that ICG fluorescence may significantly reduce the rate of severe complications in colorectal surgery (31). Kin et al also performed a case-matched study in colectomy and proctectomy patient, but concluded that the technology was not associated with a reduced colorectal anastomotic leak (32). Five percent of patients in the fluorescent angiography group had a revision of the planned resection margin. In the 173 pairs, they found 7.5% of the fluorescent angiography group developed an anastomotic leak versus 6.4% of controls. Multivariate analysis demonstrated that level of anastomosis and surgeon was associated with leaks (32). There were several issues with this study’s design that could account for the divergent results found in this work compared to other studies. The study was retrospective, with a historic control group used for matching. There was also a small overall sample size, and even smaller number of anastomotic leaks, (n=24), resulting in an underpowered analysis. With the lack of controlled studies and conflicting data, additional studies are necessary to determine whether this technology is beneficial for colorectal surgery. A recent systematic review of 10 cohort studies (916 patient) reported that intraoperative fluorescent angiography was associated with a reduced risk of anastomotic leakage (n = 23/693; 3.3 % (95 % CI 1.97-4.63 %) compared with no fluorescent angiography
assessment (n = 19/223; 8.5%; 95% CI 4.8-12.2%) (33). In the PILLAR-II prospective, multicenter, open-label, clinical trial performed at 11 centers across the United States, 139 patients undergoing left-sided colectomy and anterior resection had perfusion assessment (5). Fluorescence angiography changed the operative plan in 8% of patients (n=11). The overall anastomotic leak rate was 1.4%, and there were no anastomotic leaks in the 11 patients who had a change in operative plan. While the existing body of literature is promising, we await randomized data to define the role for fluorescence angiography in routine clinical practice. The PILLAR III multi-center randomized controlled trial is currently underway, and we anxiously await the results (ClinicalTrials.gov identifier: NCT02205307) (34).

One limitation that deserves mention in the currently available NIR imaging systems is the subjective evaluation of fluorescence intensity and lack of quantitative data to measure tissue perfusion—leaving surgeons with the question, “How green is green?”

Nerup et al aimed to establish a method to quantify perfusion in ICG fluorescence angiography in an animal model. After marking regions of interest in the stomach, the authors administered ICG and neutron-activated microspheres, performed NIR fluorescence, and then sampled the marked tissue to calculate the regional blood flow (35). They found a strong correlation between regional blood flow and the slope of the fluorescence curves, with acceptable correlation to predictive algorithms they developed. While the models show quantification of perfusion is feasible, the clinical applications of this theoretical work in animal models is pending (35). In 2 separate single-institution studies in human subjects, the absolute perfusion units were reported
as a recommendation of when to resect; the threshold value for absolute perfusion units in these reports were 18-20 (14, 36). Wada et al aimed to quantify assessment of intestinal perfusion by reviewing recorded video images and creating a time–fluorescence intensity curve at the point of bowel transection (37). They found that the fluorescence difference between maximum and baseline was the most indicative factor for anastomotic leak, with a sensitivity of 100% and a specificity of 92.5%. Further, no correlation between anastomotic leak and the time from ICG injection to the first visible fluorescence signal was found (37). While these studies are promising, controlled trials with larger sample sizes are needed to standardize and validate the quantitative models. In the meantime, the adequacy of anastomotic perfusion remains subjective.

**Emerging Uses of Fluorescence Angiography in Colorectal Surgery**

While developed to assess perfusion, various other uses of fluorescence angiography are being reported, which compliment the standard of care in colorectal surgery, and could facilitate improving patient outcomes.

**Ureter Visualization**

Iatrogenic ureteral injury is a relatively rare but devastating complication in colorectal surgery, with significant morbidity (38, 39). While rare overall, a higher incidence is reported with laparoscopic colorectal surgery, especially in rectal cancer resections (40, 41). Preoperative ureteral stenting is an option, but it has not been shown to decrease the risk of ureteral injury- only helping with early identification of ureteral injuries (42). In addition, there are complications and costs associated with placing stents, including
increased operative time and an increase in length of stay (42, 43). ICG fluorescence of the ureters has value for facilitating ureter identification, especially in minimally invasive colorectal surgery (FIGURE 2). The ICG binds proteins on urothelial layer on the ureter, reversibly staining the inner lining of the ureter, and allowing visualization of the green-illuminated ureter under NIR fluorescence (44). As ICG fluorescence penetrates tissue, the intraureteral ICG may be used to localize the entire course of the ureters, even when obscured in fibrotic tissue. While the intraureteral ICG cannot detect thermal or devascularization injuries, full-thickness ureteral injuries can be identified cause readily apparent leakage of ICG into the surgical field (FIGURE 3). Studies have described the technique, feasibility, and utility for visual identification of the ureters during laparoscopic (45–47) and robotic surgery using the Firefly® system (44, 48–50). The protocols described between 0.125-1mg/kg of methylene blue (45, 47) and 10-25mg of ICG dissolved in 10-mL of sterile water and injected through the open catheter, either as a bolus or through slow infusion over 5 minutes (44, 48–50). Maximum fluorescence was reported between 9 and 20 minutes from administration (45). The reported results describe bilateral ureters fluorescing green in 50-91% when methylene blue was used, and 100% of patients with ICG (44, 45, 47–50). The fluorescence persisted through the duration of the operative procedure (mean operative times 121-171.3 minutes), despite some variation in brightness from the depth of the ureter to the peritoneal surface across patients (44, 48–50). No intraoperative or postoperative adverse effects attributable to fluorophore administration were reported (44, 47–49). Further study will determine the role for this useful adjunct to prevent iatrogenic ureteral injury during pelvic dissection in colorectal surgery.
Endoscopic Tattooing Of Colorectal Lesions

Preoperative endoscopic tattooing of colorectal neoplasms facilitates localization at the time of surgery, and is crucial for appropriate resection and treatment planning (51). Tattooing has become especially important in the era of minimally invasive surgery, with the lack of tactile feedback afforded during resection (52). The correct measurement and appropriate margins facilitated by precise localization could directly impact recurrence and survival. India ink has been the standard marking agent, but complications have been reported, including perforation, peritonitis, abscesses, inflammatory pseudotumors, and adhesion-related ileus (53–55). ICG has been reported as the endoscopic tattoo agent in several studies to guide colorectal resections (56–61). In these reports, peritumoral injection before open or minimally invasive resection using NIR fluorescence to identify the lesion has been reported to be safe, feasible, and effective. No adverse effects have been reported and all colorectal lesions were visualized upon open or laparoscopic resection. The median time for injection of ICG has been reported as 4 days prior to resection, but the ICG has been described as visible up to 7 days after endoscopic injection; after 9-10 days, the agent was visible in only 20% of patients (58, 60). A comparative study was performed by Nagata et al, who endoscopically injected 24 patients with both ICG and India ink injected at separate sites within 4 days of laparoscopic colorectal surgery (59). They found the NIR fluorescence showed tumor location clearly and accurately in 100% of patients, while 10 of 24 patients had negative staining of India ink. The site of ICG injection did not reveal fibrosis, inflammation, necrosis or microabscesses, while the India Ink injection sites had vasculitis, necrosis,
edema, and neutrophilic infiltration of the bowel wall. While large scale, controlled trials are needed to define the site, concentration, and timing, from the benefits reported, ICG has value as a preoperative marking agent to identify tumor location.

**Lymphatic Mapping and Sentinel Lymph Node Identification**

In oncologic resections, sentinel lymph node mapping allows targeted identification and harvesting of potentially metastatic lymph nodes (62). Taking a clue from breast and gynecologic malignancies (63, 64), ICG is emerging as a powerful tool for lymphatic mapping, identification of micrometastatic disease, and focused target nodal assessment in colorectal cancer. **There is the potential that the NIR information could change the operative course and recommendations for adjuvant therapy postoperatively.** A few studies have reported the ICG immunofluorescence provided valuable information for finding lymph nodes; if these nodes are present outside of the traditional resection field, the operative strategy could be changed from this information (61, 65–68). **It was noted that ICG may be may be less reliable due to neoadjuvant therapy in rectal cancers** (61). Near-infrared fluorescence mapping of the pelvic side-wall in low rectal cancer can also guide the need for extended lymphadenectomy in appropriate patients, **and spare patients with tumor-negative sentinel nodes from the morbidity of a lateral pelvic sidewall node dissection** (67, 69). Intraoperative lymphatic mapping may also help define the lymphatic course for tumors that have variable drainage or in reoperative cases, where lymphatic-bearing tissue has been excised (56). The sentinel lymph node biopsy could have specific utility in transanal resections of rectal cancer, where identification could change the treatment course to a more radical resection. Arezzo et al performed a submucosal ICG injection in 3 patients, then used NIR fluorescence to map
the mesorectal lymphatics and guide full-thickness resection with any involved lymph nodes (70). In all 3 patients, the pathologist confirmed presence of excised nodes, lack of metastasis, and the patients were spared radical surgery. Published studies have described the accuracy and benefit with this tool. Cahill et al used ICG with intraoperative fluorescence for real-time identification of the lymphatic drainage and sentinel mesocolic lymph nodes in 18 patients undergoing laparoscopic surgery for colorectal cancer (56). They found the fluorescence helped identify the sentinel nodes—whether within the specimen or outside of the standard resection field—prior to formal dissection, and was valid to guide their resection. In their series, four of 18 patients had mesocolic sentinel lymph nodes outside of the traditional resection field identified with ICG immunofluorescence. Hirche et al evaluated 26 patients with colon cancer with ICG, then visualized their lymphatic mapping and sentinel lymph nodes with immunofluorescence, comparing results to the histopathology for clinical feasibility, detection rate, and sensitivity (66). The found the ICG could have improved accuracy over conventional methods. Fluorescence imaging identified the sentinel lymph node in 25/26 patients (detection rate, 96%); metastatic involvement was found in 9/11 nodal positive patients by conventional histopathology, a sensitivity of 82% (66). To date, the rate of lymph node detection has been described from 89% to 98%, with a false negative rate of 18-67% (71). Nishigori et al reported in 21 patients undergoing laparoscopic resection for colorectal cancer with intraoperative immunofluorescence, the lymph node mapping demonstrated 23.5% of patients required modifications in the extent of lymphadenectomy and 16.7% required a change in the plan of the intestinal resection (68). A recent systematic review and meta-analysis review was performed to
determine the overall sensitivity and specificity of ICG NIR fluorescence in sentinel lymph node detection in colorectal cancer. For 12 studies across 248 patients, the authors reported accuracy rate of 75.7% and pooled sensitivity and specificity rates of 71% and 84.6%, respectively (72). With further validation and accuracy, ICG immunofluorescence lymph node mapping could become a valuable tool to guide tailored, oncologic colorectal resections. However, further controlled trials are needed studies to determine the true impact and role on outcomes, especially in patients that have received neoadjuvant therapy.

Detection of Colorectal Peritoneal Carcinomatosis

In carcinomatosis with colorectal cancer, staging and the completeness of the cytoreductive surgery are important prognostic factors. (73) Fluorescence imaging may be a tool that facilitates intraoperative assessment of tumoral margins beyond the present methods of palpation and visual inspection (74). In a proof of concept study, Filippello et al showed ICG accumulation into the tumor mass of peritoneal carcinomatosis from colorectal cancer was due to the enhanced permeability and retention, which could be augmented with bevacizumab to inhibits neoangiogenesis (74). Barabino et al perfomed a pilot study of 10 patients to validate if ICG could to detect colorectal carcinomatosis in patients scheduled for cytoreductive surgery (73). After receiving 0.25 mg/kg of ICG intravenously 24 hours before surgery, 42/58 cancerous lesions were correctly identified (72.4%). However, 12/30 non-cancerous lesions were also identified, showing the test is sensitive but not specific. To improve the accuracy, Liberale et al looked at tumor-to-background ratios in suspected lesions in patients with peritoneal metastases from colorectal cancer.
undergoing cytoreductive surgery. Patients received 0.25mg/kg intravenous intraoperatively, then had a tumor-to-background ratio calculated for all resected lesions, finding a significant difference between malignant and benign nodules. In 29%, this information modified the surgical decision-making (75). While promising, further work is needed to improve the accuracy of this application.

ICG to Guide Safe Transanal Total Mesorectal Excision (TaTME)

A new procedure gaining popularity for TME dissection is to perform the procedure via a combined laparoscopic abdominal and transanal approach: transanal total mesorectal excision (TaTME) (76, 77). This new technique requires specific training and distinct pattern recognition from the abdominal approach (78). Consequently, there is potential to easily enter an incorrect plane, resulting in bleeding, autonomic nerve injury, and urethral injury – usually of the membranous portion from inadvertent mobilization of the prostate (79, 80). FI with ICG has been described to facilitate identifying the transanal TME dissection plane, which could potentially help avoid injuries, including to the urethra. Dapri et al demonstrated the intraluminal dissection plane NIR during a single TaTME case, helping to identify the presacral fascia and correct plane of dissection (81). In a cadaver proof of concept study, Barnes et al demonstrated the feasibility, with fluorescence to visualize the urethra specifically, demonstrating the urethra in eight of eight cadavers (82). Further application of this tool and integration into TaTME training
models could improve the safety and complication profile for this new procedure, especially during the learning curve.

ICG to Confirm Traditional Anatomic Perfusion Models

ICG has also been used to verify blood flow (or lack thereof) in cases of ischemia, and to demonstrate the watershed areas and marginal arteries. Nowak et al used fluorescence angiography in 4 emergent cases of mesenteric ischemia, where recoverable, perfused regions of colon were visualized, and the length of bowel resected was significantly changed in 1 of the 4 cases (83). Watanabe et al used ICG to evaluate perfusion in patients who underwent left-sided and anterior resections with the blood flow through the last sigmoid arterial branch interrupted, leaving the rectosigmoid junction supplied by only the marginal artery (15). They found only 57.1% had a good anastomosis of the marginal artery near the rectosigmoid junction, with the remainder having either diminished, delayed or no fluorescence. Ryu et al describe successful application of intraoperative ICG fluorescence angiography to confirm a reported watershed area in the Superior Rectal artery (84). With the new understanding of the mesentery as a continuous and contiguous structure with avascular areas between arterial trunks, this could provide useful information to guide precise anatomic resections (85, 86). In a single case, Atallah et al successfully used the technology for quantitative, real-time adequate perfusion assessment of an anorectal advancement flap (87). Future studies will no doubt expand on the use of fluorescence imaging to delineate anatomy and help guide appropriate resection.
Metastatic Colorectal Lesions to the Liver

ICG fluorescence imaging has been used in the field of hepatobiliary surgery for intraoperative identification of the bile ducts and hepatic tumors for years before its application in colorectal perfusion assessment (88–90). ICG fluorescence imaging is emerging as a navigation tool for resecting metastatic hepatic tumors in laparoscopic hepatectomy. The tool may help surgeons safely and accurately identify colorectal metastatic lesions and complete laparoscopic hepatectomies, compensating for the limitations in tactile feedback and ultrasound intraoperative of the hepatic surfaces (91–94). The accumulation of ICG in metastatic liver tumors is associated with biliary excretion disorders, which can be caused by morphological obstruction in the biliary system and/or functional decrease in biliary transport, and result in accumulation of ICG in the cancerous tissues after preoperative intravenous administration (95–98) (FIGURE 4). While the tissue permeability of the fluorescence signal is limited to 5-8mm from the hepatic surfaces, the technique highly sensitive for detecting small and grossly unidentifiable metastatic lesions in real time, enhancing the accuracy of liver segmentation for resection and operative staging. (88, 99).

The technical details regarding dose and timing of ICG administration have not yet been standardized for the intraoperative fluorescence imaging of metastatic hepatic tumors. In most cases, the ICG was administered for preoperative liver function test at a dose of 0.25-0.5 mg/kg body weight between 1 day and 2 weeks before surgery (91, 100). Since ICG-fluorescence imaging of hepatic tumors is not a cancer-specific modality, the false positive rates in the present technique are relatively high (~40%) (99). With this high
false positive rate, lesions newly detected by ICG-fluorescence imaging should be confirmed by other modalities, such as palpation or intraoperative ultrasonography, before being resected. The false-positive rate can be reduced by not administering ICG on the day before surgery, especially in patients with decreased liver function from cirrhosis or preoperative chemotherapy. With the development of cancer-specific fluorophores, the specificity of intraoperative fluorescence to identify colorectal cancerous tissues can be improved, and could potentially become standard of care (101).

CONCLUSIONS

ICG fluorescence imaging is a valuable tool that aids decision-making and potentially improves outcomes in colorectal surgery. While initially introduced as an agent to assess anastomotic perfusion, the applications of fluorescence angiography continue to evolve. The current literature demonstrates the tool is safe, feasible, and beneficial in a broad array of colorectal-related procedures. As we await prospective randomized controlled trials to define the standards for utilization, the tool continues to have a significant clinical benefit in minimally invasive colorectal surgery. **Future studies will need to address the fluorophores used in fluorescence imaging. Currently, there are only two clinically available NIR fluorophores that support image-guided surgery: ICG (described here) and methylene blue. However, neither provides optimum specificity and stability for targeted image guidance (102). Therefore, it is of paramount importance to develop targeted NIR fluorophores for unmet clinical needs. New fluorophores are in development that are water soluble, biocompatible, have**
absorption and emission maxima within the desired NIR spectra, and improved fluorescence performance (103–105). Novel agents are currently being tested in animal models, and some are used as molecular endoscopically tools, providing an enhanced, visual picture of the mucosal and molecular targets (106, 107). Fluorophores could also be used to label antibodies and other biomarkers, repurposing a therapeutic agent into a diagnostic agent (108–114). Thus, developing fluorophores with the properties absent in the currently and may contribute towards the on-going expansion of NIR-fluorescence guidance in colorectal surgery. With this vast potential for expanded use, studies will need to define the cost-effectiveness of fluorescence imaging in colorectal surgery. The cost of new technology can be prohibitive for resource-limited healthcare systems, limiting widespread application. While fluorescence imaging seems to have a cost benefit in reducing complications and guiding appropriate resections, studies on the true value, considering outcomes, costs, and the patient experience, are warranted.
Author statement form

Dr. Keller - Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

Dr. Ishizawa - Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

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Mr. Chand - Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

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FIGURE LEGEND

Figure 1: Fluorescence Angiography to Assess Anastomotic Perfusion. Using ICG fluorescence angiography demonstrates a non-perfused segment in bowel that looked grossly healthy.

Figure 2: ICG infusion in the ureter.

Figure 3: ICG demonstrating a ureteral injury.

Figure 4: Identification of colorectal liver metastasis by ICG-fluorescence imaging during laparoscopic hepatectomy. a. Fluorescence imaging delineates small subcapsular hepatic metastasis located in segment 8 (right), which is unidentified by white-light color imaging (left) or intraoperative ultrasonography; b. Hepatic transection line is set using fusion-fluorescence imaging of the tumor, which enables superimposition of fluorescence signals on color images; c. The resected specimen is cut with the use of fluorescence imaging, and 3 mm-metastatic nodule appears on a cut surface of the specimen (left, arrow). The tumor shows rim-type fluorescence signals (right).
REFERENCES


82. Barnes TG, Penna M, Hompes R, Cunningham C. Fluorescence to highlight the urethra: a human cadaveric study. Tech Coloproctol. 2017


99. Abo T, Nanashima A, Tobinaga S et al. Usefulness of intraoperative diagnosis of


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ABSTRACT
Indocyanine green (ICG) fluorescence imaging is a surgical tool with increasing applications in colorectal surgery. This tool has received acceptance in various surgical disciplines as a potential method to enhance surgical field visualization, improve lymph node retrieval, and decrease anastomotic leaks. In colorectal surgery specifically, small studies have shown intraoperative fluorescence imaging is a safe and feasible method to assess anastomotic perfusion, and its use may impact anastomotic leak rates. Controlled trials are underway to validate these conclusions. In the meantime, new indications for ICG continue to develop, including innovative options for detecting and guiding management of colorectal metastasis to the liver. These advances could offer great value for surgeons and patients, by improving the accuracy and outcomes of oncologic resections. The purpose of this article is to review the literature on the current state of immunofluorescence in colon and rectal surgery and emerging applications.

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Immunofluorescence, indocyanine green (ICG); fluorescence angiography; fluorescence imaging; colorectal surgery; colorectal cancer
Introduction to Fluorescence Imaging

Fluorescence imaging (FI) is emerging as a major contributor to intraoperative decision making during surgical procedures. With FI, the tissue of interest is illuminated with light at an excitation wavelength (750 -800 nm) while observing it emit fluorescence at longer emission wavelengths (over 800 nm) (1). Indocyanine green (ICG) is the fluorophore most commonly used in fluorescence imaging. ICG is a water-soluble, tricarbocyanine dye that binds to blood lipoproteins after intravenous injection and remains confined in the intravascular compartment until elimination. It is selectively taken up at the first pass by hepatocytes and excreted unchanged into the bile. This fluorophore has tissue penetration up to 5mm, a plasma half-life of 3–5 min, with biliary excretion in 15–20 min, making it ideal for repeated applications (2). ICG has been used in other clinical applications, such as determining cardiac output, hepatic function, liver blood flow, and ophthalmic angiography for several decades (1, 3, 4). Fluorescence imaging with the application of ICG is an area of new development, and the uses continue to grow in colorectal surgery. The long-standing safety and efficacy of ICG greatly facilitates its introduction to new applications in this field (1). To date, the most publicized application in colorectal surgery has been as fluorescence angiography for perfusion analysis of the colorectal anastomosis (5). However, the indications continue to expand, and these advances could offer great value for surgeons and patients, by improving the visualization, accuracy and outcomes of colorectal resections. The purpose of this article is to review the literature on the current state of immunofluorescence in colon and rectal surgery and emerging applications.
Search strategy and selection criteria

For this narrative review, a team of surgeons that are subject matter experts in fluorescence angiography for gastrointestinal and hepatic surgery met to determine the clinically relevant areas and applications of fluorescence imaging with ICG in colorectal surgery. The consensus was: review of the available equipment, fluorescence angiography for anastomotic assessment, ureter visualization, endoscopic tattooing of colorectal lesions, lymphatic and sentinel lymph node mapping, and colorectal liver metastasis. Within each defined area, searches using the PubMed and Medline electronic databases were performed from database inception to May 2017 for original articles on the use if ICG in the setting of clinical studies in colorectal surgery. The following search terms were used: “fluorescence imaging”, “fluorescence angiography”, “Indocyanine green”, and “ICG” with “colorectal”, “colorectal surgery”, “perfusion”, “lymph node”, “sentinel lymph node”, “colorectal cancer”, “anastomotic leak”, “ureter” “hepatocellular cancer”, and “liver metastasis”, and “fluorophores” AND “imaging” AND “surgery”. Reference lists and specific authors who the team recognized as other subject matter experts in the field were also hand searched. Articles were included if published in English and full text was available. Conference proceedings and videos were not included. Abstracts were initially independently reviewed by 1 author (DK) for relevance to the defined sections and novelty. Then, full text for the selected articles was reviewed by all authors, minus the section on metastatic liver lesions, which was performed by TI. The content was written in a narrative format for full breadth of the details on the technical aspects, current state, and future application of the field.

Fluorescence Angiography: The Technical Details
There are a few options for fluorescent angiography systems for laparoscopic and open surgery, including the Stryker 1588 AIM Platform (Portage, Michigan, USA), PINPOINT™ (Novadaq, Mississauga, Ontario, Canada), the D-Light NIR/ICG™ (Karl Storz, Tuttingen, Germany), IC-View® (Pulsion Medical Systems, Munich, Germany), PDE-neo System™ (Hamamatsu Photonics K.K., Hamamatsu, Japan), the SPY Elite™ Kit (LifeCell Corporation, Bridgewater, New Jersey, USA), and da Vinci robotic system-Firefly™ (Intuitive Surgical Inc., Sunnyvale, California, USA). These systems function as a conventional laparoscope in white light mode, but can be activated into NIR mode, where the ICG is visualized as white fluorescence on a black background. The Novadaq system offers an additional PINPOINT dual display mode, where the green fluorescence is superimposed over the white-light image, providing a highlighted view of the tissue of interest (6). Surgeons should understand that the signal detectability in fluorescence imaging differs greatly among laparoscopic imaging systems and, in general, is inferior to open imaging systems (7). Studies have demonstrated the feasibility and safety of intraoperative fluorescence angiography using these systems, with no adverse effects related to the infusion of ICG (2, 5, 8–18).

**Fluorescence Angiography to Assess Anastomotic Perfusion**

Anastomotic leaks are a dreaded complication in gastrointestinal surgery. Despite technical advances in colorectal surgery, the rate of anastomotic leaks has been steady, reported in 3%-20% of colorectal cases (19, 20). The etiology of anastomotic leaks is multifactorial, and their impact is widespread- adding a significant clinical and economic
burden to the patient and healthcare system, as well as a predisposition to local cancer recurrence (20–26). The current diagnostic tests— including intraoperative endoscopic assessment, air leak testing, assessment of anastomotic donuts, measurement of local tissue oxygenation, CT scan, and water-soluble contrast enema—often fail to establish an anastomotic leak at stage early enough to allow timely intervention and minimize morbidity and mortality (24). And while a proximal diverting stoma can minimize the consequences of a leak, it does not reduce the risk (27). Thus, further study and new technology are warranted to address the issue of anastomotic leaks.

Perfusion is vital for healing, and inadequate blood flow can result in failure of anastomotic healing and leak (28). Adequate perfusion of the anastomosis is commonly confirmed by assessing palpable pulses in the mesentery, lack of bowel discoloration, and pulsatile bleeding from its cut ends; however, this method of assessment can be subjective and difficult to quantify.

A valuable tool to visualize perfusion of the bowel anastomosis is fluorescence angiography. Fluorescence angiography can help confirm anastomotic perfusion by visualizing the bowel perfusion intraoperatively in real-time. For detection of blood flow, 2–3 mL of ICG solution (2.5 mg/mL) is injected intravenously during the operation by the anesthetist. After intravenous injection, ICG is visualized as green when excited by light in the near-infrared (NIR) spectrum with a NIR camera in 30-60 seconds (29). This assessment allows the surgeon to confirm or revise a proposed resection margin after
ligation of the vascular pedicle and before the anastomosis is created (5, 8). This interrogation of perfusion before the anastomosis is especially relevant in non-anatomic resections, where aberrant or altered vascular anatomy can impair perfusion to the remaining colon (FIGURE 1). In addition, the surgeon can verify perfusion of the completed anastomosis with an endoscopic mucosal angiography evaluation of the join (9).

There is a growing body of literature supporting intraoperative assessment of perfusion impacting intraoperative management and patient outcomes, including anastomotic leak rate in colorectal surgery. Many studies evaluating immunofluorescence in colorectal anastomosis are retrospective case series in both laparoscopic and robotic colorectal resections (2, 5, 8, 10–18). These studies describe the technique as simple, taking approximately 5 minutes to perform whether laparoscopic or robotic (2, 18), and providing useful intraoperative information about the vascular perfusion during minimally invasive colorectal surgery, with promise to reduce anastomotic leak rates. Boni et al reported results of 107 laparoscopic colorectal resections (40 right colectomies, 10 splenic flexure segmental resections, 35 left colectomies, and 22 anterior resections), where fluorescence demonstrated an insufficiently perfused bowel margin in 4/107 patients, which was revised before anastomosis; none of these patients had a clinical leak (10). Gröne et al reported outcomes for consecutive rectal cancer patients who underwent laparoscopic anterior and lower anterior resection with fluorescence angiography over a 5-months period, finding perfusion imaging influenced surgical decision making in 28% of the patients; there was 1 anastomotic leak (6%, coloanal anastomosis) in all patients
during that period (11). Hellan et al. reported the outcome of perfusion assessment in 40 patients having robotic left-sided colon or rectal resection, where angiography led to a change in the proposed transection line in 40% of cases; anastomotic leak occurred in 2 patients whose site of transection had been revised (days 15 and 40) (18). Jafari et al evaluated 16 robotic low anterior resections with ICG perfusion, where the use of fluorescence angiography resulted in revision of the proximal transection point before formation of the anastomosis in 19% of patients; only 1 anastomotic leak occurred in the fluorescence angiography group compared to 4 in a matched control group (8). Ris et al reported outcomes in 30 consecutive elective minimally invasive colorectal resections (24 left-sided and 6 right-sided resections), where in addition to achieving no postoperative anastomotic leaks, the authors reported that visualizing the perfusion also encouraged avoidance of defunctioning stomas in three patients with low anastomoses (2).

With increasing utilization of the tool, the quality and volume of the evidence for fluorescence angiography in anastomotic assessment continues to grow. Boni et al completed a case-matched study, comparing 42 fluorescent angiography patients to 38 historic controls undergoing laparoscopic low anterior resection. From fluorescent angiography-demonstrated hypoperfusion, the planned anastomotic level was revised in 4.7%. There were 2 anastomotic leaks in the control group, and none in the fluorescent angiography group (30). While the sample size is too small to power conclusions, they felt it could lead to a reduction in the anastomotic leakage after total mesorectal excisions. Kudszus et al performed a case-matched study in 402 colorectal cancer patients, with a subgroup analysis by elective/emergent case status and age, for the rate
of anastomotic revision for leakage. They found an overall reduction in the absolute revision rate of the anastomosis of 4% in with fluorescence angiography, with significantly reduced rate of revision in the subgroup analysis of patients undergoing elective resections (3.1% vs. 7.7%, p = 0.04, risk of revision reduced by 60%) and in patients older than 70 years (4.3% vs. 11.9%; p = 0.04, risk of revision reduced by 64%), supporting that ICG fluorescence may significantly reduce the rate of severe complications in colorectal surgery (31). Kin et al also performed a case-matched study in colectomy and proctectomy patient, but concluded that the technology was not associated with a reduced colorectal anastomotic leak (32). Five percent of patients in the fluorescent angiography group had a revision of the planned resection margin. In the 173 pairs, they found 7.5% of the fluorescent angiography group developed an anastomotic leak versus 6.4% of controls. Multivariate analysis demonstrated that level of anastomosis and surgeon was associated with leaks (32). There were several issues with this study’s design that could account for the divergent results found in this work compared to other studies. The study was retrospective, with a historic control group used for matching. There was also a small overall sample size, and even smaller number of anastomotic leaks, (n=24), resulting in an underpowered analysis. With the lack of controlled studies and conflicting data, additional studies are necessary to determine whether this technology is beneficial for colorectal surgery. A recent systematic review of 10 cohort studies (916 patient) reported that intraoperative fluorescent angiography was associated with a reduced risk of anastomotic leakage (n = 23/693; 3.3 % (95 % CI 1.97-4.63 %) compared with no fluorescent angiography assessment (n = 19/223; 8.5 %; 95 % CI 4.8-12.2 %) (33). In the PILLAR-II prospective, multicenter, open-label,
clinical trial performed at 11 centers across the United States, 139 patients undergoing left-sided colectomy and anterior resection had perfusion assessment (5). Fluorescence angiography changed the operative plan in 8% of patients (n=11). The overall anastomotic leak rate was 1.4%, and there were no anastomotic leaks in the 11 patients who had a change in operative plan. While the existing body of literature is promising, we await randomized data to define the role for fluorescence angiography in routine clinical practice. The PILLAR III multi-center randomized controlled trial is currently underway, and we anxiously await the results (ClinicalTrials.gov identifier: NCT02205307) (34).

One limitation that deserves mention in the currently available NIR imaging systems is the subjective evaluation of fluorescence intensity and lack of quantitative data to measure tissue perfusion- leaving surgeons with the question, “How green is green?” Nerup et al aimed to establish a method to quantify perfusion in ICG fluorescence angiography in an animal model. After marking regions of interest in the stomach, the authors administered ICG and neutron-activated microspheres, performed NIR fluorescence, and then sampled the marked tissue to calculate the regional blood flow (35). They found a strong correlation between regional blood flow and the slope of the fluorescence curves, with acceptable correlation to predictive algorithms they developed. While the models show quantification of perfusion is feasible, the clinical applications of this theoretical work in animal models is pending (35). In 2 separate single-institution studies in human subjects, the absolute perfusion units were reported as a recommendation of when to resect; the threshold value for absolute perfusion units in
these reports were 18-20 (14, 36). Wada et al aimed to quantify assessment of intestinal perfusion by reviewing recorded video images and creating a time–fluorescence intensity curve at the point of bowel transection (37). They found that the fluorescence difference between maximum and baseline was the most indicative factor for anastomotic leak, with a sensitivity of 100% and a specificity of 92.5%. Further, no correlation between anastomotic leak and the time from ICG injection to the first visible fluorescence signal was found (37). While these studies are promising, controlled trials with larger sample sizes are needed to standardize and validate the quantitative models. In the meantime, the adequacy of anastomotic perfusion remains subjective.

**Emerging Uses of Fluorescence Angiography in Colorectal Surgery**

While developed to assess perfusion, various other uses of fluorescence angiography are being reported, which compliment the standard of care in colorectal surgery, and could facilitate improving patient outcomes.

**Ureter Visualization**

Iatrogenic ureteral injury is a relatively rare but devastating complication in colorectal surgery, with significant morbidity (38, 39). While rare overall, a higher incidence is reported with laparoscopic colorectal surgery, especially in rectal cancer resections (40, 41). Preoperative ureteral stenting is an option, but it has not been shown to decrease the risk of ureteral injury- only helping with early identification of ureteral injuries (42). In addition, there are complications and costs associated with placing stents, including increased operative time and an increase in length of stay (42, 43). ICG fluorescence of
the ureters has value for facilitating ureter identification, especially in minimally invasive colorectal surgery (FIGURE 2). The ICG binds proteins on urothelial layer on the ureter, reversibly staining the inner lining of the ureter, and allowing visualization of the green-illuminated ureter under NIR fluorescence (44). As ICG fluorescence penetrates tissue, the intraureteral ICG may be used to localize the entire course of the ureters, even when obscured in fibrotic tissue. While the intraureteral ICG cannot detect thermal or devascularization injuries, full-thickness ureteral injuries can be identified cause readily apparent leakage of ICG into the surgical field (FIGURE 3). Studies have described the technique, feasibility, and utility for visual identification of the ureters during laparoscopic (45–47) and robotic surgery using the Firefly® system (44, 48–50). The protocols described between 0.125-1mg/kg of methylene blue (45, 47) and 10-25mg of ICG dissolved in 10-mL of sterile water and injected through the open catheter, either as a bolus or through slow infusion over 5 minutes (44, 48–50). Maximum fluorescence was reported between 9 and 20 minutes from administration (45). The reported results describe bilateral ureters fluorescing green in 50-91% when methylene blue was used, and 100% of patients with ICG (44, 45, 47–50). The fluorescence persisted through the duration of the operative procedure (mean operative times 121-171.3 minutes), despite some variation in brightness from the depth of the ureter to the peritoneal surface across patients (44, 48–50). No intraoperative or postoperative adverse effects attributable to fluorophore administration were reported (44, 47–49). Further study will determine the role for this useful adjunct to prevent iatrogenic ureteral injury during pelvic dissection in colorectal surgery.
Endoscopic Tattooing Of Colorectal Lesions

Preoperative endoscopic tattooing of colorectal neoplasms facilitates localization at the time of surgery, and is crucial for appropriate resection and treatment planning (51). Tattooing has become especially important in the era of minimally invasive surgery, with the lack of tactile feedback afforded during resection (52). The correct measurement and appropriate margins facilitated by precise localization could directly impact recurrence and survival. India ink has been the standard marking agent, but complications have been reported, including perforation, peritonitis, abscesses, inflammatory pseudotumors, and adhesion-related ileus (53–55). ICG has been reported as the endoscopic tattoo agent in several studies to guide colorectal resections (56–61). In these reports, peritumoral injection before open or minimally invasive resection using NIR fluorescence to identify the lesion has been reported to be safe, feasible, and effective. No adverse effects have been reported and all colorectal lesions were visualized upon open or laparoscopic resection. The median time for injection of ICG has been reported as 4 days prior to resection, but the ICG has been described as visible up to 7 days after endoscopic injection; after 9-10 days, the agent was visible in only 20% of patients (58, 60). A comparative study was performed by Nagata et al, who endoscopically injected 24 patients with both ICG and India ink injected at separate sites within 4 days of laparoscopic colorectal surgery (59). They found the NIR fluorescence showed tumor location clearly and accurately in 100% of patients, while 10 of 24 patients had negative staining of India ink. The site of ICG injection did not reveal fibrosis, inflammation, necrosis or microabscesses, while the India Ink injection sites had vasculitis, necrosis, edema, and neutrophilic infiltration of the bowel wall. While large scale, controlled trails
are needed to define the site, concentration, and timing, from the benefits reported, ICG has value as a preoperative marking agent to identify tumor location.

**Lymphatic Mapping and Sentinel Lymph Node Identification**

In oncologic resections, sentinel lymph node mapping allows targeted identification and harvesting of potentially metastatic lymph nodes (62). Taking a clue from breast and gynecologic malignancies (63, 64), ICG is emerging as a powerful tool for lymphatic mapping, identification of micrometastatic disease, and focused target nodal assessment in colorectal cancer. There is the potential that the NIR information could change the operative course and recommendations for adjuvant therapy postoperatively. A few studies have reported the ICG immunofluorescence provided valuable information for finding lymph nodes; if these nodes are present outside of the traditional resection field, the operative strategy could be changed from this information (61, 65–68). It was noted that ICG may be less reliable due to neoadjuvant therapy in rectal cancers (61).

Near-infrared fluorescence mapping of the pelvic side-wall in low rectal cancer can also guide the need for extended lymphadenectomy in appropriate patients, and spare patients with tumor-negative sentinel nodes from the morbidity of a lateral pelvic sidewall node dissection (67, 69). Intraoperative lymphatic mapping may also help define the lymphatic course for tumors that have variable drainage or in reoperative cases, where lymphatic-bearing tissue has been excised (56). The sentinel lymph node biopsy could have specific utility in transanal resections of rectal cancer, where identification could change the treatment course to a more radical resection. Arezzo et al performed a submucosal ICG injection in 3 patients, then used NIR fluorescence to map the mesorectal lymphatics and guide full-thickness resection with any involved lymph nodes (70). In all 3 patients, the
pathologist confirmed presence of excised nodes, lack of metastasis, and the patients were spared radical surgery. Published studies have described the accuracy and benefit with this tool. Cahill et al used ICG with intraoperative fluorescence for real-time identification of the lymphatic drainage and sentinel mesocolic lymph nodes in 18 patients undergoing laparoscopic surgery for colorectal cancer (56). They found the fluorescence helped identify the sentinel nodes - whether within the specimen or outside of the standard resection field - prior to formal dissection, and was valid to guide their resection. In their series, four of 18 patients had mesocolic sentinel lymph nodes outside of the traditional resection field identified with ICG immunofluorescence. Hirche et al evaluated 26 patients with colon cancer with ICG, then visualized their lymphatic mapping and sentinel lymph nodes with immunofluorescence, comparing results to the histopathology for clinical feasibility, detection rate, and sensitivity (66). The found the ICG could have improved accuracy over conventional methods. Fluorescence imaging identified the sentinel lymph node in 25/26 patients (detection rate, 96%); metastatic involvement was found in 9/11 nodal positive patients by conventional histopathology, a sensitivity of 82% (66). To date, the rate of lymph node detection has been described from 89% to 98%, with a false negative rate of 18-67% (71). Nishigori et al reported in 21 patients undergoing laparoscopic resection for colorectal cancer with intraoperative immunofluorescence, the lymph node mapping demonstrated 23.5% of patients required modifications in the extent of lymphadenectomy and 16.7% required a change in the plan of the intestinal resection (68). A recent systematic review and meta-analysis review was performed to determine the overall sensitivity and specificity of ICG NIR fluorescence in sentinel lymph node detection in colorectal cancer. For 12 studies across
248 patients, the authors reported accuracy rate of 75.7% and pooled sensitivity and specificity rates of 71% and 84.6%, respectively (72). With further validation and accuracy, ICG immunofluorescence lymph node mapping could become a valuable tool to guide tailored, oncologic colorectal resections. However, further controlled trials are needed studies to determine the true impact and role on outcomes, especially in patients that have received neoadjuvant therapy.

Detection of Colorectal Peritoneal Carcinomatosis

In carcinomatosis with colorectal cancer, staging and the completeness of the cytoreductive surgery are important prognostic factors. (73) Fluorescence imaging may be a tool that facilitates intraoperative assessment of tumoral margins beyond the present methods of palpation and visual inspection (74). In a proof of concept study, Filippello et al showed ICG accumulation into the tumor mass of peritoneal carcinomatosis from colorectal cancer was due to the enhanced permeability and retention, which could be augmented with bevacizumab to inhibits neoangiogenesis (74). Barabino et al perfomed a pilot study of 10 patients to validate if ICG could to detect colorectal carcinomatosis in patients scheduled for cytoreductive surgery (73). After receiving 0.25 mg/kg of ICG intravenously 24 hours before surgery, 42/58 cancerous lesions were correctly identified (72.4%). However, 12/30 non-cancerous lesions were also identified, showing the test is sensitive but not specific. To improve the accuracy, Liberale et al looked at tumor-to-background ratios in suspected lesions in patients with peritoneal metastases from colorectal cancer undergoing cytoreductive surgery. Patients received 0.25mg/kg intravenous intraoperatively, then had a tumor-to-background ratio calculated for all resected lesions, finding a significant difference between malignant and benign nodules.
In 29%, this information modified the surgical decision-making (75). While promising, further work is needed to improve the accuracy of this application.

**ICG to Guide Safe Transanal Total Mesorectal Excision (TaTME)**

A new procedure gaining popularity for TME dissection is to perform the procedure via a combined laparoscopic abdominal and transanal approach: transanal total mesorectal excision (TaTME) (76, 77). This new technique requires specific training and distinct pattern recognition from the abdominal approach (78). Consequently, there is potential to easily enter an incorrect plane, resulting in bleeding, autonomic nerve injury, and urethral injury – usually of the membranous portion from inadvertent mobilization of the prostate (79, 80). FI with ICG has been described to facilitate identifying the transanal TME dissection plane, which could potentially help avoid injuries, including to the urethra. Dapri et al demonstrated the intraluminal dissection plane NIR during a single TaTME case, helping to identify the presacral fascia and correct plane of dissection (81). In a cadaver proof of concept study, Barnes et al demonstrated the feasibility, with fluorescence to visualize the urethra specifically, demonstrating the urethra in eight of eight cadavers (82). Further application of this tool and integration into TaTME training models could improve the safety and complication profile for this new procedure, especially during the learning curve.

**ICG to Confirm Traditional Anatomic Perfusion Models**
ICG has also been used to verify blood flow (or lack thereof) in cases of ischemia, and to demonstrate the watershed areas and marginal arteries. Nowak et al used fluorescence angiography in 4 emergent cases of mesenteric ischemia, where recoverable, perfused regions of colon were visualized, and the length of bowel resected was significantly changed in 1 of the 4 cases (83). Watanabe et al used ICG to evaluate perfusion in patients who underwent left-sided and anterior resections with the blood flow through the last sigmoid arterial branch interrupted, leaving the rectosigmoid junction supplied by only the marginal artery (15). They found only 57.1% had a good anastomosis of the marginal artery near the rectosigmoid junction, with the remainder having either diminished, delayed or no fluorescence. Ryu et al describe successful application of intraoperative ICG fluorescence angiography to confirm a reported watershed area in the Superior Rectal artery (84). With the new understanding of the mesentery as a continuous and contiguous structure with avascular areas between arterial trunks, this could provide useful information to guide precise anatomic resections (85, 86). In a single case, Atallah et al successfully used the technology for quantitative, real-time adequate perfusion assessment of an anorectal advancement flap (87). Future studies will no doubt expand on the use of fluorescence imaging to delineate anatomy and help guide appropriate resection.

Metastatic Colorectal Lesions to the Liver

ICG fluorescence imaging has been used in the field of hepatobiliary surgery for intraoperative identification of the bile ducts and hepatic tumors for years before its
application in colorectal perfusion assessment (88–90). ICG fluorescence imaging is emerging as a navigation tool for resecting metastatic hepatic tumors in laparoscopic hepatectomy. The tool may help surgeons safely and accurately identify colorectal metastatic lesions and complete laparoscopic hepatectomies, compensating for the limitations in tactile feedback and ultrasound intraoperative of the hepatic surfaces (91–94). The accumulation of ICG in metastatic liver tumors is associated with biliary excretion disorders, which can be caused by morphological obstruction in the biliary system and/or functional decrease in biliary transport, and result in accumulation of ICG in the cancerous tissues after preoperative intravenous administration (95–98) (FIGURE 4). While the tissue permeability of the fluorescence signal is limited to 5-8mm from the hepatic surfaces, the technique highly sensitive for detecting small and grossly unidentifiable metastatic lesions in real time, enhancing the accuracy of liver segmentation for resection and operative staging. (88, 99). The technical details regarding dose and timing of ICG administration have not yet been standardized for the intraoperative fluorescence imaging of metastatic hepatic tumors. In most cases, the ICG was administered for preoperative liver function test at a dose of 0.25-0.5 mg/kg body weight between 1 day and 2 weeks before surgery (91, 100). Since ICG-fluorescence imaging of hepatic tumors is not a cancer-specific modality, the false positive rates in the present technique are relatively high (~40%) (99). With this high false positive rate, lesions newly detected by ICG-fluorescence imaging should be confirmed by other modalities, such as palpation or intraoperative ultrasonography, before being resected. The false-positive rate can be reduced by not administering ICG on the day before surgery, especially in patients with decreased liver function from cirrhosis or preoperative
chemotherapy. With the development of cancer-specific fluorophores, the specificity of intraoperative fluorescence to identify colorectal cancerous tissues can be improved, and could potentially become standard of care (101).

CONCLUSIONS

ICG fluorescence imaging is a valuable tool that aids decision-making and potentially improves outcomes in colorectal surgery. While initially introduced as an agent to assess anastomotic perfusion, the applications of fluorescence angiography continue to evolve. The current literature demonstrates the tool is safe, feasible, and beneficial in a broad array of colorectal-related procedures. As we await prospective randomized controlled trials to define the standards for utilization, the tool continues to have a significant clinical benefit in minimally invasive colorectal surgery. Future studies will need to address the fluorophores used in fluorescence imaging. Currently, there are only two clinically available NIR fluorophores that support image-guided surgery: ICG (described here) and methylene blue. However, neither provides optimum specificity and stability for targeted image guidance (102). Therefore, it is of paramount importance to develop targeted NIR fluorophores for unmet clinical needs. New fluorophores are in development that are water soluble, biocompatible, have absorption and emission maxima within the desired NIR spectra, and improved fluorescence performance (103–105). Novel agents are currently being tested in animal models, and some are used as molecular endoscopically tools, providing an enhanced, visual picture of the mucosal and molecular targets (106, 107). Fluorophores could also be used to label antibodies and
other biomarkers, repurposing a therapeutic agent into a diagnostic agent (108–114). Thus, developing fluorophores with the properties absent in the currently and may contribute towards the on-going expansion of NIR-fluorescence guidance in colorectal surgery. With this vast potential for expanded use, studies will need to define the cost-effectiveness of fluorescence imaging in colorectal surgery. The cost of new technology can be prohibitive for resource-limited healthcare systems, limiting widespread application. While fluorescence imaging seems to have a cost benefit in reducing complications and guiding appropriate resections, studies on the true value, considering outcomes, costs, and the patient experience, are warranted.
Author statement form

Dr. Keller- Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

Dr. Ishizawa- Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

Mr. Cohen- Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

Mr. Chand- Contributed to study design, data collection and synthesis of information used, drafting the manuscript, critical revisions, selecting and approving figures, final approval of the submitted work

Declaration of interests and source of funding statements

The authors have no relevant conflicts of interest

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FIGURE LEGEND

Figure 1: Fluorescence Angiography to Assess Anastomotic Perfusion. Using ICG fluorescence angiography demonstrates a non-perfused segment in bowel that looked grossly healthy.

Figure 2: ICG infusion in the ureter

Figure 3: ICG demonstrating a ureteral injury

Figure 4: Identification of colorectal liver metastasis by ICG-fluorescence imaging during laparoscopic hepatectomy. a. Fluorescence imaging delineates small subcapsular hepatic metastasis located in segment 8 (right), which is unidentified by white-light color imaging (left) or intraoperative ultrasonography; b. Hepatic transection line is set using fusion-fluorescence imaging of the tumor, which enables superimposition of fluorescence signals on color images; c. The resected specimen is cut with the use of fluorescence imaging, and 3 mm-metastatic nodule appears on a cut surface of the specimen (left, arrow). The tumor shows rim-type fluorescence signals (right).
REFERENCES


82. Barnes TG, Penna M, Hompes R, Cunningham C. Fluorescence to highlight the urethra: a human cadaveric study. Tech Coloproctol. 2017
99. Abo T, Nanashima A, Tobinaga S et al. Usefulness of intraoperative diagnosis of


Reviewers' comments:

Reviewer #1:

Introduction
1. The introduction essentially revolves around anastomotic leakage which gives an impression that the article is basically discussing this problem whereas the article is about ICG fluorescence and its uses in colorectal surgery which include, but not limited to, perfusion assessment. I propose to start the introduction with a background on ICG itself, its structure, development, uses in other medical fields then gradually proceed to discuss its primary indication in colorectal surgery highlighting anastomotic leakage in this context.

Per the reviewer’s preference, the introduction and initial sections have been completely restructured, and these focused, additional details have been added into the text.

2. First paragraph: "the current diagnostic tests often fail to…” it would be useful to enumerate these current diagnostic tests to the reader.

Per the reviewer’s request, the details of intraoperative endoscopic assessment, air leak testing, assessment of anastomotic donuts, measurement of local tissue oxygenation, CT scan, and water-soluble contrast enema have been added into the text.

3. Second paragraph: "Adequate perfusion of the anastomosis is commonly confirmed by the assessing palpable pulses in the mesentery", I assume it would read as "commonly confirmed by assessing palpable pulses in the mesentery"

This has been corrected.

Technical details
4. Please clarify the meaning of the abbreviation "ICG" at the first appearance in the text as you did in the abstract.

This has been expanded.

5. Last sentence reads "Studies have demonstrated the feasibility and safety of intraoperative fluorescence angiography using these systems, with no adverse effects related to the infusion of ICG", however none of these studies were cited to support this sentence.

Per the reviewer’s request, references were added here.

Fluorescence Angiography to Assess Anastomotic Perfusion

6. Page 7, line 4: the felt or they felt?

This has been corrected.

7. While all studies reported that ICG fluorescence effectively managed to reduce anastomotic leakage; Kim et al (30) were the only authors who concluded that this technology was not associated with a reduced leak rate, is there something unique with this particular study regarding patients' characteristics, indications for or type of surgery
performed that led to this contradiction?
The article by Kin et al was divergent from the rest of the literature. There were several issues with this study’s design that could account for the divergent results found in this work compared to other studies. The study was retrospective, with a historic control group used for matching. There was also a small overall sample size, and even smaller number of anastomotic leaks, (n=24), resulting in an underpowered analysis. This has been added to the text.

8. Since the PILLAR II trial is the only RCT included, it would be helpful to extend the discussion on its outcome. Also, the registered protocol of the PILLAR III trial could be cited as well in the text. PILLAR II was a prospective trial, but not RCT. Additional details of the PILLAR II outcomes are under the section for “Fluorescence Angiography to Assess Anastomotic Perfusion”. This has been highlighted for the reviewer’s easy review. PILLAR III is an RCT. The registered protocol for PILLAR III was added, as requested.

9. The subjective assessment of fluorescence intensity is indeed a shortcoming, a recent study employed quantitative assessment of fluorescence angiography in animal model (N Nerup et al. Langenbecks Arch Surg 402 (4), 655-662) and concluded that quantitative assessment is not only feasible but also easy to perform, perhaps the authors may like to refer to this trial in their review. This animal model study has been added into the text as requested.

Ureter Visualization
10. Six studies evaluating the utility of ICG fluorescence in the visual identification of the ureters during Laparoscopic or robotic colectomy were cited, however no mention of the outcome was made. Some information needs to be displayed such as: was the ureter clearly identified in all patients? Time elapsed between ICG injection and visualization of the ureter, and the incidence of ureteral injury in each study. Details for these studies have been added to the text.

Lymphatic mapping and sentinel lymph node identification
Some recent publications on the role of ICG fluorescence in sentinel LN mapping could be added to this section as:

11. A recent meta-analysis reported the median and pooled sensitivity and specificity rates of ICG fluorescence in sentinel LN mapping in colorectal cancer (Emile SH et al, J Surg Oncol. 2017 Jun 1. doi: 10.1002/jso.24701), this meta-analysis emphasized the importance of standardizing the technique of ICG fluorescence to achieve optimal outcomes. Per request, the reviewer’s work was included in the text.

ICG to Confirm Traditional Anatomic Perfusion Models
13. I suggest to move this section to be included to the previous section discussing 
anastomotic perfusion since both sections reflect the same utility (perfusion assessment). 
We thank the author for his suggestion. This section details visualization of the ureters- 
not perfusion assessment, while the prior details anastomotic perfusion assessment.

Primary and Metastatic Liver Lesions
14. This section details the use of ICG fluorescence in hepatic malignancies overall 
which is not really pertinent to the topic of the review. Since the review is all about the 
uses of ICG in colorectal surgery, a discussion of the utility of ICG-FI in detecting 
colorectal liver metastasis would be more appropriate and relevant whilst highlighting its 
mechanism referring to its original indication in primary hepatic malignancies. 
This section has been revised and focused, as suggested.

Additional comments
15. Please add a section on other emerging clinical applications of ICG fluorescence in 
colorectal surgery that were not discussed in this review such as:

1. Detection of colorectal peritoneal carcinomatosis (Flippelo et al, Surg Innov. 2017 
2. Application of ICG-FI in transanal TME to identify the plane of dissection (Dapri 
   the urethra (Barnes et al, Tech Coloproctol. 2017 May 30. doi: 10.1007/s10151-017- 
   1615-y).
3. Assessment of perfusion of anodermal/anorectal advancement flap (Atallah et al, 
   Dis Colon Rectum. 2013 Jun;56(6):797. doi: 10.1097/DCR.0b013e31828e1b71), (Turner 
4. A revolutionary, yet still experimental, application of ICG-FI is in the assessment 
   2012;7(9):e44310. doi: 10.1371/journal.pone.0044310)

Per the reviewer’s comment, sections have been added or references incorporated into 
existing sections.

16. One of the shortcomings of ICG-fluorescence systems is their high cost that can be 
prohibitive for resource-limited communities, and hence limits its wide application. This 
should be discussed in the manuscript with emphasis on the cost-effectiveness of this 
technology as demonstrated in the literature.
This limitation has been added to the conclusions.

Reviewer #2: Although on page 4 of their manuscript the authors describe multiple 
commercially available systems for ICG imaging their illustrations focus strictly upon the 
equipment available by one manufacturer. The only way that this manuscript would be
suitable for publication is if the authors offered images from a variety of manufacturers to give a better balance. For example, figures 1 and 2 do not lend any particular importance or help clarify comprehension of the subject, but seem almost like marketing for the Stryker company. Thank you for the suggestion- the company specific images were removed.

[Editor's note: we agree, please remove the images of the specific systems from the manuscript]
This was removed- thank you!

Reviewer #3:

1. Firstly, with reference to the originality of the study, it does not stand out from other reviews published so far on the same subject. The introduction and the description of the technical details are correct, but somewhat brief. Since it is a review of the literature, I believe that it would be appropriate to explain initially all the aspects that will be analyzed in the article, and not only the suture failure, which, although important, is not the only issue at hand, since other applications and their general use in colorectal surgery are also discussed. Introduction provide a background for the paper but don´t state the specific purpose (only named it in the abstract).
This article synthesizes all current and emerging applications of ICG fluorescence in colorectal surgery imaging into a detailed narrative review, a unique addition to the literature to the best of our knowledge. The purpose was added to the introduction, with explanation that the review will describe popular current and emerging applications.

2. Then it goes on to speak directly about the uses of the ICG without a section of material and methods. A section of material and methods should be presented, explaining how the literature review was carried out, which databases were used, what type of articles they analyzed and their number, as well as inclusion and exclusion criteria. Reference to a minimum number of patients in the series collected, design of the studies analyzed, deadline for antiquity, literature only in English or not, etc. (Meta-analyses, systematic reviews, review articles, randomised controlled trials and non-randomised comparative studies including controlled clinical trials and cohort studies were included or not?). It would be useful also to know how the Quality Assessment of the included studies was performed.

We thank the author for his suggestion. A “Search Strategy and Selection Criteria” was included at the end of the paper. This was a narrative review, not a systematic review or meta-analysis, where the included studies are graded, but the methods were detailed as requested.

[Editor's note: this should be included as a Search Strategy and Selection Criteria panel at the end of the paper.] 

After explaining the context of the study, it goes on to explain one by one the uses of
ICG,

3. Fluorescence Angiography to Assess Anastomotic Perfusion
In this section an adequate summary of the current situation of the use of ICG to assess anastomotic perfusion. The studies to which they refer are current and relevant. They also refer to future studies such as PILLAR III and also talk about new methods to evaluate the use of indocyanine green objectively and not subjectively as has been done so far. We thank the author for their comment that this section is adequate. There are no specific points to address.

4. Emerging Uses of Fluorescence Angiography in Colorectal Surgery
This section discusses the use of ICG in ureter visualization, endoscopic tattooing of colorectal lesions, lymphatic mapping and sentinel lymph node identification, anatomic perfusion models and primary and metastatic liver lesions. The review of each topic is detailed, identifying in each case relevant literature on the subject, explaining sufficiently recent and innovative studies that support the objective to be addressed.
Maybe in this section could have been added a reference to the use in the future of fluorescent agents such as icg in what is believed to be going to revolutionize the role of the fluorescence imaging system in oncologic surgery, the discovery of a contrast agent that binds to tumor-specific targets. To date, new fluorophores have been developed. They are characterized by improved fluorescent properties and can be conjugated to specific ligands. These contrast agents have already been tested in animals. The next step is to transfer the achieved results to human clinical trials.
Discussion on the current and future state of fluorophores was added to the text.

[Editor's note: we have published a review on molecular imaging in the digestive tract (http://www.thelancet.com/journals/langas/article/PIIS2468-1253(16)30027-9/fulltext) and a research article on molecular fluorescence guided surgery of peritoneal metastases of colorectal origin (http://www.thelancet.com/journals/langas/article/PIIS2468-1253(16)30082-6/fulltext), which may be of use while researching this last component]
Thank you for the provided references- they were used in the revision.

5. The conclusions are short but clear, although they do not bring any novelty to highlight Additional text considering cost studies and the requested items on other fluorophores was added for novelty.

Editorial comments:
1. Please add a conflict of interest statement that matches the declarations on your author statement forms, which were submitted with your first draft. Authors should be referred to by their initials in this section. If there are none, then please state "The authors declared no conflicts of interest" or "The other authors declared no conflicts of interest". This has been added. The authors declare no conflicts of interest

2. We require confirmation that the paper has not been submitted to another journal, and has not been published in whole or in part elsewhere previously.
This paper has not been submitted to another journal, and has not been published in whole or in part elsewhere previously. A statement to this effect has been added to the cover page.

3. Please check with your co-authors, and confirm, that all names are spelt correctly, and affiliations listed correctly. We cannot guarantee that we will be able to correct names and affiliations after publication of your article.
This is correct, thank you

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5. Please supply the photographic figures as high-resolution TIF format. For more information, see download.thelancet.com/flatcontentassets/authors/artwork-guidelines.pdf.
The images have been reformatted per the guidelines.

6. Please supply a section entitled "Search strategy and selection criteria". This should state clearly the sources (databases, journals, or book reference lists, etc) of the material covered and the criteria used to include or exclude studies. Please state which search terms, languages and date ranges were used.
This has been added, thank you for allowing us to revise.

--- End of editorial comments ---
Figure 3
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